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## INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

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<b>(21) International Application Number:</b> PCT/US92/07175 <b>(22) International Filing Date:</b> 21 August 1992 (21.08.92)  <b>(30) Priority data:</b> 749,451 23 August 1991 (23.08.91) US 834,044 11 February 1992 (11.02.92) US  <b>(60) Parent Applications or Grants</b> <b>(63) Related by Continuation</b> US 07/934,161 (CIP) Filed on 21 August 1992 (21.08.92) US 07/834,044 (CIP) Filed on 11 February 1992 (11.02.92) US 07/749,451 (CIP) Filed on 23 August 1991 (23.08.91)  <b>(71) Applicant (for all designated States except US):</b> NPS PHARMACEUTICALS, INC. [US/US]; 420 Chipeta Way, Salt Lake City, UT 84108 (US).		<b>(72) Inventors; and</b> <b>(75) Inventors/Applicants (for US only) :</b> NEMETH, Edward, F. [US/US]; 3258 E. Fortuna Drive, Salt Lake City, UT 84124 (US). VAN WAGENEN, Bradford, C. [US/US]; 1070 East 300 South, #507, Salt Lake City, UT 84102 (US). BALANDRIN, Manuel, F. [US/US]; 5787 South 1585 East, Salt Lake City, UT 84121 (US).  <b>(74) Agents:</b> WARBURG, Richard, J. et al.; Lyon & Lyon, 611 West Sixth Street, 34th Floor, Los Angeles, CA 90017 (US).  <b>(81) Designated States:</b> AT, AU, BB, BG, BR, CA, CH, CS, DE, DK, ES, FI, GB, HU, JP, KP, KR, LK, LU, MG, MN, MW, NL, NO, PL, RO, RU, SD, SE, US , Euro- pean patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, SN, TD, TG).  <b>Published</b> <i>With international search report.</i>
<b>(54) Title:</b> CALCIUM RECEPTOR ACTIVE MOLECULES  <b>(57) Abstract</b>  Method and composition useful for treating a patient having a disease characterized by an abnormal level of one or more components, the activity of which is regulated or affected by activity of one or more Ca <sup>2+</sup> receptors. Novel compounds useful in these methods and compositions are also provided. The method includes administering to the patient a therapeutically effective amount of a molecule active at one or more Ca <sup>2+</sup> receptors as an agonist or antagonist. Preferably, the molecule is able to act as either a selective agonist or antagonist at a Ca <sup>2+</sup> receptor of one or more but not all cells chosen from the group consisting of parathyroid cells, bone osteoclasts, juxtaglomerular kidney cells, proximal tubule kidney cells, keratinocytes, parafollicular thyroid cells and placental trophoblasts and a pharmaceutically acceptable carrier.		

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DESCRIPTIONCalcium Receptor Active MoleculesRelated Application

This application is a continuation-in-part of Nemeth, et al., entitled Calcium Receptor Active Molecules, filed February 11, 1992, which is a continuation-in-part of  
5 Nemeth et al., entitled Calcium Receptor Activators, U.S. Serial No. 07/749,451, filed August 23, 1991, the whole of both of which, including drawings, are hereby incorporated herein by reference.

Field of the Invention

10 This invention relates to the design, development, composition and use of novel calcimimetic molecules able to act in a manner analogous to extracellular calcium ions on cells, to calcilytic molecules which block the activity of extracellular calcium ions on cells, and to methods for  
15 their use and identification.

Background of the Invention

The following description provides a summary of information relevant to the present invention. It is not an admission that any of the information provided herein  
20 is prior art to the presently claimed invention, nor that any of the publications specifically or implicitly referenced are prior art to that invention.

Certain cells in the body respond not only to chemical signals, but also to ions such as extracellular  
25 calcium ions ( $\text{Ca}^{2+}$ ). Changes in the concentration of extracellular  $\text{Ca}^{2+}$  (referred to herein as " $[\text{Ca}^{2+}]$ ") alter the functional responses of these cells. One such specialized cell is the parathyroid cell which secretes parathyroid hormone (PTH). PTH is the principal endocrine  
30 factor regulating  $\text{Ca}^{2+}$  homeostasis in the blood and extracellular fluids.



PTH, by acting on bone and kidney cells, increases the level of  $\text{Ca}^{2+}$  in the blood. This increase in  $[\text{Ca}^{2+}]$  then acts as a negative feedback signal, depressing PTH secretion. The reciprocal relationship between  $[\text{Ca}^{2+}]$  and PTH secretion forms the essential mechanism maintaining bodily  $\text{Ca}^{2+}$  homeostasis.

Extracellular  $\text{Ca}^{2+}$  acts directly on the parathyroid cell to regulate PTH secretion. The existence of a parathyroid cell surface protein which detects changes in  $[\text{Ca}^{2+}]$  has been suggested. This protein acts as a receptor for extracellular  $\text{Ca}^{2+}$  ("the  $\text{Ca}^{2+}$  receptor"), and is suggested to detect changes in  $[\text{Ca}^{2+}]$  and to initiate a functional cellular response, PTH secretion. For example, the role of  $\text{Ca}^{2+}$  receptors and extracellular  $\text{Ca}^{2+}$  in the regulation of intracellular  $\text{Ca}^{2+}$  and cell function is reviewed in Nemeth et al., 11 Cell Calcium 319, 1990; the role of  $\text{Ca}^{2+}$  receptors in parafollicular and parathyroid cells is discussed in Nemeth, 11 Cell Calcium 323, 1990; and the role of  $\text{Ca}^{2+}$  receptors on bone osteoclasts is discussed by Zaidi, 10 Bioscience Reports 493, 1990.

Other cells in the body, specifically the osteoclast in bone, the juxtaglomerular and the proximal tubule cells in the kidney, the keratinocyte in the epidermis, the parafollicular cell in the thyroid, and the trophoblast in the placenta, have the capacity to sense changes in  $[\text{Ca}^{2+}]$ . It has been suggested that cell surface  $\text{Ca}^{2+}$  receptors may also be present on these cells, imparting to them the ability to detect and to initiate or enable a response to changes in  $[\text{Ca}^{2+}]$ .

In parathyroid cells, osteoclasts, parafollicular cells (C-cells), keratinocytes, juxtaglomerular cells and trophoblasts, an increase in  $[\text{Ca}^{2+}]$  evokes an increase in intracellular free  $\text{Ca}^{2+}$  concentration (" $[\text{Ca}^{2+}]_i$ "). Such an increase may be caused by influx of extracellular  $\text{Ca}^{2+}$  or by mobilization of  $\text{Ca}^{2+}$  from intracellular organelles. Changes in  $[\text{Ca}^{2+}]_i$  are readily monitored and quantitated

using fluorimetric indicators such as fura-2 or indo-1 (Molecular Probes, Eugene, OR). Measurement of  $[Ca^{2+}]_i$  provides an assay to assess the ability of molecules to act as agonists or antagonists at the  $Ca^{2+}$  receptor.

5 In parathyroid cells, increases in the concentration of extracellular  $Ca^{2+}$  evoke rapid and transient increases in  $[Ca^{2+}]_i$  which are followed by lower yet sustained increases in  $[Ca^{2+}]_i$ . The transient increases in  $[Ca^{2+}]_i$  arise from the mobilization of intracellular  $Ca^{2+}$ , whereas  
10 the lower, sustained increases result from the influx of extracellular  $Ca^{2+}$ . The mobilization of intracellular  $Ca^{2+}$  is accompanied by increased formation of inositol-1,4,5-trisphosphate ( $IP_3$ ) and diacylglycerol, two biochemical indicators which are associated with receptor-dependent  
15 mobilization of intracellular  $Ca^{2+}$  in various other cells.

In addition to  $Ca^{2+}$ , various other di- and trivalent cations, such as  $Mg^{2+}$ ,  $Sr^{2+}$ ,  $Ba^{2+}$ ,  $La^{3+}$ , and  $Gd^{3+}$  also cause the mobilization of intracellular  $Ca^{2+}$  in parathyroid cells.  $Mg^{2+}$  and  $La^{3+}$  also increase the formation of  $IP_3$ ;  
20 all these inorganic cations depress the secretion of PTH. The postulated  $Ca^{2+}$  receptor on the parathyroid cell is therefore promiscuous because it detects a variety of extracellular di- and trivalent cations.

The ability of various compounds to mimic extracellular  $Ca^{2+}$  in vitro is discussed by Nemeth et al.,  
25 (spermine and spermidine) in "Calcium-Binding Proteins in Health and Disease", 1987, Academic Press, Inc., pp. 33-35; Brown et al., (e.g., neomycin) 128 Endocrinology 3047, 1991; Chen et al., (diltiazem and its analog, TA-3090)  
30 5 J. Bone and Mineral Res. 581, 1990; and Zaidi et al., (verapamil) 167 Biochem Biophys Res Comm 807, 1990.

Brown et al., 6 J. Bone and Mineral Res. 11, 1991 discuss the existing theories regarding the effects of  $Ca^{2+}$  ions on parathyroid cells, and propose that the results  
35 may be explained by both a receptor-like mechanism and a receptor-independent mechanism as follows:

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5 Polyvalent cations [e.g., divalent and  
trivalent cations] exert a variety of  
effects on parathyroid function, such as  
inhibition of parathyroid hormone (PTH)  
secretion and cAMP accumulation, stimula-  
tion of the accumulation of inositol phos-  
phates, and elevation of the cytosolic  
calcium concentration. These actions are  
thought to be mediated through a "receptor-  
like" mechanism. The inhibition of  
10 agonist-stimulated cAMP accumulation by  
divalent and trivalent cations, for exam-  
ple, is blocked following preincubation  
with pertussis toxin. Thus, the putative  
polyvalent cation receptor may be coupled  
15 to inhibition of adenylate cyclase by the  
inhibitory guanine nucleotide regulatory  
(G) protein,  $G_i$ .

20 We recently showed that the poly-  
cationic antibiotic, neomycin, mimics the  
actions of di- and trivalent cations in  
several aspects of parathyroid function.  
To determine whether these actions were  
specific to this agent or represented a  
25 more generalized action of polycations, we  
tested the effects of the highly basic pep-  
tides, polyarginine and polylysine, as well  
as protamine on the same parameters in  
dispersed bovine parathyroid cells. The  
30 results demonstrate that the parathyroid  
cell responds to a variety of polycations  
as well as to polyvalent cations, poten-  
tially via similar biochemical pathways.  
These results are discussed in terms of the  
recently postulated, "receptor-independent"  
35 modulation of G proteins by polycations in  
other systems.

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\* \* \*

The  $\text{Ca}^{2+}$  receptor has been presumed to be analogous to other G protein-coupled receptors [e.g., a glycoprotein], but recent studies with other cell types have raised the possibility that polycations can modulate cell function by alternative or additional mechanisms. In mast cells, for example, a variety of amphipathic cations, including mastoparan, a peptide from wasp venom, 48/80, a synthetic polycation, and polylysine, enhance secretion by a pertussis toxin-sensitive mechanism, suggesting the involvement of a G protein. No classic cell surface receptor has been identified that could mediate the actions of these diverse agents. Furthermore, these same compounds have been shown to activate directly purified G proteins in solution or in artificial phospholipid vesicles. On the basis of these observations, it has been proposed that amphipathic cations activate G proteins and, in turn, mast cell secretion by a "receptor-independent" mechanism.

Polycations have also been shown to interact strongly with acidic phospholipids. Polylysines of varying chain lengths (20-1000 amino acids) bind to artificial phospholipid vesicles with dissociation constants in the range of 0.5 nM to 1.5  $\mu\text{M}$ . The binding affinity is directly related to the length of the polylysine chain, with polymers of 1000 amino acids having a  $K_d$  of 0.5 nM, shorter polymers having higher  $K_d$  values, and lysine not interacting to a significant

extent. This relationship between potency and chain length is similar to that observed for the effects of polylysine <sup>10,200</sup>, polylysine <sup>3800</sup>, and lysine on parathyroid function.

It is possible that the binding of polycations to biomembranes produces some of their biologic actions. The permeabilization of the plasma membrane induced in some cell types by a variety of pore-forming agents, including polycations, has been postulated to be mediated by their interaction with a phosphatidylserine-like structure. In addition, the "receptor-independent" activation of purified G proteins by amphipathic cations is potentiated when these proteins are incorporated into phospholipid vesicles.

Calcium ions, in the millimolar concentration range, also produce marked changes in membrane structure. In some cases, calcium can either antagonize or potentiate the interaction of polycations with membrane lipids. These considerations raise the possibility that the actions of both polyvalent cations and polycations on parathyroid cells could involve a receptor-independent mechanism not requiring the presence of a classic, cell surface, G protein-coupled receptor. Further studies, however, are required to elucidate the molecular basis for  $\text{Ca}^{2+}$  sensing by this and other cell types. [Citations omitted.]

Shoback and Chen (6 (Supplement 1), J. Bone and Mineral Res. 1991, S135) and Racke et al. (6 (Supplement 1), J. Bone and Mineral Res. 1991, S118) describe experiments which are said to indicate that a  $\text{Ca}^{2+}$  receptor or

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Ca<sup>2+</sup> sensor is present in parathyroid cells. Messenger RNA isolated from such cells can be expressed in oocytes and caused to provide those oocytes with a phenotype which might be explained by the presence of a Ca<sup>2+</sup> receptor protein.

#### Summary of the Invention

Applicant has demonstrated that Ca<sup>2+</sup> receptor proteins enable certain specialized cells involved in bodily Ca<sup>2+</sup> metabolism to detect and respond to changes in the concentration of extracellular Ca<sup>2+</sup>. Although these receptors share certain general characteristics, they can be selectively affected by different pharmacological agents. As detailed below, certain molecules are identified with selective activity on Ca<sup>2+</sup> receptors at parathyroid cells, osteoclasts, and C-cells.

Ca<sup>2+</sup> receptors constitute discrete molecular targets for a new class of molecules that mimic ("calcimimetics") or antagonize ("calcilytics") the actions of extracellular Ca<sup>2+</sup>. Such receptors are present on cell surfaces and have a low affinity for extracellular Ca<sup>2+</sup> (apparent K<sub>d</sub> generally greater than about 0.5 mM). Such receptors may include a free or bound effector mechanism, as defined by Cooper, Bloom and Roth, "The Biochemical Basis of Neuropharmacology", Ch. 4. Such receptors are thus distinct from intracellular Ca<sup>2+</sup> receptors, e.g., calmodulin and the troponins. Calcimimetics, for example, act on Ca<sup>2+</sup> receptors selectively to directly or indirectly depress the function of parathyroid cells or osteoclasts or to stimulate the function of C-cells. Calcimimetics and calcilytics of this invention allow novel therapies for hyperparathyroidism, osteoporosis and other Ca<sup>2+</sup>-related diseases. This application concerns targeting Ca<sup>2+</sup> receptors on each of these three cell types and other cell types that detect and respond to changes in [Ca<sup>2+</sup>].

Applicant is the first to demonstrate a Ca<sup>2+</sup> receptor protein in parathyroid cells, and to pharmacologically

differentiate such  $\text{Ca}^{2+}$  receptors in other cells, such as C-cells and osteoclasts. Applicant is also the first to describe methods by which molecules active at these  $\text{Ca}^{2+}$  receptors can be identified and used as lead molecules in the discovery, development, design, modification and/or construction of useful calcimimetics or calcilytics which are active at  $\text{Ca}^{2+}$  receptors. Such calcimimetics or calcilytics are useful in the treatment of various disease states characterized by abnormal levels of one or more components, e.g., polypeptides such as hormones, enzymes or growth factors, the expression and/or secretion of which is regulated or affected by activity at one or more  $\text{Ca}^{2+}$  receptors. Further, the identification of different  $\text{Ca}^{2+}$  receptors in different cell types, and the specific response of such receptors to different lead molecules allows design and construction of specific molecules active in treatment of specific diseases which can be affected by action at such specific  $\text{Ca}^{2+}$  receptors. For example, abnormal levels of parathyroid hormone secretion can be affected by such specific molecules without affecting the level of secretion of other  $\text{Ca}^{2+}$  regulated hormones and the like.

Identification of such lead molecules was impeded by the prior lack of a high-throughput screening system to discover active molecules, and the absence of a structural data base upon which to design effective drug candidates. These barriers are now removed by cloning the parathyroid cell  $\text{Ca}^{2+}$  receptor and functionally related receptors, and systematically examining the structural features of certain lead molecules that activate such cloned  $\text{Ca}^{2+}$  receptors and functionally related receptors. Cloning of the  $\text{Ca}^{2+}$  receptor also enables development of transfected cell lines suitable for high-throughput screening of natural product or molecule libraries and synthetic molecules. This, together with structure-activity studies discussed below, provides the technology necessary to develop novel calcimimetics and calcilytics.

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Applicant enables such procedures in this application. For example, the human parathyroid cell  $\text{Ca}^{2+}$  receptor cDNA can be cloned by screening for functional expression in Xenopus oocytes, and the structural features  
5 of organic molecules necessary for activity on the  $\text{Ca}^{2+}$  receptor can be determined through the testing of selected natural products or other molecule libraries and subsequent structure-activity studies.

Thus, in a first aspect, the invention features a  
10 pharmaceutical composition including a molecule which either mimics the activity of extracellular  $\text{Ca}^{2+}$  by evoking an increase in  $[\text{Ca}^{2+}]_i$  in a cell, or blocks an increase in  $[\text{Ca}^{2+}]_i$  elicited by extracellular  $\text{Ca}^{2+}$ . The molecule has an  $\text{EC}_{50}$  of less than or equal to 5  $\mu\text{M}$ , and is not protamine.

15 By "mimic" is meant that the molecule has one or more of the specific actions of extracellular  $\text{Ca}^{2+}$  on an extracellular  $\text{Ca}^{2+}$  responsive cell. The term does not require that all of the biological functions of extracellular  $\text{Ca}^{2+}$  are mimicked, but rather than at least one such function  
20 is mimicked. In addition it does not require that the molecule bind to the same site on the receptor as does extracellular  $\text{Ca}^{2+}$  receptor (see for example, the novel compound NPS 467 and its action in Example 20 below). By "block" is meant that one such action of  $\text{Ca}^{2+}$  is reduced or  
25 prevented by the molecule. The  $\text{EC}_{50}$  can be determined in assays as described below, where the activity mimicked is measured and the concentration of molecule which mimics at half the maximum mimicking effect is the  $\text{EC}_{50}$ . Conversely, the  $\text{IC}_{50}$  of a calcilytic is that amount which blocks half  
30 maximal activity. Preferably, such assays measure  $[\text{Ca}^{2+}]_i$  increases and are confirmed to be specific to a  $\text{Ca}^{2+}$  receptor by methods described below, or their equivalent.

In preferred embodiments, bioassays described herein demonstrated that the increase in  $[\text{Ca}^{2+}]_i$  in a cell is  
35 transient, having a duration of less than one minute, and the increase in  $[\text{Ca}^{2+}]_i$  is rapid, occurring within thirty seconds; and the molecule also (a) evokes a sustained

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increase (greater than thirty seconds) in  $[Ca^{2+}]_i$ , (b) evokes an increase in inositol-1,4,5-trisphosphate and/or diacylglycerol levels, e.g., within less than 60 seconds, and (c) inhibits dopamine- or isoproterenol-stimulated cyclic AMP formation. In addition, the transient increase in  $[Ca^{2+}]_i$  is abolished by pretreatment of the cell for ten minutes with 10 mM sodium fluoride, or the transient increase is diminished by brief pretreatment (not more than ten minutes) of the cell with an activator of protein kinase C, e.g., phorbol myristate acetate (PMA), mezerein or (-)indolactam V.

In a parathyroid cell, those molecules which are active in all of the assays described above are particularly useful in this invention since they are specific in their actions to a  $Ca^{2+}$  receptor of such a cell. This is particularly true for the PMA pretreatment effect described above.

In a more preferred embodiment, the cell is a parathyroid cell, and the molecule inhibits parathyroid hormone secretion from the cell; and the molecule elicits an increase in  $Cl^-$  conductance in a Xenopus oocyte injected with mRNA from a parathyroid cell, bone osteoclast, juxtaglomerular kidney cell, proximal tubule kidney cell, keratinocyte, parafollicular thyroid cell or placental trophoblast.

In other preferred embodiments, the molecule evokes the mobilization of intracellular  $Ca^{2+}$  to cause the increase in  $[Ca^{2+}]_i$ ; the cell is a C-cell or an osteoclast and the molecule inhibits bone resorption in vivo; the cell is an osteoclast and the molecule inhibits bone resorption in vitro; or the cell is a C-cell and the molecule stimulates calcitonin secretion in vitro or in vivo; and most preferably the molecule is either a calcimimetic or calcilytic having an  $EC_{50}$  or  $IC_{50}$  at a  $Ca^{2+}$  receptor of less than or equal to 5  $\mu M$ , and even more preferably less than or equal to 1  $\mu M$ , 100 nmolar, 10 nmolar, or 1 nmolar. Such lower  $EC_{50}$ 's or  $IC_{50}$ 's are

advantageous since they allow lower concentrations of molecules to be used in vivo or in vitro for therapy or diagnosis. The discovery of molecules with such low  $EC_{50}$ 's and  $IC_{50}$ 's enables the design and synthesis of similarly  
5 potent and efficacious molecules.

By "calcimimetic" molecule is meant any molecule which has one or more activities of extracellular  $Ca^{2+}$ , and preferably mimics the activity of  $Ca^{2+}$  at a  $Ca^{2+}$  receptor. For example, when used in reference to a parathyroid cell  
10 it is a molecule, which when tested on parathyroid cells, in vitro, possesses one or more, and preferably all of the following characteristics as measured by techniques well known to those in the art:

1. The molecule causes a rapid (time to peak  
15 < 5 sec) and transient increase in  $[Ca^{2+}]_i$  that is refractory to inhibition by  $1 \mu M$   $La^{3+}$  or  $Gd^{3+}$ . The increase in  $[Ca^{2+}]_i$  persists in the absence of extracellular  $Ca^{2+}$  but is abolished by pretreatment with ionomycin (in the absence of extracellular  $Ca^{2+}$ );

20 2. The increase in  $[Ca^{2+}]_i$  elicited by extracellular  $Ca^{2+}$  is not inhibited by dihydropyridines.

3. The transient increase in  $[Ca^{2+}]_i$  caused by the molecule is abolished by pretreatment for 10 min. with 10 mM sodium fluoride;

25 4. The transient increase in  $[Ca^{2+}]_i$  caused by the molecule is diminished by pretreatment with an activator of protein kinase C (PKC), such as phorbol myristate acetate (PMA), mezerein or (-)-indolactam V. The overall effect of the protein kinase C activator is to shift the  
30 concentration-response curve of the molecule to the right without affecting the maximal response;

5. The molecule causes a rapid (< 30 sec.) increase in the formation of inositol-1,4,5-trisphosphate and or diacylglycerol;

35 6. The molecule inhibits dopamine- or isoproterenol-stimulated cyclic AMP formation;

7. The molecule inhibits PTH secretion;

8. Pretreatment with pertussis toxin (100 ng/ml for > 4 hrs.) blocks the inhibitory affect of the molecule on cyclic AMP formation but does not effect increases in  $[Ca^{2+}]_i$ , inositol-1,4,5-trisphosphate, or diacylglycerol, nor decreases in PTH secretion;

9. The molecule elicits increases in  $Cl^-$  conductance in Xenopus oocytes injected with poly(A)<sup>+</sup>- enriched mRNA from bovine or human parathyroid cells but is without effect in Xenopus oocytes injected with water or rat brain or liver mRNA; and

10. Similarly, using a cloned receptor from parathyroid cells, the molecule will elicit a response in Xenopus oocytes injected with the specific cDNA or mRNA encoding the receptor.

15 By "calcilytic" molecule is meant any molecule which blocks one or more of the activities of extracellular  $Ca^{2+}$  on an extracellular  $Ca^{2+}$ -sensing cell, preferably by acting as an antagonist at the  $Ca^{2+}$  receptor. For example, when used in reference to a parathyroid cell, it is a molecule which, when tested on parathyroid cells in vitro, possesses one or more, and preferably all of the following characteristics as measured by techniques well known to those in the art:

25 1. The molecule blocks, either partially or completely, the ability of increased concentrations of extracellular  $Ca^{2+}$  to:

- a) increase  $[Ca^{2+}]_i$ ,
- b) mobilize intracellular  $Ca^{2+}$ ,
- c) increase the formation of inositol-1,4,5-trisphosphate,
- d) decrease dopamine- or isoproterenol-stimulated cyclic AMP formation, and
- e) inhibit PTH secretion;

35 2. At low  $[Ca^{2+}]$ , i.e., 0.5 mM, the molecule by itself does not change  $[Ca^{2+}]_i$

3. The molecule blocks increases in  $Cl^-$  conductance in Xenopus oocytes injected with poly(A)<sup>+</sup>-mRNA from bovine

or human parathyroid cells elicited by extracellular  $\text{Ca}^{2+}$  or calcimimetic compounds but not in Xenopus oocytes injected with water or rat brain or liver mRNA;

4. Similarly, using a cloned receptor from parathyroid cells, the molecule will block a response in Xenopus oocytes injected with the specific cDNA or mRNA encoding the  $\text{Ca}^{2+}$  receptor, elicited by extracellular  $\text{Ca}^{2+}$  or a calcimimetic compound.

Parallel definitions of useful calcimimetics and calcilytics at  $\text{Ca}^{2+}$  receptors on other cell types are evident from the examples provided below.

The  $\text{Ca}^{2+}$  receptor is able to detect and respond to certain inorganic polycations and polycationic organic molecules. For example, the parathyroid cell is unable to distinguish increases in extracellular  $\text{Ca}^{2+}$  concentration from the addition of these organic polycations, presumably because these organic molecules act just like extracellular  $\text{Ca}^{2+}$  at the  $\text{Ca}^{2+}$  receptor. The calcimimetic molecules of this invention are particularly good agonists of the  $\text{Ca}^{2+}$  receptor and may be used as drugs that alter selected cellular functions, e.g., secretion of PTH from parathyroid cells. Unlike  $\text{Ca}^{2+}$  most of these molecules act only at one or more, but not all  $\text{Ca}^{2+}$  receptors, and thus provide an ability to specifically target one  $\text{Ca}^{2+}$  receptor.

These molecules also provide lead structures for the development of further novel therapeutics effective in the treatment of various diseases where  $[\text{Ca}^{2+}]_i$  and  $[\text{Ca}^{2+}]_o$  play a role, such as hyperparathyroidism, osteoporosis, Paget's disease, hypertension, renal disease, and cancer.

The calcimimetics and calcilytics can be formulated as pharmaceutical compositions which are useful for regulating the level of extracellular free  $\text{Ca}^{2+}$  in a patient and for mimicking the effect of extracellular  $\text{Ca}^{2+}$  on a cell selected from the group described above, by administering to the patient such a pharmaceutical composition. Prior to this invention, applicant was unaware of any such

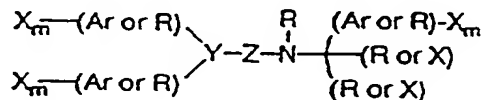
molecules acting on the  $\text{Ca}^{2+}$  receptor useful in treatment of diseases caused by irregularity in operation or regulation of a  $\text{Ca}^{2+}$  receptor or diseases in an animal having normal  $\text{Ca}^{2+}$  receptors but which can be treated by activating or deactivating such  $\text{Ca}^{2+}$  receptors.

In yet another preferred embodiment, the molecule has an  $\text{EC}_{50}$  less than or equal to 5  $\mu\text{M}$  at one or more but not all cells chosen from the group consisting of parathyroid cells, bone osteoclasts, juxtaglomerular kidney cells, proximal tubule kidney cells, keratinocytes, parafollicular thyroid cells (C-cells) and placental trophoblasts.

It is the specificity of action of such molecules that is particularly advantageous in this invention since it allows specific in vivo and in vitro therapy and diagnosis and discovery of additional calcimimetic or calci-lytic molecules.

In specific preferred embodiments, the molecule is positively charged at physiological pH, and is selected from the group consisting of branched or cyclic polyamines, positively charged polyamino acids, and arylalkylamines, e.g., the branched polyamine has the formula  $\text{H}_2\text{N}-(\text{CH}_2)_j-(\text{NR}_i-(\text{CH}_2)_j)_k-\text{NH}_2$  where  $k$  is an integer from 1 to 10, each  $j$  is the same or different and is an integer from 2 to 20, and each  $R_i$  is the same or different and is selected from the group consisting of hydrogen and  $-(\text{CH}_2)_j-\text{NH}_2$ , where  $j$  is as defined above, and at least one  $R_i$  is not hydrogen.

In an alternative embodiment, the molecule has the formula

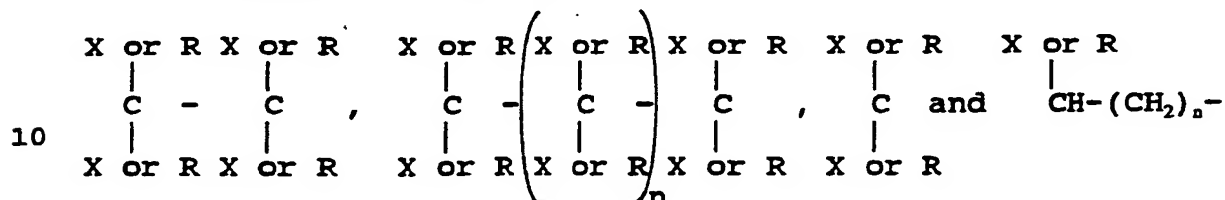


where each X independently is selected from the group consisting of H,  $\text{CH}_3$ ,  $\text{CH}_3\text{O}$ ,  $\text{CH}_3\text{CH}_2\text{O}$ , Br, Cl, F,  $\text{CF}_3$ ,  $\text{CHF}_2$ ,  $\text{CH}_2\text{F}$ ,  $\text{CF}_3\text{O}$ ,  $\text{CH}_3\text{S}$ , OH,  $\text{CH}_2\text{OH}$ ,  $\text{CONH}_2$ , CN,  $\text{NO}_2$ , and  $\text{CH}_3\text{CH}_2$ ; Ar is a hydrophobic entity; each R independently is selected from the group consisting of hydrogen, methyl, ethyl, propyl,

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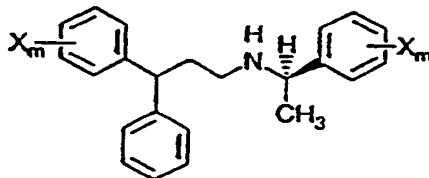
15

isopropyl, butyl, isobutyl, cyclopentyl, cyclohexyl, cycloheptyl, cyclooctyl, indenyl, indanyl, dihydroindolyl, thiodihydroindolyl, 2-, 3-, or 4-piperid(in)yl; Y is selected from the group consisting of CH, nitrogen and an  
 5 unsaturated carbon; Z is selected from the group consisting of oxygen, nitrogen, sulfur,



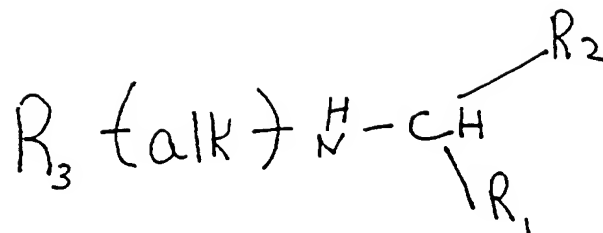
where each n is independently between 1 and 4 inclusive, and each m is independently between 0 and 5 inclusive.  
 15 Most preferably the molecule is either a calcimimetic or calcilytic.

In preferred embodiments, the hydrophobic entity is selected from the group consisting of phenyl, 2-, 3-, or 4-pyridyl, 1- or 2-naphthyl, 1- or 2-quinolinyl, 2- or 3-  
 20 indolyl, benzyl, and phenoxy; the molecule is an R-diphenylpropyl- $\alpha$ -phenethylamine derivative, and the molecule has the formula:



with each X preferably being independently selected from the group consisting of Cl, F, CF<sub>3</sub>, CH<sub>3</sub>, and CH<sub>3</sub>O.

25 According to a preferred aspect of the present invention, novel phenyl- $\alpha$ -phenethylamine analogs and derivatives are provided having the formula:



wherein alk is straight or branched chain alkylene of from 1 to 6 carbon atoms; R<sub>1</sub> is lower alkyl of from 1 to 3 carbon atoms or lower haloalkyl of from 1 to 3 carbon atoms substituted with from 1 to 7 halogen atoms; R<sub>2</sub> and R<sub>3</sub> are independently selected carbocyclic aryl or cycloalkyl groups, either monocyclic or bicyclic, having 5- or 6-membered rings optionally substituted with 1 to 5 substituents independently selected from lower alkyl of 1 to 3 carbon atoms, lower haloalkyl of 1 to 3 carbon atoms substituted with 1 to 7 halogen atoms, lower alkoxy of 1 to 3 carbon atoms, halogen, nitro, amino, alkylamino, amido, lower alkylamido of 1 to 3 carbon atoms, cyano, hydroxy, acyl of 2 to 4 carbon atoms lower hydroxyalkyl of 1 to 3 carbon atoms or lower thioalkyl of 1 to 3 carbon atoms. Suitable carbocyclic aryl groups are groups having one or two rings, at least one of which having aromatic character and include carbocyclic aryl groups such as phenyl and bicyclic carbocyclic aryl groups such as naphthyl. As is apparent from the above formula, the compounds encompassed therein may exist as racemic mixtures and as individual stereoisomers. Especially preferred are R-phenylpropyl

$\alpha$ -phenethylamine derivatives which are believed to exhibit enhanced activity in lowering serum ionized calcium.

Preferred compounds include those where alk is n-propylene. Also preferred are compounds where  $R_1$  is methyl. Also preferred are those compounds where  $R_2$  and  $R_3$  are optionally substituted phenyl.

Especially preferred compounds include those where  $R_2$  is monosubstituted phenyl, more preferably meta-substituted. Especially preferred  $R_3$  groups include unsubstituted or monosubstituted phenyl, especially ortho-substituted. Preferred substituents for  $R_2$  include halogen, haloalkyl, preferably trihalomethyl, and alkoxy, preferably methoxy. Preferred substituents for  $R_3$  include halogen.

In a second related aspect, the invention features a method for treating a patient having a disease or condition characterized by an abnormal  $[Ca^{2+}]$  or  $[Ca^{2+}]_i$  in one or more cells or in the blood or plasma or extracellular fluids. The method includes the step of administering to the patient a therapeutically effective amount of a molecule which either mimics the activity of extracellular  $Ca^{2+}$  by evoking an increase in  $[Ca^{2+}]_i$  in a cell or blocks an increase in  $[Ca^{2+}]_i$  elicited by extracellular  $Ca^{2+}$ .

By "abnormal" is meant that the patient, compared to the general population, has a different  $Ca^{2+}$  metabolism that is affected by one or more proteins (e.g., hormones) in the blood or extracellular body fluids, or other molecules which affect the level of extracellular and/or intracellular  $Ca^{2+}$ . Thus, the diseases include hyperparathyroidism, osteoporosis and other bone and mineral-related disorders, and the like (as described, e.g., in standard medical text books, such as "Harrison's Principles of Internal Medicine"). Such diseases are treated in this invention by molecules which mimic or block one or more of the effects of  $Ca^{2+}$  and thereby directly or indirectly affect the levels of the proteins or other molecules in the body of the patient.



By "therapeutically effective amount" is meant an amount that relieves to some extent one or more symptoms of the disease or condition in the patient. Additionally, by "therapeutically effective amount" is meant an amount  
5 that returns to normal, either partially or completely, physiological or biochemical parameters associated with or causative of the disease or condition. Generally, it is an amount between about 1 nmole and 1  $\mu$ mole of the molecule, dependent on its  $EC_{50}$  and on the age, size, and  
10 disease associated with the patient.

In preferred embodiments, the molecule has an  $EC_{50}$  of less than or equal to 5  $\mu$ M, and is not protamine; and most preferably interacts at a  $Ca^{2+}$  receptor as a calcimimetic or calcilytic. Most preferably the molecule is chosen  
15 from one of those described above.

In other preferred embodiments, the patient has a disease characterized by an abnormal level of one or more components the level of which is regulated or affected by activity of one or more  $Ca^{2+}$  receptors, and the molecule is  
20 active on a  $Ca^{2+}$  receptor of a cell selected from the group consisting of parathyroid cells, bone osteoclasts, juxta-glomerular kidney cells, proximal tubule kidney cells, keratinocytes, parafollicular thyroid cells, and placental trophoblasts.

In still other preferred embodiments, the molecule reduces the level of parathyroid hormone in the serum of the patient, e.g., to that level present in a normal individual, or to a degree sufficient to cause a decrease in plasma  $Ca^{2+}$ ; and the molecule is provided in an amount  
30 sufficient to have a therapeutically relevant effect on the patient.

In a third aspect, the invention features a method for diagnosis of a disease or condition in a patient by identifying the number and/or location (and/or functional  
35 integrity) of one or more  $Ca^{2+}$  receptors within the patient and comparing that number and/or location (and/or func-

tional integrity) with that observed in normal patients as an indication of the presence of the disease or condition.

In preferred embodiments, the method is an immunoassay in which an antibody to a  $\text{Ca}^{2+}$  receptor is used to  
5 identify the number and/or location and/or functional integrity of the  $\text{Ca}^{2+}$  receptors, or the assay involves providing a labelled calcimimetic or calcilytic molecule which binds to a  $\text{Ca}^{2+}$  receptor; and the disease diagnosed  
10 is a cancer, e.g., an ectopic tumor of the parathyroid, or a condition characterized by an above normal level in the number of osteoclasts in bone or an increased level of activity of osteoclasts in bone.

In a fourth aspect, the invention features a method for identifying a molecule useful as a therapeutic mole-  
15 cule. The method includes screening a potentially useful molecule for either an ability to mimic the activity of extracellular  $\text{Ca}^{2+}$  in a cell, or to block an increase in  $[\text{Ca}^{2+}]_i$  elicited by extracellular  $\text{Ca}^{2+}$ , and determining whether the molecule has an  $\text{EC}_{50}$  or  $\text{IC}_{50}$  of less than or  
20 equal to 5  $\mu\text{M}$ .

In other aspects, the invention features a recombinant  $\text{Ca}^{2+}$  receptor, a cell including a recombinant  $\text{Ca}^{2+}$  receptor, purified nucleic acid encoding a  $\text{Ca}^{2+}$  receptor, the biological activity and use of the molecule NPS 019,  
25 the novel compounds or compositions of matter of NPS 459, NPS 467, NPS 551, and NPS 568 (see Fig. 36) and a method for identifying a useful calcimimetic or calcilytic molecule by identifying a molecule which mimics or blocks one or more activities of  $\text{Ca}^{2+}$  at a first  $\text{Ca}^{2+}$  receptor but not  
30 at a second  $\text{Ca}^{2+}$  receptor, e.g., by use of a recombinant  $\text{Ca}^{2+}$  receptor.

By "recombinant" is meant to include any  $\text{Ca}^{2+}$  receptor produced by recombinant DNA techniques such that it is distinct from the naturally occurring  $\text{Ca}^{2+}$  receptor either  
35 in its location, purity or structure. Generally, such a receptor will be present in a cell in an amount different from those normally observed in nature.

By "purified" is meant that the antibody or nucleic acid is distinct from naturally occurring antibody or nucleic acid, being separated from antibody or nucleic acid with which it naturally occurs, e.g., in a vector system, such that it can be used to express recombinant  $\text{Ca}^{2+}$  receptor. Preferably, the antibody or nucleic acid is provided as a homogeneous preparation by standard techniques.

Such cloned receptors can be expressed in a desired cell, and isolated and crystallized to allow structure determination. Such a structure will allow design of useful molecules of this invention which can bind to the  $\text{Ca}^{2+}$  receptor. In addition, equivalent such receptors can be cloned using a first clone as a probe for clones in other cell, cDNA or genomic libraries.

Antibodies to the cloned receptor can be isolated and used as therapeutics in this invention, or as diagnostic tools for determining  $\text{Ca}^{2+}$  receptor numbers and/or locations and/or functional integrity to diagnose  $\text{Ca}^{2+}$ -related diseases or conditions. Such antibodies can also be used in vivo by intravenous administration as calcimimetics or calcilytics.

Thus, in general, the invention features calcimimetic or calcilytic molecules able to act as either selective agonists or antagonists respectively at a  $\text{Ca}^{2+}$  receptor of one or more but not all cells chosen from the group consisting of parathyroid cells, bone osteoclasts, juxtaglomerular kidney cells, proximal tubule kidney cells, keratinocytes, parafollicular thyroid cells and placental trophoblasts. Such a composition may include any pharmaceutically acceptable carrier known to those in the art to provide a pharmaceutical composition.

The invention also features modulation of the the number of  $\text{Ca}^{2+}$  receptors in a patient by standard techniques, e.g., antisense and related technologies (e.g., ribozymes), as a therapeutic for a disease state.

This invention provides methods for identifying molecules which affect the activity of a  $\text{Ca}^{2+}$  receptor using assays, as defined below, to detect calcimimetics and/or calcilytics. Further, molecules found to be effective to  
5 reduce or enhance expression of  $\text{Ca}^{2+}$  receptor at a transcriptional or translational level by use of the assays or antibodies or other techniques described below can be defined for therapeutic uses.

Other features and advantages of the invention will  
10 be apparent from the following description of the preferred embodiments thereof, and from the claims.

#### Description of the Preferred Embodiments

The drawings will first briefly be described.

#### Drawings

15 Fig. 1 depicts representative molecules useful in the invention.

Fig. 2 is a graphical representation showing increases in  $[\text{Ca}^{2+}]_i$  induced by extracellular  $\text{Ca}^{2+}$  in quin-2- or fura-2-loaded bovine parathyroid cells. The initial  
20  $[\text{Ca}^{2+}]$  was 0.5 mM (using  $\text{CaCl}_2$ ) and, at each of the arrows, was increased in 0.5 mM increments.

Fig. 3 is a graphical representation showing mobilization of  $[\text{Ca}^{2+}]_i$  in bovine parathyroid cells. The initial  $[\text{Ca}^{2+}]$  was 0.5 mM and was decreased to  $< 1 \mu\text{M}$  by the addition of EGTA as indicated. (a) Extracellular  $\text{Mg}^{2+}$  (8 mM, final) elicits an increase in  $[\text{Ca}^{2+}]_i$  in the absence of extracellular  $\text{Ca}^{2+}$ . (b) Pretreatment with ionomycin ( $1 \mu\text{M}$ ) blocks the response to  $\text{Mg}^{2+}$ . (c) Pretreatment with 5  $\mu\text{M}$  molecule 1799 (a mitochondrial uncoupler) is without  
25 effect on the response to  $\text{Mg}^{2+}$ .  
30

Fig. 4 is a graphical representation showing preferential inhibitory effects of a low concentration of  $\text{Gd}^{3+}$  on steady-state increases in  $[\text{Ca}^{2+}]_i$  and that a high concentration of  $\text{Gd}^{3+}$  elicits a transient increase in  $[\text{Ca}]_i$ . Top  
35 panel: Control. Initial concentration of extracellular

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$\text{Ca}^{2+}$  was 0.5 mM and was increased by 0.5 mM at each of the arrowheads. Middle panel:  $\text{Gd}^{3+}$  (5  $\mu\text{M}$ ) blocks steady-state but not transient increases in  $[\text{Ca}^{2+}]_i$  elicited by extracellular  $\text{Ca}^{2+}$ . Lower panel:  $\text{Gd}^{3+}$  (50  $\mu\text{M}$ ) elicits a transient increase in  $[\text{Ca}^{2+}]_i$  and abolishes both transient and sustained responses to extracellular  $\text{Ca}^{2+}$ . In the middle and lower panels, just enough EGTA was added to chelate preferentially  $\text{Gd}^{3+}$ : the block of  $\text{Ca}^{2+}$  influx is removed and  $[\text{Ca}^{2+}]_i$  rises promptly.

10 Fig. 5 is a graphical representation showing that the effects of PMA on  $[\text{Ca}^{2+}]_i$ , IP<sub>3</sub> formation, and PTH secretion are overcome by increasing concentrations of extracellular  $\text{Ca}^{2+}$ . For each variable, there is a shift to the right in the concentration-response curve for extracellular  $\text{Ca}^{2+}$ .  
15 Note also that the concentration-response curves vary sigmoidally as  $[\text{Ca}^{2+}]$  increases linearly.

Fig. 6 is a graphical representation showing that increases in  $[\text{Ca}^{2+}]_i$  elicited by spermine are progressively depressed by increasing  $[\text{Ca}^{2+}]$ . Spermine (200  $\mu\text{M}$ ) was  
20 added at the time shown by arrowheads. In this and all subsequent figures, the numbers accompanying the traces are  $[\text{Ca}^{2+}]_i$  in nM.

Fig. 7 is a graphical representation showing that spermine mobilizes intracellular  $\text{Ca}^{2+}$  in bovine parathyroid  
25 cells. EGTA was added to reduce  $[\text{Ca}^{2+}]$  to  $<1 \mu\text{M}$  before the addition of spermine (200  $\mu\text{M}$ ) as indicated (left trace). Pretreatment with ionomycin (1  $\mu\text{M}$ ) blocks the response to spermine (right trace).

Figs. 8A and B are graphical representations showing  
30 that spermine increases  $[\text{Ca}^{2+}]_i$  and inhibits PTH secretion in bovine parathyroid cells similarly to extracellular  $\text{Ca}^{2+}$ . The data points for the spermine dose-concentration response curves are the means of two experiments.

Fig. 9 is a graphical representation showing the contrasting effects of PMA on responses to extracellular  $\text{Ca}^{2+}$   
35 and on responses to  $\text{ATP}\gamma\text{S}$  in bovine parathyroid cells.

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Left panel: The concentration-response curve for extracellular  $\text{Ca}^{2+}$ -induced inhibition of cyclic AMP formation is shifted to the right by PMA (100 nM). Middle panel: PMA does not affect the ability of  $\text{ATP}\gamma\text{S}$  to increase  $[\text{Ca}^{2+}]_i$ .

- 5 Note also that the concentration-response curve to  $\text{ATP}\gamma\text{S}$  shows classical sigmoidal behavior as a function of the log concentration, in contrast to extracellular divalent cations.

Fig. 10 is a graphical representation showing mobilization of intracellular  $\text{Ca}^{2+}$  in human parathyroid cells evoked by extracellular  $\text{Mg}^{2+}$ . Cells were obtained from an adenoma and bathed in buffer containing 0.5 mM extracellular  $\text{Ca}^{2+}$ . (a) Transient and sustained increases in  $[\text{Ca}^{2+}]_i$  elicited by extracellular  $\text{Mg}^{2+}$  (10 mM, final) shows that  
15 sustained increases are not affected by nimodipine (1  $\mu\text{M}$ ) but are depressed by  $\text{La}^{3+}$  (1  $\mu\text{M}$ ) and return promptly when  $\text{La}^{3+}$  is selectively chelated by a low concentration of EGTA. (b)  $\text{La}^{3+}$  (1  $\mu\text{M}$ ) blocks the sustained but not the transient increase in  $[\text{Ca}^{2+}]_i$  elicited by extracellular  
20  $\text{Mg}^{2+}$ . (c) Cytosolic  $\text{Ca}^{2+}$  transients elicited by extracellular  $\text{Mg}^{2+}$  persist in the absence of extracellular  $\text{Ca}^{2+}$ .

Fig. 11 is a graphical representation showing mobilization of intracellular  $\text{Ca}^{2+}$  evoked by neomycin or protamine in bovine parathyroid cells. In all traces, the initial  $[\text{Ca}^{2+}]$  and  $[\text{Mg}^{2+}]$  was 0.5 and 1 mM, respectively. In  
25 trace (a) and (b), the  $\text{Ca}^{2+}$  and  $\text{Mg}^{2+}$  concentrations were increased to 2 and 8 mM, from 0.5 and 1 mM respectively. In the other traces, (c) through (i), neomycin B (30  $\mu\text{M}$ ) or protamine (1  $\mu\text{g/ml}$ ) were added as indicated.  $\text{La}^{3+}$   
30 (1  $\mu\text{M}$ ) EGTA (1 mM), or ionomycin (100 nM) were added as indicated. Each trace is representative of the pattern seen in 5 or more trials using at least 3 different cell preparations. Bar = 1 min.

Fig. 12 is a graphical representation showing that  
35 neomycin B blocks transient but does not block steady-state increases in  $[\text{Ca}^{2+}]_i$  elicited by extracellular  $\text{Ca}^{2+}$ . Left control:  $[\text{Ca}^{2+}]$  was initially 0.5 mM and was

increased in 0.5 mM increments at each of the open arrow-heads before the addition of neomycin B (30  $\mu$ M). Right: Neomycin B (30  $\mu$ M) was added before  $[Ca^{2+}]_i$ . Bar = 1 min.

Fig. 13 is a graphical representation showing that  
5 neomycin B or protamine inhibit PTH secretion at concentrations which evoked increases in  $[Ca^{2+}]_i$ . Cells were incubated with the indicated concentrations of organic polycation for 30 min. in the presence of 0.5 mM extracellular  $Ca^{2+}$ . Open symbols: control responses for PTH  
10 secretion in the presence of 0.5 (circles) or 2 mM (diamonds) extracellular  $Ca^{2+}$ . Values for  $[Ca^{2+}]_i$  are diamond symbols. Bovine cells were used in the experiments with protamine and human (adenoma) parathyroid cells were used in the experiments with neomycin B. Each point is the  
15 mean  $\pm$  SEM of 3 experiments.

Fig. 14 is a graphical representation showing the preferential inhibitory effects of PMA on cytosolic  $Ca^{2+}$  transients elicited by spermine. Initial  $[Ca^{2+}]$  was 0.5 mM; spermine (200  $\mu$ M) or ATP (50  $\mu$ M) were added as indicated. Bar = 1 min.  
20

Fig. 15 is a graphical representation showing that PMA shifts to the right the concentration-response curves for extracellular  $Ca^{2+}$ - and neomycin B-induced increases in  $[Ca^{2+}]_i$ . Cells were pretreated with PMA for 1 min. before  
25 increasing  $[Ca^{2+}]$  or before adding neomycin B as indicated. Each point is the mean  $\pm$  SEM of 3 to 5 experiments.

Fig. 16 is a graphical representation showing that PMA shifts to the right the concentration-response curves for extracellular  $Ca^{2+}$ - and spermine-induced inhibition of  
30 PTH secretion. Cells were incubated with the indicated  $[Ca^{2+}]$  and spermine for 30 min. in the presence (closed circles) or absence (open circles) of 100 nM PMA. Each point is the mean  $\pm$  SEM of 3 experiments.

Fig. 17 is a graphical representation showing that  
35 protamine increases the formation of inositol phosphates. Parathyroid cells were incubated overnight in culture media containing 4 uCi/ml  $^3H$ -myo-inositol, washed, and

incubated with the indicated concentration of protamine at 37°. After 30 sec. the reaction was terminated by the addition of CHCl<sub>3</sub>:MeOH:HCl and IP<sub>1</sub> (circles) and IP<sub>3</sub> (triangles) separated by anion exchange chromatography.

5 Each point is the mean of 2 experiments, each performed in triplicate.

Fig. 18 is a graphical representation showing that PMA depresses the formation of IP<sub>1</sub> evoked by extracellular Ca<sup>2+</sup> or spermine. <sup>3</sup>H-myoinositol-labeled cells were  
10 exposed to the indicated [Ca<sup>2+</sup>] or spermine for 30 sec. before terminating the reaction and determining IP<sub>1</sub> by anion exchange chromatography. Hatched columns: Cells were pretreated with PMA (100 nM) for 5 min. before increasing [Ca<sup>2+</sup>] or adding spermine. Each value is the  
15 mean of 2 experiments, each performed in triplicate.

Fig. 19 is a graphical representation showing transient and sustained increases in [Ca<sup>2+</sup>]<sub>i</sub> elicited by neomycin B in human (adenoma) parathyroid cells. [Ca<sup>2+</sup>] was 0.5 mM. (a) The sustained increase in [Ca<sup>2+</sup>]<sub>i</sub> elicited by  
20 neomycin B (10 μM) was depressed by La<sup>3+</sup>. (b) The transient increase in [Ca<sup>2+</sup>]<sub>i</sub> evoked by neomycin B was unaffected by La<sup>3+</sup>. (c) Transient increases in [Ca<sup>2+</sup>]<sub>i</sub> persisted in the absence of extracellular Ca<sup>2+</sup>.

Fig. 20 is a graphical representation showing that  
25 neomycin B evokes oscillating increases the Cl<sup>-</sup> conductance in Xenopus oocytes expressing the Ca<sup>2+</sup> receptor. Upper trace from an oocyte three days after injection with human (hyperplastic) parathyroid cell poly(A)<sup>+</sup>-mRNA. Lower trace from an oocyte injected with water. Neomycin B failed to  
30 elicit a response in five water-injected oocytes and carbachol elicited a response in one, which is shown. In both traces, the holding potential was -76 mV.

Fig. 21 is a graphical representation showing that neomycin B fails to affect basal or evoked increases in  
35 C-cells. Control, left trace: Fura-2-loaded rMTC 6-23 cells were initially bathed in buffer containing 1 mM Ca<sup>2+</sup>

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before increasing  $[Ca^{2+}]$  to 3 mM. Right trace: pretreatment with 5 mM neomycin B.

Fig. 22 is a graphical representation showing that extracellular  $Ca^{2+}$  evokes increases in  $[Ca^{2+}]_i$  in rat osteoclasts. Microfluorimetric recording in a single rat osteoclast loaded with indo-1 and superfused for the indicated times (bars) with buffer containing the indicated  $[Ca^{2+}]$ . Normal buffer, superfused between the bars, contained 1 mM  $Ca^{2+}$ .

Fig. 23 is a graphical representation showing that spermine or neomycin B fail to evoke increases in  $[Ca^{2+}]_i$  in rat osteoclasts. An indo-1-loaded osteoclast was superfused with the indicated concentration of spermine or neomycin B (open bars) alone or together with 20 mM  $Ca^{2+}$  (solid bars).

Fig. 24 is a graphical representation showing the differential effects of argiotoxin (shown as argiopine in the figure, structures also shown in Fig. 1). 659 and argiotoxin 636 on  $[Ca^{2+}]_i$  in bovine parathyroid cells. The initial  $[Ca^{2+}]$  was 0.5 mM and was increased to 1.5 mM where indicated (right trace). Where indicated, argiotoxin 659 (300  $\mu$ M) or argiotoxin 636 (400  $\mu$ M) was added.

Fig. 25 is a graphical representation showing that extracellular  $Mg^{2+}$  or  $Gd^{3+}$  evoke oscillatory increases in  $Cl^-$  conductance in Xenopus oocytes injected with bovine parathyroid cell poly(A)<sup>+</sup>-mRNA. In trace (a), the concentration of extracellular  $Ca^{2+}$  was < 1  $\mu$ M and in trace (b), 0.7 mM. Trace (c) shows that extracellular  $Mg^{2+}$  fails to elicit a response in an oocyte injected only with the mRNA for the substance K receptor, although superfusion with substance K evokes a response. Holding potential was -70 to -80 mV.

Fig. 26 is a graphical representation showing that extracellular  $Ca^{2+}$  elicits oscillatory increases in  $Cl^-$  conductance in Xenopus oocytes injected with human (hyperplastic) parathyroid tissue poly(A)<sup>+</sup>-mRNA. The oocyte was tested for responsivity to extracellular  $Ca^{2+}$  three days

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after injection of 50 ng poly(A)<sup>+</sup>-mRNA. Holding potential was -80 mV.

Fig. 27 is a graphical representation showing the mobilization of intracellular Ca<sup>2+</sup> in bovine parathyroid cells elicited by budmunchiamine. Budmunchiamine (300 μM, structure also shown) was added where indicated.

Fig. 28 is a graphical representation showing that the ability to mobilize intracellular Ca<sup>2+</sup> in parathyroid cells is stereospecific. Bovine parathyroid cells loaded with fura-2 were initially suspended in buffer containing 0.5 mM extracellular Ca<sup>2+</sup> before the addition of the indicated concentration of each molecule.

Fig. 29 is a graphical representation showing effects of La<sup>3+</sup> on [Ca<sup>2+</sup>]<sub>i</sub> in osteoclasts. A representative trace from a single rat osteoclast loaded with indo-1 is shown. At low concentrations, La<sup>3+</sup> partially blocks increases in [Ca<sup>2+</sup>]<sub>i</sub> elicited by extracellular Ca<sup>2+</sup>.

Figs. 30A and B are graphical representations showing the mobilization of intracellular Ca<sup>2+</sup> elicited by extracellular Mn<sup>2+</sup> in rat osteoclasts. Extracellular Mn<sup>2+</sup> evokes concentration-dependent increases in [Ca<sup>2+</sup>]<sub>i</sub> (Fig. 30A) that persist in the absence of extracellular Ca<sup>2+</sup> (Fig. 30B).

Figs. 31A and 31B are graphical representations showing mobilization of [Ca<sup>2+</sup>]<sub>i</sub> in rat osteoclasts elicited by a molecule termed NPS 449 (see Fig. 38). Isolated rat osteoclasts loaded with indo-1 were superfused with the indicated concentrations of NPS 449 in the presence (Fig. 31A) or absence (Fig. 31B) of 1 mM extracellular CaCl<sub>2</sub>.

Fig. 32 is a graphical representation showing the mobilization of intracellular Ca<sup>2+</sup> in C-cells evoked by NPS 019 (see Fig. 1). rMTC 6-23 cells were loaded with fura-2 and bathed in buffer containing 0.5 mM [Ca<sup>2+</sup>]. Where indicated, NPS 019 was added to a final concentration of 10 μM. Representative traces show that the transient increase in [Ca<sup>2+</sup>]<sub>i</sub> elicited by NPS 019 is refractory to

inhibition by  $\text{La}^{3+}$  (middle trace) and persists in the absence of extracellular  $\text{Ca}^{2+}$  (right trace).

Fig. 33 is a graphical representation showing that NPS 456 (Fig. 36) evokes oscillatory increases in  $\text{Cl}^-$  current in Xenopus oocytes which have been injected with bovine parathyroid cell poly(A)<sup>+</sup>-mRNA.

Fig. 34 is a graphical representation showing that extracellular  $\text{Ca}^{2+}$  evokes oscillatory increases in  $\text{Cl}^-$  current in Xenopus oocytes which have been injected with human osteoclast mRNA. The oocyte was tested for responsiveness to extracellular  $\text{Ca}^{2+}$  three days after injection of 50 ng of total poly(A)<sup>+</sup> mRNA.

Fig. 35 is a graphical representation showing that the parathyroid cell  $\text{Ca}^{2+}$  receptor is encoded by mRNA in a size range of 2.5-3.5 kb. Bovine parathyroid cell poly(A)<sup>+</sup>-mRNA was size fractionated on denaturing glycerol gradients and pooled into ten fractions. Each fraction was injected (50 ng/fraction) separately into Xenopus oocytes. After three days, the oocytes were examined for their ability to respond to extracellular  $\text{Ca}^{2+}$  with oscillatory increases in the  $\text{Cl}^-$  conductance.

Fig. 36 shows the chemical structures of molecules derived from diphenylpropyl- $\alpha$ -phenethylamine illustrating a family of molecules which were prepared and screened to find the useful molecules of the invention.

Fig. 37 is a graphical representation showing that NPS 021 is a calcilytic compound that blocks the effects of extracellular  $\text{Ca}^{2+}$  on  $[\text{Ca}^{2+}]_i$  in bovine parathyroid cells. Cells were initially bathed in buffer containing 0.5 mM  $\text{CaCl}_2$  and, where indicated, the  $[\text{Ca}^{2+}]$  was increased to a final of 2mM (left trace). The addition of NPS 021 (200  $\mu\text{M}$ ) caused no change in  $[\text{Ca}^{2+}]_i$  but inhibited the increase in  $[\text{Ca}^{2+}]_i$  elicited by extracellular  $\text{Ca}^{2+}$  (right trace).

Fig. 38 is a graph showing in vivo  $\text{Ca}^{2+}$  response to NPS 467.

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Fig. 39 is a graph showing in vivo PTH response to NPS 467.

Fig. 40 is a graph showing in vivo  $\text{Ca}^{2+}$  response to 25 mg/kg NPS 467.

5 Figs. 41 and 42 are graphs showing in vivo  $\text{Ca}^{2+}$  responses to different enantiomers of NPS 467.

Fig. 43a depicts a reaction scheme for the preparation of fendiline or fendiline analogs or derivatives depicted in Figure 36. Fig. 43b depicts a reaction scheme  
10 for the synthesis of NPS 467.

Fig. 44 depicts a dose response curve showing that NPS 467 lowers serum ionized calcium when administered orally.

#### Calcimimetic and Calcilytic Molecules

15 Calcimimetic and calcilytic molecules useful in the invention are generally described above. These molecules can be readily identified using screening procedures to define molecules which mimic or antagonize the activity of  $\text{Ca}^{2+}$  at  $\text{Ca}^{2+}$  receptors. Examples of such procedures are  
20 provided below. These examples are not limiting in the invention but merely illustrate methods which are readily used or adapted by those skilled in the art.

Generally, calcimimetic and calcilytic molecules are identified by screening molecules which are modelled after  
25 those described below (called lead molecules). As can be seen below there are several specific calcimimetics and calcilytics useful at various  $\text{Ca}^{2+}$  receptors. Derivative molecules are readily designed by standard procedures and tested in one of many protocols known to those skilled in  
30 the art. Many molecules may be screened easily to identify the most useful in this invention.

Organic cationic molecules which mimic or antagonize the actions of  $\text{Ca}^{2+}$  in other systems contain the requisite structure for activity on a  $\text{Ca}^{2+}$  receptor. Rational design  
35 of other useful molecules involves the study of a molecule known to be calcimimetic or calcilytic and then modifying

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the structure of the known molecule. For example, polyamines are potentially calcimimetic since spermine mimics the action of  $\text{Ca}^{2+}$  in several in vitro systems. Results show that spermine does indeed cause changes in  $[\text{Ca}^{2+}]_i$  and PTH secretion reminiscent of those elicited by extracellular di- and trivalent cations (see below). The experiments outlined below are therefore aimed at demonstrating that this phenomenology, obtained with spermine, involves the same mechanisms used by extracellular  $\text{Ca}^{2+}$ . To do this, the effects of spermine on a variety of physiological and biochemical parameters which characterize activation of the  $\text{Ca}^{2+}$  receptor were assessed. Those molecules having similar effects are useful in this invention and can be discovered by selecting or making molecules having a structure similar to spermine. Once another useful molecule is discovered this selection process can be readily repeated.

For clarity, below is provided a specific series of screening protocols to identify such useful molecules which are active at a parathyroid cell  $\text{Ca}^{2+}$  receptor, or which act as agonists or antagonists of the cellular response to changes in  $[\text{Ca}^{2+}]$ . Equivalent assays can be used for molecules active at other  $\text{Ca}^{2+}$  receptors, or which otherwise mimic or antagonize cellular functions regulated by  $[\text{Ca}^{2+}]$ . These assays exemplify the procedures which are useful to find calcimimetic molecules of this invention. Equivalent procedures can be used to find calcilytic molecules by screening for those molecules most antagonistic to the actions of extracellular  $\text{Ca}^{2+}$ . In vitro assays can be used to characterize the selectivity, saturability, and reversibility of these calcimimetics and calcilytics by standard techniques.

#### Screening Procedure

Generally, bovine parathyroid cells loaded with fura-2 are initially suspended in buffer containing 0.5 mM  $\text{CaCl}_2$ . The test substance is added to the cuvette in a

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small volume (5-15  $\mu$ l) and any change in the fluorescence signal noted. Cumulative increases in the concentration of the test substance are made in the cuvette until some predetermined concentration is achieved or changes in  
5 fluorescence noted. If no changes in fluorescence are noted, the molecule is considered inactive and no further testing is performed. In initial studies, e.g., with polyamine-type molecules, molecules were tested at concentrations as high as 5 or 10 mM. As more potent mole-  
10 cules are now known (see below), the ceiling concentration is lowered. For example, newer molecules are tested at concentrations up to 500  $\mu$ M or less. If no changes in fluorescence are noted at this concentration, the molecule can be considered inactive.

15 Molecules causing increases in  $[Ca^{2+}]_i$  are subjected to additional testing. The two essential characteristics of the molecule important for its consideration as a calcimimetic molecule are the mobilization of intracellular  $Ca^{2+}$  and sensitivity to PKC activators. Molecules causing the  
20 mobilization of intracellular  $Ca^{2+}$  in a PMA-sensitive manner have invariably been found to be calcimimetic molecules and to inhibit PTH secretion. Additional testing can, if needed, be performed to solidify this belief. Typically, all the various tests for calcimimetic or  
25 calcilytic activity (see above) are not performed. Rather, if a molecule causes the mobilization of intracellular  $Ca^{2+}$  in a PMA-sensitive manner, it is advanced to screening on human parathyroid cells. For example, measurements of  $[Ca^{2+}]_i$  are performed to determine the  $EC_{50}$ , and  
30 to measure the ability of the molecule to inhibit PTH secretion in human parathyroid cells which have been obtained from patients undergoing surgery for primary or secondary hyperparathyroidism. The lower the  $EC_{50}$  or  $IC_{50}$  the more potent the molecule as a calcimimetic or  
35 calcilytic.

Measuring  $[Ca^{2+}]_i$  with fura-2 provides a very rapid means of screening new organic molecules for activity. In

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a single afternoon, 10-15 molecules can be examined and their ability to mobilize intracellular  $\text{Ca}^{2+}$  (or not) assessed. The sensitivity of any observed increase in  $[\text{Ca}^{2+}]_i$  to depression by PMA can also be assessed. Moreover, a single cell preparation can provide data on  $[\text{Ca}^{2+}]_i$ , cyclic AMP levels,  $\text{IP}_3$  and PTH secretion. A typical procedure is to load cells with fura-2 and then split the cell suspension in two; most of the cells are used for measurement of  $[\text{Ca}^{2+}]_i$  and the remainder are incubated with molecules to assess their effects on cyclic AMP and PTH secretion. Because of the sensitivity of the radioimmunoassays for cyclic AMP and PTH, both variables can be determined in a single incubation tube containing 0.3 ml cell suspension (about 500,000 cells). Measurements of inositol phosphates are a time-consuming aspect of the screening. However, ion exchange columns eluted with chloride (rather than formate) provide a very rapid means of screening for  $\text{IP}_3$  formation since rotary evaporation (which takes around 30 hrs) is not required. This method allows processing of nearly 100 samples in a single afternoon. Those molecules that prove interesting, as assessed by measurements of  $[\text{Ca}^{2+}]_i$ , cyclic AMP,  $\text{IP}_3$ , and PTH are then subjected to a more rigorous analysis by examining formation of various inositol phosphates and assessing their isomeric form by HPLC.

Interesting molecules detected in these protocols are then assessed for specificity, e.g., by examining their effects on  $[\text{Ca}^{2+}]_i$  in calcitonin-secreting C-cells using, e.g., the rat MTC 6-23 cell line.

The following is illustrative of methods useful in these screening procedures. Examples of typical results for various test calcimimetic or calcilytic molecules are provided in Figs. 2-34.

#### Parathyroid Cell Preparation

Parathyroid glands were obtained from freshly slaughtered calves (12-15 weeks old) at a local abattoir and

transported to the laboratory in ice-cold parathyroid cell buffer (PCB) which contains (mM): NaCl, 126; KCl, 4; MgCl<sub>2</sub>, 1; Na-HEPES, 20; pH 7.4; glucose, 5.6, and variable amounts of CaCl<sub>2</sub>, e.g., 1.25 mM. Human parathyroid glands, obtained from patients undergoing surgical removal of parathyroid tissue for primary or uremic hyperparathyroidism (HPT), were treated similarly to bovine tissue. Glands were trimmed of excess fat and connective tissue and then minced with a fine scissors into approximate cubes of 2-3 mm. Dissociated parathyroid cells were prepared by collagenase digestion. Dissociated cells were then purified by centrifugation in Percoll buffer. The resultant parathyroid cell preparation was essentially devoid of red blood cells, adipocytes, and capillary tissue as assessed by phase contrast microscopy and Sudan black B staining. Dissociated and purified parathyroid cells were present as small clusters containing 5 to 20 cells. Cellular viability, as indexed by exclusion of trypan blue or ethidium bromide, was routinely 95%.

Although cells can be used for experimental purposes at this point, physiological responses (suppressibility of PTH secretion and resting levels of [Ca<sup>2+</sup>]<sub>i</sub>) are better after culturing the cells overnight. Primary culture also has the advantage that cells can be labeled with isotopes to near isotopic equilibrium, as is necessary for studies involving measurements of inositol phosphate metabolism (see below). After purification on Percoll gradients, cells were washed several times in a 1:1 mixture of Ham's F12-Dulbecco's modified Eagles medium (GIBCO) supplemented with 50 ug/ml streptomycin, 100 U/ml penicillin, 5 ug/ml gentamicin and ITS<sup>+</sup>. ITS<sup>+</sup> is a premixed solution containing insulin, transferrin, selenium, and bovine serum albumin (BSA)-linolenic acid (Collaborative Research, Bedford, MA). The cells were then transferred to plastic flasks (75 or 150 cm<sup>2</sup>; Falcon) and incubated at 37°C in a humid atmosphere of 5% CO<sub>2</sub>. No serum is added to these overnight cultures, since its presence allows the cells to attach to

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the plastic, undergo proliferation, and dedifferentiate. Cells cultured under the above conditions were readily removed from the flasks by decanting, and show the same viability as freshly prepared cells.

5        Measurement of Cytosolic  $\text{Ca}^{2+}$

Purified parathyroid cells were resuspended in 1.25 mM  $\text{CaCl}_2$ -2% BSA-PCB containing 1  $\mu\text{M}$  fura-2-acetoxymethylester and incubated at 37°C for 20 min. The cells were then pelleted, resuspended in the same buffer lacking the  
10 ester, and incubated a further 15 min at 37°C. The cells were subsequently washed twice with PCB containing 0.5 mM  $\text{CaCl}_2$  and 0.5% BSA and maintained at room temperature (about 20°C). Immediately before use, the cells were diluted five-fold with prewarmed 0.5 mM  $\text{CaCl}_2$ -PCB to obtain  
15 a final BSA concentration of 0.1%. The concentration of cells in the cuvette used for fluorescence recording was  $1-2 \times 10^6/\text{ml}$ .

The fluorescence of indicator-loaded cells was measured at 37°C in a spectrofluorimeter (Biomedical Instrumentation Group, University of Pennsylvania, Philadelphia, PA) equipped with a thermostated cuvette holder and magnetic stirrer using excitation and emission wavelengths of 340 and 510 nm, respectively. This fluorescence indicates the level of cytosolic  $\text{Ca}^{2+}$ . Fluorescence signals were  
25 calibrated using digitonin (50  $\mu\text{g}/\text{ml}$ , final) to obtain maximum fluorescence ( $F_{\text{max}}$ ), and EGTA (10 mM, pH 8.3, final) to obtain minimal fluorescence ( $F_{\text{min}}$ ), and a dissociation constant of 224 nM. Leakage of dye is dependent on temperature and most occurs within the first 2 min after  
30 warming the cells in the cuvette; dye leakage increases only very slowly thereafter. To correct the calibration for dye leakage, cells were placed in the cuvette and stirred at 37°C for 2-3 min. The cell suspension was then removed, the cells pelleted, and the supernatant returned  
35 to a clean cuvette. The supernatant was then treated with digitonin and EGTA as above to obtain as estimate of dye

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leakage, which is typically 10-15% of the total  $\text{Ca}^{2+}$ -dependent fluorescent signal. This estimate was subtracted from the apparent  $F_{\min}$ .

#### Measurement of PTH Secretion

5 In most experiments, cells loaded with fura-2 were used in studies of PTH secretion. Loading parathyroid cells with fura-2 does not change their secretory response to extracellular  $\text{Ca}^{2+}$ . Cells were suspended in PCB containing 0.5 mM  $\text{CaCl}_2$  and 0.1% BSA. Incubations were performed in plastic tubes (Falcon 2058) containing 0.3 ml of the cell suspension with or without small volumes of  $\text{CaCl}_2$  and/or organic polycations. After incubation at 37°C for various times (typically 30 min), the tubes were placed on ice and the cells pelleted at 2°C. Samples of the supernatant were brought to pH 4.5 with acetic acid and stored at -70°C. This protocol was used for both bovine and human parathyroid cells.

For bovine cells, the amount of PTH in sample supernatants was determined by a homologous radioimmunoassay using GW-1 antibody or its equivalent at a final dilution of 1/45,000.  $^{125}\text{I}$ -PTH (65-84; INCSTAR, Stillwater, MN) was used as tracer and fractions separated by dextran-activated charcoal. Counting of samples and data reduction were performed on a Packard Cobra 5005 gamma counter.

25 For human cells, a commercially available radioimmunoassay kit (INS-PTH; Nichols Institute, Los Angeles, CA) which recognizes intact and N-terminal human PTH was used because GW-1 antibody recognizes human PTH poorly.

#### Measurement of cyclic AMP

30 Cells were incubated as above for PTH secretion studies and at the end of incubation, a 0.15 ml sample was taken and transferred to 0.85 ml hot (70°C) water and heated at this temperature for 5-10 min. The tubes were subsequently frozen and thawed several times and the cellular debris sedimented by centrifugation. Portions of

the supernatant were acetylated and cyclic AMP concentrations determined by radioimmunoassay.

#### Measurement of Inositol Phosphate Formation

Membrane phospholipids were labeled by incubating  
5 parathyroid cells with 4  $\mu\text{Ci/ml}$   $^3\text{H}$ -myo-inositol for 20-24  
hrs. Cells were then washed and resuspended in PCB con-  
taining 0.5 mM  $\text{CaCl}_2$  and 0.1% BSA. Incubations were per-  
formed in microfuge tubes in the absence or presence of  
various concentrations of organic polycation for different  
10 times. Reactions were terminated by the addition of 1 ml  
chloroform/methanol/12 N HCl (200:100:1; v/v/v). Phytic  
acid hydrolysate (200  $\mu\text{l}$ ; 25  $\mu\text{g}$  phosphate/tube) water was  
then added. The tubes were centrifuged and 600  $\mu\text{l}$  of the  
aqueous phase was diluted into 10 ml water.

15 Inositol phosphates were separated by ion exchange  
chromatography using AG1-X8 in either the chloride- or  
formate-form. When only  $\text{IP}_3$  levels were to be determined,  
the chloride-form was used, whereas the formate form was  
used to resolve the major inositol phosphates ( $\text{IP}_3$ ,  $\text{IP}_2$ ,  
20 and  $\text{IP}_1$ ). For determination of just  $\text{IP}_3$ , the diluted  
sample was applied to the chloride-form column and the  
column washed with 10 ml 30 mM HCl followed by 6 ml 90 mM  
HCl and the  $\text{IP}_3$  eluted with 3 ml 500 mM HCl. The last  
eluate was diluted and counted. For determination of all  
25 major inositol phosphates, the diluted sample was applied  
to the formate-form column and  $\text{IP}_1$ ,  $\text{IP}_2$ , and  $\text{IP}_3$  eluted  
sequentially by increasing concentrations of formate  
buffer. The eluted samples from the formate columns were  
rotary evaporated, the residues brought up in cocktail,  
30 and counted.

The isomeric forms of  $\text{IP}_3$  were evaluated by HPLC. The  
reactions were terminated by the addition of 1 ml 0.45 M  
perchloric acid and stored on ice for 10 min. Following  
centrifugation, the supernatant was adjusted to pH 7-8  
35 with  $\text{NaHCO}_3$ . The extract was then applied to a Partisil  
SAX anion-exchange column and eluted with a linear gra-

dient of ammonium formate. The various fractions were then desalted with Dowex followed by rotary evaporation prior to liquid scintillation counting in a Packard Tri-carb 1500 LSC.

- 5 For all inositol phosphate separation methods, appropriate controls using authentic standards were used to determine if organic polycations interfered with the separation. If so, the samples were treated with cation-exchange resin to remove the offending molecule prior to  
10 separation of inositol phosphates.

#### Measurement of Cytosolic $\text{Ca}^{2+}$ in C-cells

- Neoplastic C-cells derived from a rat medullary thyroid carcinoma (rMTC 6-23) obtained from American Type Culture Collection (ATCC No. 1607) were cultured as mono-  
15 layers in Dulbecco's Modified Eagle's medium (DMEM) plus 15% horse serum in the absence of antibiotics. For measurements of  $[\text{Ca}^{2+}]_i$ , the cells were harvested with 0.02% EDTA/0.05% trypsin, washed twice with PCB containing 1.25 mM  $\text{CaCl}_2$  and 0.5% BSA, and loaded with fura-2 as described  
20 above for parathyroid cells. Measurements of  $[\text{Ca}^{2+}]_i$  were performed as described above with appropriate corrections for dye leakage.

#### Measurement of $[\text{Ca}^{2+}]_i$ in Rat Osteoclasts

- Osteoclasts were obtained from 1-2 day old Sprague-  
25 Dawley rats using aseptic conditions. The rat pups were sacrificed by decapitation, the hind legs removed, and the femora rapidly freed of soft tissue and placed in pre-warmed F-12/DMEM media (DMEM containing 10% fetal calf serum and antibiotics (penicillin-streptomycin-gentamicin;  
30 100 U/ml-100 ug/ml-100 ug/ml)). The bones from two pups were cut lengthwise and placed in 1 ml culture medium. Bone cells were obtained by gentle trituration of the bone fragments with a plastic pipet and diluted with culture medium. The bone fragments were allowed to settle and  
35 equal portions (about 1 ml) of the medium transferred to

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a 6 well culture plate containing 25 mm glass coverslips. The cells were allowed to settle for 1 hr at 37°C in a humidified 5% CO<sub>2</sub>-air atmosphere. The coverslips were then washed 3 times with fresh media to remove nonadherent  
5 cells. Measurements of [Ca<sup>2+</sup>]<sub>i</sub> in osteoclasts were performed within 6-8 hrs of removing nonadherent cells.

Cells attached to the coverslip were loaded with indo-1 by incubation with 5 μM indo-1 acetoxymethylester / 0.01% Pluronic F28 for 30 min at 37°C in F-12/DMEM  
10 lacking serum and containing instead 0.5% BSA. The coverslips were subsequently washed and incubated an additional 15 min at 37°C in F-12/DMEM lacking ester before being transferred to a superfusion chamber mounted on the stage of a Nikon Diaphot inverted microscope equipped for micro-  
15 fluorimetry. Osteoclasts were easily identified by their large size and presence of multiple nuclei. The cells were superfused with buffer (typically PCB containing 0.1% BSA and 1 mM Ca<sup>2+</sup>) at 1 ml/min with or without test substance. The fluorescence emitted by excitation at 340 nm  
20 was directed through the video port of the microscope onto a 440 nm dichroic mirror and fluorescence intensity at 495 and 405 nm collected by photomultiplier tubes. The outputs from the photomultiplier tubes were amplified, digitized, and stored in an 80386 PC. Ratios of fluorescence  
25 intensity were used to estimate [Ca<sup>2+</sup>]<sub>i</sub>.

#### Oocyte Expression

In additional studies, Xenopus oocytes injected with mRNA from bovine or human parathyroid cells were used in screening protocols, and Cl<sup>-</sup> conductance measured as an  
30 indirect means of monitoring increases in [Ca<sup>2+</sup>]<sub>i</sub>. The following is an example to test the effect of neomycin.

Oocytes were injected with poly(A)<sup>+</sup>-enriched mRNA from human parathyroid tissue (hyperplastic glands from a case of secondary HPT). After 3 days, the oocytes were tested  
35 for their response to neomycin. Neomycin B evoked oscillatory increases in the Cl<sup>-</sup> conductance which ceased upon

superfusion with drug-free saline (see Fig. 20). Responses to neomycin B were observed at concentrations between 100  $\mu$ M and 10 mM. To ensure that the response evoked by neomycin B was contingent upon injection of parathyroid mRNA, the effect of neomycin B on currents in water-injected oocytes was determined. In each of five oocytes examined, neomycin B (10 mM) failed to cause any change in the current. About 40% of oocytes are known to respond to carbachol, an effect mediated by an endogenous muscarinic receptor. In five oocytes examined, one showed inward currents in response to carbachol, and this is shown in the lower trace of Fig. 20. Thus, in cells expressing a muscarinic receptor coupled to increases in  $[Ca^{2+}]_i$  and  $Cl^-$  conductance, neomycin B fails to evoke a response. This shows that the response to neomycin B depends on expression of a specific protein encoded by parathyroid cell mRNA. It suggests quite strongly that in intact cells, neomycin B acts directly on the  $Ca^{2+}$  receptor to alter parathyroid cell function.

## 20 Drug Design From Lead Molecules

Certain organic molecules mimic or antagonize the action of extracellular  $Ca^{2+}$  by acting at the  $Ca^{2+}$  receptor as shown herein. The molecules tested, however, are not necessarily suitable as drug candidates, but they serve to demonstrate that the hypothesis underlying  $Ca^{2+}$  receptor-based therapies is correct. These molecules can be used to determine the structural features that enable them to act on the  $Ca^{2+}$  receptor, and thus to select molecules useful in this invention.

30 An example of one such analytical procedure follows: This example is detailed in the examples below, but is used here to demonstrate the rationale that can be used to design useful molecules of this invention from lead molecules discussed herein. Those in the art will recognize the analytic steps defined in the example and that analogous analysis can be conducted on other lead molecules

until the most desired calcimimetic or calcilytic is defined.

Other examples are also provided below. Together the data presented demonstrate that useful lead molecules will have aromatic groups which are preferably substituted at one or more positions, and may have branched or linear substituted or unsubstituted alkyl groups as desired. In addition, it is important to choose molecules of correct stereospecificity to ensure higher affinity for the desired  $\text{Ca}^{2+}$  receptor. These data thus point those in the art to appropriate lead molecules which can be derivatised to find optimum desired molecules of this invention, much as described below.

Although structurally diverse, molecules that are tested may have common features that can be studied. In this example, the correlation between net positive charge and potency in mobilizing intracellular  $\text{Ca}^{2+}$  was tested. Protamine (+21;  $\text{EC}_{50} = 40 \text{ nM}$ ) was more effective than neomycin B (+6;  $\text{EC}_{50} = 20 \text{ }\mu\text{M}$  in human parathyroid cells and 40  $\mu\text{M}$  in bovine parathyroid cells) which was more effective than spermine (+4;  $\text{EC}_{50} = 150 \text{ }\mu\text{M}$ ) in causing the mobilization of  $[\text{Ca}^{2+}]_i$  in parathyroid cells. These results raise the question of whether positive charge alone determines potency, or if there are other structural features that contribute to activity on the  $\text{Ca}^{2+}$  receptor. This is important to determine at the outset because it profoundly impacts on the view that the  $\text{Ca}^{2+}$  receptor can be targeted with effective and specific therapeutic molecules. Thus, a variety of other organic polycations related to neomycin B and spermine can be studied to determine the relationship between the net positive charge of a molecule and its potency to mobilize intracellular  $\text{Ca}^{2+}$ .

The first series of molecules studied were the aminoglycosides. The molecules were examined on bovine parathyroid cells and their  $\text{EC}_{50}$ 's for the mobilization of intracellular  $\text{Ca}^{2+}$  determined. For the aminoglycosides, the rank order of potency for eliciting cytosolic  $\text{Ca}^{2+}$

transients was neomycin B ( $EC_{50} = 20$  or  $40 \mu M$ ) > gentamicin ( $150 \mu M$ ) > bekanamycin ( $200 \mu M$ ) > streptomycin ( $600 \mu M$ ). Kanamycin and lincomycin were without effect when tested at a concentration of  $500 \mu M$ . The net positive charge on these aminoglycosides at pH 7.3 is neomycin B (+6) > gentamicin (+5) = bekanamycin (+5) > kanamycin (average +4.5) > streptomycin (+3) > lincomycin (+1). Within the aminoglycoside series, then, there is some correlation between net positive charge but it is not absolute, and kanamycin, which would be predicted to be more potent than streptomycin, is without activity.

Testing of various polyamines revealed additional and more marked discrepancies between net positive charge and potency. Three structural classes of polyamines were examined: (1) straight chain, (2) branched chain, and (3) cyclic. The structures of the polyamines tested are provided in Fig. 1. Amongst the straight chain polyamines, spermine (+4;  $EC_{50} = 150 \mu M$ ) was more potent than pentaethylenehexamine (+6;  $EC_{50} = 500 \mu M$ ) and tetraethylenepentamine (+5;  $EC_{50} = 2.5 mM$ ) even though the latter molecules have a greater net positive charge.

We synthesized some branched chain polyamines that have different numbers of secondary and primary amino groups and thus vary in net positive charge. Two of these molecules, NPS 381 and NPS 382, were examined for effects on  $[Ca^{2+}]_i$  in bovine parathyroid cells. NPS 382 (+8;  $EC_{50} = 50 \mu M$ ) was about twice as potent as NPS 381 (+10;  $EC_{50} = 100 \mu M$ ) even though it contains two fewer positive charges.

A similar discrepancy between positive charge and potency was noted in experiments with cyclic polyamines. For example, hexacyclen (+6;  $EC_{50} = 20 \mu M$ ) was more potent than NPS 383 (+8;  $EC_{50} = 150 \mu M$ ). The results obtained with these polyamines show that positive charge is not the sole factor contributing to potency.

Additional studies provided insights into the structural features of molecules that impart activity on the



parathyroid cell  $\text{Ca}^{2+}$  receptor. One of the structurally important features is the intramolecular distance between the nitrogens (which carry the positive charge). Thus, spermine is 50-fold more potent than triethylenetetramine (EC<sub>50</sub> = 8 mM) in evoking increases in  $[\text{Ca}^{2+}]_i$  in bovine parathyroid cells yet both molecules carry a net positive charge of +4. The only difference in structure between these two polyamines is the number of methylenes separating the nitrogens: in spermine it is 3-4-3 whereas in triethylenetetramine it is 2-2-2. This seemingly minor change in the spacing between nitrogens has profound implications for potency and suggests that the conformational relationships of nitrogens within the molecule are critical. Supporting this are results obtained with hexacyclen and pentaethylenhexamine. The former molecule is simply the cyclic analog of the latter and contains the same number of methylenes between all nitrogens, yet the presence of the ring structure increases potency 25-fold. These results indicate that positive charge per se is not the critical factor determining the activity of an organic molecule on the  $\text{Ca}^{2+}$  receptor.

Another series of experiments reveals the importance of aromatic groups in determining activity on the  $\text{Ca}^{2+}$  receptor. The results were obtained with two arylalkylamines isolated from the venom of the spider Argiope lobata. These molecules, argiotoxin 636 and argiotoxin 659, have identical polycationic portions linked to different aromatic groups (Fig. 24). Argiotoxin 659 evoked transient increases in  $[\text{Ca}^{2+}]_i$  in bovine parathyroid cells when tested at concentrations of 100 to 300  $\mu\text{M}$ . In contrast, argiotoxin 636 was without effect when tested at similar concentrations (Fig. 24). The only difference in structure between these two arylalkylamines is in the aromatic portion of the molecules: argiotoxin 659 contains a 4-hydroxyindole moiety whereas argiotoxin 636 contains a 2,4-dihydroxyphenyl group. The net positive charge on these two arylalkylamines is the same (+4), so their dif-

ferent potencies must result from the different aromatic groups. This shows that net positive charge alone does not determine potency. The real importance of these findings, however, is the discovery that aromatic groups contribute significantly to the ability of molecules to activate the  $\text{Ca}^{2+}$  receptor.

Agatoxin 489 (NPS 017) and Agatoxin 505 (NPS 015) both cause the mobilization of intracellular  $\text{Ca}^{2+}$  in parathyroid cells with  $\text{EC}_{50}$ 's of 6 and 22  $\mu\text{M}$ , respectively. The only difference in the structure of these molecules is a hydroxyl group on the indole moiety (Fig. 1). This shows that substitutions on the aromatic region of the molecule can influence potency. This indicates that further lead molecules to be studied will include those molecules having substituted aromatic moieties.

The structural features to be varied systematically from lead molecules described herein include (1) net positive charge, (2) number of methylenes separating nitrogens, and (3) cyclic versions of, e.g., polyamines, with and without changes in methylene spacing and net positive charge. In addition systematic variations in the structure and location of aromatic groups can be examined, e.g., in a variety of arylalkylamines isolated from the venoms of wasps and spiders; and synthetic molecules can be prepared by the coupling of commercially available aromatic moieties to the argiotoxin polyamine moiety. The argiotoxin polyamine moiety can be readily coupled to any aromatic moiety containing a carboxylic acid. Thus, it is simple to systematically screen the hydroxy and methoxy derivatives of phenylacetic acid and benzoic acid as well as the hydroxyindoleacetic acid series. Analogs containing heteroaromatic functionalities can also be prepared and assessed for activity.

Comparisons of potency and efficacy among such molecules will reveal the optimal structure and location of the aromatic group at a constant positive charge.

One of the structural variations on the polyamine motif that seems to increase potency is the presence of the cyclic version of the straight chain-parent molecule. Budmunchiamine A, isolated from the plant Albizia amara,  
5 is a cyclic derivative of spermine (Fig. 1). The addition of budmunchiamine A to bovine parathyroid cells caused a rapid and transient increase in  $[Ca^{2+}]_i$  that persisted in the absence of extracellular  $Ca^{2+}$  and was blunted by pre-treatment with PMA. It therefore causes the mobilization  
10 of intracellular  $Ca^{2+}$  in parathyroid cells, probably by acting on the  $Ca^{2+}$  receptor. It is about equipotent with spermine ( $EC_{50}$  about 200  $\mu M$ ) yet carries one less positive charge (+3) than does spermine.

The results obtained with budmunchiamine A demonstrate the predictive power of the structure-activity  
15 studies and the novel structural information to be gained by testing natural products. Thus, screening of natural products, selected rationally on the basis of the structural information is readily performed e.g., molecules can  
20 be selected on the basis of well-established chemotaxonomic principles using appropriate data bases, such as Napralert. For example, macrocyclic polyamine alkaloids derived from papilionoid legumes related to Albizia, such as Pithecolobium and other plant-derived molecules can be  
25 screened.

Fig. 36 provides a second example of a series of molecules which were screened to determine useful molecules of this invention. These molecules were generally derived from fendiline and tested to determine their  
30 respective  $EC_{50}$ 's. Moreover, testing of related molecules, such as NPS 447 and NPS 448 reveals stereospecific effects of molecule structure. The most active compounds tested to date are the novel compounds designated NPS 467 and NPS 568 which have  $EC_{50}$  values of less than 5  $\mu M$ . Those in the  
35 art, by reviewing this series of molecules, can determine other suitable derivatives which can be tested in the invention.

These examples demonstrate the general design and screening process useful in this invention, and indicate that additional compound and natural product libraries can be screened as desired by those in the art to determine  
5 other useful lead molecules or novel molecules of this invention.

As discussed above, examples of molecules useful as calcimimetics include branched or cyclic polyamines, positively charged polyamino acids, and arylalkylamines. In  
10 addition, other positively charged organic molecules, including naturally occurring molecules and their analogs, are useful calcimimetics. These naturally occurring molecules and their analogs preferably have positive charge-to-mass ratios that correlate with those ratios for the  
15 molecules exemplified herein. (Examples include material isolated from marine animals, arthropod venoms, terrestrial plants and fermentation broths derived from bacteria and fungi.) It is contemplated that one group of preferred naturally occurring molecules and analogs useful as  
20 calcimimetics will have a ratio of positive charge: molecular weight (in daltons) from about 1:40 to 1:200, preferably from about 1:40 to 1:100. More specific examples of such molecules are provided below.

#### Polyamines

25 The polyamines useful as calcimimetics in this invention may be either branched or cyclic. Branched or cyclic polyamines potentially have higher calcimimetic activity than their straight-chain analogs. That is, branched or cyclic polyamines tend to have a lower  $EC_{50}$  than their  
30 corresponding linear polyamines with the same effective charge at physiological pH (see Table 1).

Table 1

	Molecule	Net (+) Charge	EC <sub>50</sub> ( $\mu$ M)
	Neomycin	+6	20 or 40
5	Hexacyclen	+6	20
	NPS 382	+8	50
	NPS 381	+10	100
	NPS 383	+8	150
	Gentamicin	+5	150
10	Spermine	+4	150
	Bekanamycin	+5	200
	Argiotoxin-659	+4	300
	Pentaethylenhexamine (PEHA)	+6	500
	Streptomycin	+3	600
15	Spermidine	+3	2000
	Tetraethylenepentamine (TEPA)	+5	2500
	1,12-diaminododecane (DADD)	+2	3000
	Triethylenetramine (TETA)	+4	8000

"Branched polyamines" as used herein refers to a chain molecule consisting of short alkyl bridges or alkyl groups joined together by amino linkages, and also containing points at which the chain branches. These "branch points" can be located at either a carbon atom or a nitrogen atom, preferably at a nitrogen atom. A nitrogen atom branch point is typically a tertiary amine but it may also be quaternary. A branched polyamine may have 1 to 20 branch points, preferably 1 to 10 branch points.

Generally, the alkyl bridges and alkyl branches in a branched polyamine are from 1 to 50 carbon atoms in length, preferably from 2 to 6 carbon atoms. The alkyl branches may also be interrupted by one or more heteroatoms (nitrogen, oxygen or sulfur) or substituted with functional groups such as: halo, including fluoro, chloro, bromo, or iodo; hydroxy; nitro; acyloxy (R'COO-), acylamido (R'CONH-), or alkoxy (-OR'), where R' may contain from 1 to 4 carbon atoms. The alkyl branches may

also be substituted with groups that are positively charged at physiological pH, such as amino or guanido. These functional substituents may add or change physical properties such as solubility to increase activity, delivery or bioavailability of the molecules.

The branched polyamines may have three or more chain and branch termination points. These termination points may be methyl groups or amino groups, preferably amino groups.

One preferred group of molecules is the group of branched polyamines having the formula:



where k is an integer from 1 to 10, each j is the same or different and is an integer from 2 to 20, and each R<sub>i</sub> is the same or different and is selected from the group consisting of hydrogen and  $-(\text{CH}_2)_j-\text{NH}_2$ , where j is as defined above, and at least one R<sub>i</sub> is not hydrogen.

Particularly preferred branched polyamines of this invention are the molecules N<sup>1</sup>,N<sup>1</sup>,N<sup>5</sup>,N<sup>10</sup>,N<sup>14</sup>,N<sup>14</sup>-hexakis-(3-aminopropyl)spermine and N<sup>1</sup>,N<sup>1</sup>,N<sup>5</sup>,N<sup>14</sup>,N<sup>14</sup>-tetrakis-(3-aminopropyl)spermine referred to as NPS 381 and NPS 382, respectively, in Figure 1.

"Cyclic polyamines" as used herein refer to heterocycles containing two or more heteroatoms (nitrogen, oxygen or sulfur), at least two of which are nitrogen atoms. The heterocycles are generally from about 6 to about 20 atoms in circumference, preferably from about 10 to about 18 atoms in circumference. The nitrogen heteroatoms are separated by 2 to 10 carbon atoms. The heterocycles may also be substituted at the nitrogen sites with aminoalkyl or aminoaryl groups (NH<sub>2</sub>R-), wherein R is aminoaryl or a lower alkyl of 2 to 6 carbon atoms.

Particularly preferred cyclic polyamines of this invention are shown in Figure 1 as hexacyclen (1,4,7,10,13,16-hexaaza-cyclooctadecane) and NPS 383.

### Polyamino Acids

The polyamino acids useful in this invention may contain two or more positively charged amino acid residues at physiological pH. These positively charged amino acids include histidine, lysine and arginine. These polypeptides will vary in length from 2 to 800 amino acids in length, more preferably from 20 to 300 amino acids in length. These polypeptides may consist of a single repeating amino acid residue, or may have the variety of a naturally occurring protein or enzyme.

The amino acid residues comprising the polyamino acids may be any of the twenty naturally occurring amino acids, or other alternative residues. Alternative residues include, for example, the  $\omega$ -amino acids of the formula  $H_2N(CH_2)_nCOOH$ , where  $n$  is from 2 to 6. These are neutral, nonpolar amino acids, as are sarcosine, t-butyl alanine, t-butyl glycine, N-methyl isoleucine, norleucine, phenyl glycine, citrulline, methionine sulfoxide, cyclohexyl alanine, and hydroxyproline. Ornithine is an alternative positively charged amino acid residue. The polyamino acids of this invention may also be chemically derivatized by known methods.

Particularly preferred polyamino acids of this invention include polyarginine, polylysine, and poly(argininyl-tyrosine), having 20-300 residues. Another preferred polyamino acid is protamine, or a protamine analog.

### Arylalkylamines

"Arylalkylamines" as used herein refer to a class of positively-charged toxins derived from arthropod venoms. Preferred arylalkylamines of this invention include philanthotoxin-433, argiotoxin-636, and argiotoxin-659, agatoxin 505, agatoxin 489, the structures of which are shown in Figure 1, and other synthetic molecules modeled after these natural products, such as NPS 019.

### Additional Components

Compositions of this invention include not only at least one calcimimetic or calcilytic, but may also include certain additional components. These additional components include targeting components, labels, and other  
5 functionalities which may be useful in the applications herein, e.g., for screening for agonists or antagonists of extracellular  $\text{Ca}^{2+}$ .

For example, an immunoglobulin or portion thereof, or a ligand specific for parathyroid cells or a  $\text{Ca}^{2+}$  receptor  
10 can be used as a target-specific component. The immunoglobulin can be a polyclonal or monoclonal antibody and may comprise whole antibodies or immunologically reactive fragments of these antibodies such as  $\text{F}_{ab}$ ,  $\text{F}_b$ , or  $(\text{F}_{ab})_2$ . Receptor-specific ligands may also be used.

15 The compositions of this invention may also contain components derivatized with a molecule or ion which acts as a label. A wide variety of labeling moieties can be used, including radioisotopes, chromophores, and fluorescent labels. Radioisotope labeling in particular can be  
20 readily detected in vivo. Radioisotopes may be coupled by coordination as cations in the porphyrin system. Useful cations include technetium, gallium, and indium. In the compositions, the positively charged molecule can be linked to or associated with a label.

## 25 Methods of Synthesis

Strategies for the syntheses and the modification of polyamines involve the use of a variety of amine protecting groups (phthalimido, BOC, CBZ, benzyl, and nitrile) which can be removed selectively to construct functional-  
30 ized molecules. The synthetic methods involved are modelled after those used to construct argiopines 636 and 659 and other arylalkylamines derived from spider venoms.

Chain extensions of 2-4 methylenes were typically accomplished by alkylation with the corresponding  
35 N-(bromoalkyl)phthalimide. A 1:1.2 mixture of amine to the bromoalkylphthalimide was refluxed in acetonitrile in



the presence of 50% KF on Celite. Chain extensions were also accomplished by alkylation of a given amine with acrylonitrile or ethylacrylate. Reaction progress was monitored by TLC and intermediates purified on silica gel using combinations of dichloromethane, methanol, and isopropylamine. Final products were purified by cation exchange (HEMA-SB) and RP-HPLC (Vydac C-18). Purity and structure verification are accomplished by  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR and high-resolution mass spectrometry (EI, CI and/or FAB).

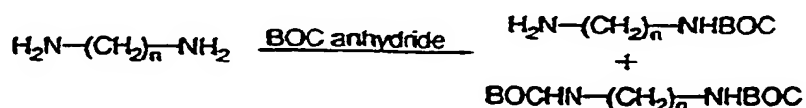
BOC protecting groups were added by the treatment of an amine ( $1^\circ$  or  $2^\circ$ ) with di-tert-butyl dicarbonate in dichloromethane in the presence of a catalytic amount of dimethylaminopyridine. Benzyl protecting groups were applied in one of two ways: (1) condensation of a  $1^\circ$  amine with benzaldehyde followed by sodium borohydride reduction or (2) alkylation of a  $2^\circ$  amine with benzylbromide in the presence of KF. Amide linkages and cyclizations were typically performed by the reaction of an amine ( $1^\circ$  or  $2^\circ$ ) with the N-hydroxysuccinimide ester of a given acid. This was accomplished directly (in the case of cyclizations) by treatment of the "amino acid" with dicyclohexylcarbodiimide under dilute conditions.

Deprotections of the phthalimido functionality were accomplished by reduction with hydrazine in refluxing methanol. Deprotections of the BOC functionality were accomplished in anhydrous TFA. Deprotection of benzyl, nitrile, and CBZ protecting functionalities was accomplished by reduction in glacial acetic acid under 55 psi hydrogen in the presence of a catalytic amount of palladium hydroxide on carbon. Nitrile functionalities (in the presence of benzyl and CBZ groups) were selectively reduced under hydrogen in the presence of sponge Raney nickel.

Specifically, branched polyamines are typically prepared from simple diaminoalkanes of the formula  $\text{NH}_2-(\text{CH}_2)_n-\text{NH}_2$ , or simple polyamines such as spermidine or spermine. One of the two primary (terminal) amines is

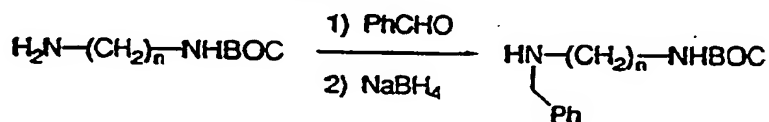
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protected or "masked" with a protecting group such as BOC (t-butyloxycarbonyl), phthalimido, benzyl, 2-ethylnitrile (the Michael condensation production product of an amine and acrylonitrile), or amide. A typical reaction is the  
 5 addition of a BOC protecting group by treatment with di-t-butyl-dicarbonate (BOC anhydride):

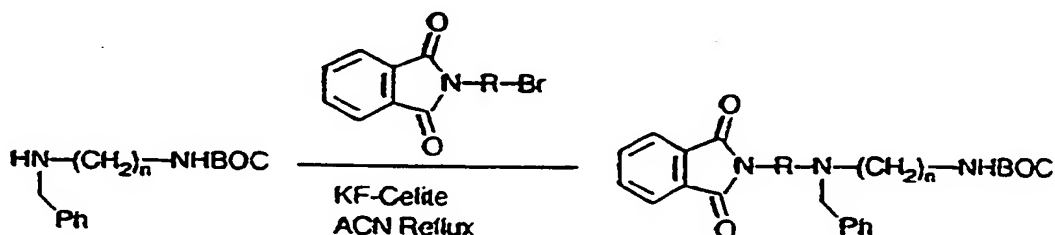


The monoprotected product is separated from the unprotected and diprotected products by simple chromatographic or distillation techniques.

10 The remaining free amine in the monoprotected product is then selectively alkylated (or acylated) with an alkylating (or acylating) agent. To ensure mono-alkylation, the free amine is partially protected by condensation with benzaldehyde followed by sodium borohydride reduction to  
 15 form the N-benzyl derivative:

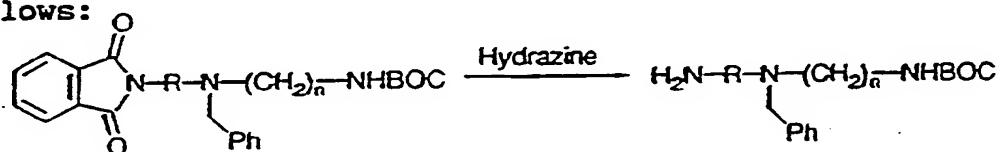


The N-benzyl derivative is then reacted with the alkylating agent. A typical alkylating agent is in an N-(bromo-alkyl)phthalimide, which reacts as follows:



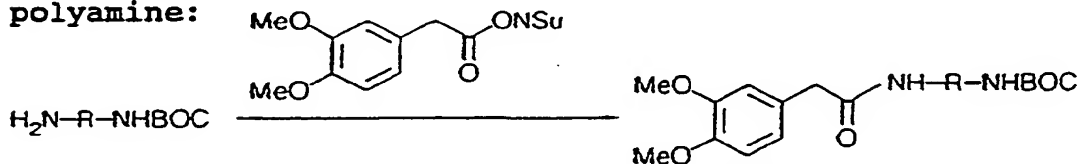
For example, N-(bromobutyl)phthalimide is used to extend  
 20 or branch the chain with four methylene units. Alternatively, reaction with acrylonitrile followed by reduction of the cyano group will extend the chain by three methylenes and an amino group.

The protecting groups of the resulting chain-extended molecule can then be selectively cleaved to yield a new free amine. For example, trifluoroacetic acid is used to remove a BOC group; catalytic hydrogenation is used to reduce a nitrile functionality and remove a benzyl group; and hydrazine is used to remove phthalimido groups as follows:



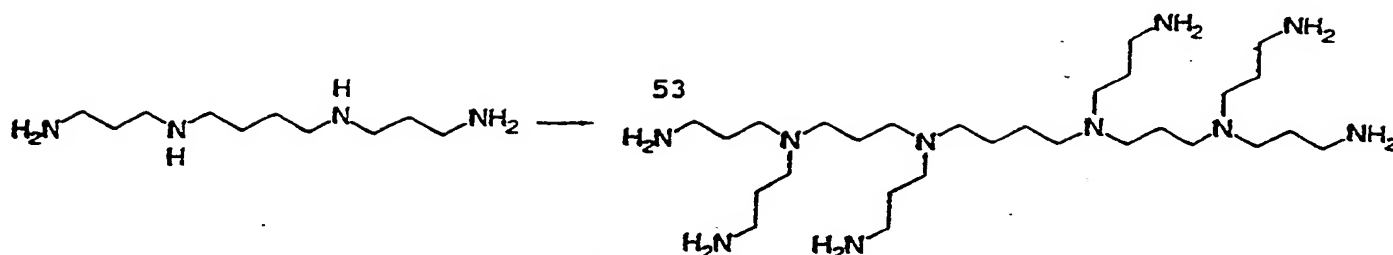
The new free amine may be alkylated (or acylated) further as above to increase the length of the polyamine. This process is repeated until the desired chain length and number of branches is obtained. In the final step, deprotection of the product results in the desired polyamine. However, further modifications may be effected at the protected end prior to deprotection in the following manner:

For example, prior to BOC-deprotection, the polyamine is acylated with the N-hydroxysuccinimide ester of 3,4-dimethoxyphenylacetic acid to yield a diprotected polyamine:



This ultimately will yield an aryl polyamine. The BOC group can then be selectively removed with trifluoroacetic acid to expose the other amino terminus which can be extended as above.

Certain branched polyamines may be formed by simultaneously alkylating or acylating the free primary and secondary amines in a polyamine formed as above. For example, treatment of spermine with excess acrylonitrile followed by catalytic reduction yields the following:



Cyclic polyamines may be prepared as above beginning with starting materials such as hexacylen (Aldrich Chem.).

The polyamino acids within the scope of the present invention can be made by recombinant techniques known in the art, or may be synthesized using standard solid-phase techniques known in the art. Solid-phase synthesis is commenced from the carboxy-terminal end of the peptide using an  $\alpha$ -amino protected amino acid. BOC protective groups can be used for all amino groups even through other protective groups are suitable. For example, BOC-lys-OH can be esterified to chloromethylated polystyrene resin supports. The polystyrene resin support is preferably a copolymer of styrene with about 0.5 to 2% divinylbenzene as a cross-linking agent which causes the polystyrene polymer to be completely insoluble in certain organic solvents. See Stewart et al., Solid-Phase Peptide Synthesis (1969), W.H. Freeman Co., San Francisco; and Merrifield, J. Am. Chem. Soc. (1963) 85:2149-2154. These and other methods of peptide synthesis are also exemplified by U.S. Patent Nos. 3,862,925; 3,842,067; 3,972,859; and 4,105,602.

The polypeptide synthesis may use manual techniques or automatically employing, for example, an Applied Biosystems 403A Peptide Synthesizer (Foster City, California) or a Biosearch SAM II automatic peptide synthesizer (Biosearch, Inc., San Rafael, California), following the instructions provided in the instruction manual supplied by the manufacturer.

The arylalkylamines of the invention are natural products isolated by known techniques, or synthesized as described in Jasys et al., Tetrahedron Lett. (1988) 29:6223-6226, and Nason et al., Tetrahedron Lett. (1989) 30:2337-2340.

One general protocol for preparation of fendiline (or fendiline analogs shown in Fig. 36) is as follows. In a 10 ml round bottom flask equipped with a magnetic stir bar and rubber septum, 1.0 mmole 3,3'-bisphenylpropylamine (or primary alkyl amine) in 2 ml ethanol was treated with 1.1 mmole phenol and 1.0 mmole acetophenone (or substituted acetophenone). To this was added 2.0 mmoles  $\text{MgSO}_4$  and 1.0 mmole  $\text{NaCNBH}_3$ . This was stirred under a nitrogen atmosphere at room temperature (about 20°C) for 24 hrs. The reaction was poured into 50 ml ether and washed 3 times with 1 N NaOH and once with brine. The ether layer was dried with anhydrous  $\text{K}_2\text{CO}_3$  and reduced in vacuo. The product was then purified by column chromatography or HPLC incorporating silica stationary phase with combinations of  $\text{CH}_2\text{Cl}_2$ -Methanol-isopropylamine (typically 3% Methanol and 0.1% isopropylamine in methylene chloride).

A preferred procedure for preparing fendiline or fendiline analogs (such as those depicted in Figure 36) uses titanium(IV) isopropoxide and was modified from methods described in J. Org. Chem. 55:2552 (1990). For the synthesis of NPS 544, titanium tetrachloride (method described in Tetrahedron Letters 31:5547 (1990)) was used in place of titanium(IV) isopropoxide. The reaction scheme is depicted in Figure 43a. In Figure 43a, R, R' and R'' depict hydrocarbyl groups. According to one embodiment, in a 4 ml vial, 1 mmole of amine (1) (typically a primary amine) and 1 mmole ketone or aldehyde (2) (generally acetophenone) are mixed, then treated with 1.25 mmoles titanium(IV) isopropoxide (3) and allowed to stand with occasional stirring at room temperature for about 30 minutes. Alternatively, a secondary amine may be used in place of (1). Note: some reactions will give heavy precipitates or solids which are warmed/heated (to their melting point) to allow for stirring/mixing several times over the course of the reaction. The reaction mixture is treated with 1 ml ethanol containing 1 mmole sodium cyanoborohydride (4) and the resulting mixture is then allowed

to stand at room temperature with occasional stirring for about 16 hours. After this time the reaction is quenched by the addition of about 500  $\mu$ l water. The reaction mixture is then diluted to about 4 ml total volume with ethyl ether and then centrifuged. The upper organic phase is removed and reduced on a rotovapor. The resulting product, (6), is partially purified by chromatography through a short column of silica (or alternatively by using preparative TLC on silica) using combination of dichloromethane:methanol:isopropylamine (typically 95:5:0.1), prior to purification by HPLC (normal phase using silica with dichloromethane:methanol:isopropylamine or reversed phase, C-18 with 0.1% TFA with acetonitrile or methanol).

If appropriate or desired, chiral resolution may be accomplished using methods such as those described in Example 21.

#### Formulation and Administration

As demonstrated herein, the molecules of the invention may be used to: (a) mimic or antagonize one or more of the effect of extracellular  $\text{Ca}^{2+}$ ; (b) affect the extracellular free  $\text{Ca}^{2+}$  level in an individual; and (c) treat diseases such as hyperparathyroidism, osteoporosis and hypertension. While the molecules have generally been shown to have an effect on parathyroid cells, they may also modulate the  $\text{Ca}^{2+}$  receptors on other cells, including bone osteoclasts, juxtaglomerular kidney cells, proximal tubule kidney cells, keratinocytes, parafollicular thyroid cells, and placental trophoblasts.

While these molecules will typically be used in therapy for human patients, they may be used to treat similar or identical diseases in other warm-blooded animal species such as other primates, farm animals such as swine, cattle, and poultry; and sports animals and pets such as horses, dogs and cats.

In therapeutic and/or diagnostic applications, the molecules of the invention can be formulated for a variety of modes of administration, including systemic and topical or localized administration. Techniques and formulations generally may be found in Remington's Pharmaceutical Sciences, Mack Publishing Co., Easton, PA.

For systemic administration, oral administration is preferred. Alternatively, injection may be used, e.g., intramuscular, intravenous, intraperitoneal, and subcutaneous. For injection, the molecules of the invention are formulated in liquid solutions, preferably in physiologically compatible buffers such as Hank's solution or Ringer's solution. In addition, the molecules may be formulated in solid form and redissolved or suspended immediately prior to use. Lyophilized forms are also included.

Systemic administration can also be by transmucosal or transdermal means, or the molecules can be administered orally. For transmucosal or transdermal administration, penetrants appropriate to the barrier to be permeated are used in the formulation. Such penetrants are generally known in the art, and include, for example, for transmucosal administration bile salts and fusidic acid derivatives. In addition, detergents may be used to facilitate permeation. Transmucosal administration may be through nasal sprays, for example, or using suppositories. For oral administration, the molecules are formulated into conventional oral administration forms such as capsules, tablets, and tonics.

For topical administration, the molecules of the invention are formulated into ointments, salves, gels, or creams, as is generally known in the art.

As shown in the examples below, the amounts of various compounds of this invention which must be administered can be determined by standard procedure. Generally it is an amount between about 1 and 50 mg/kg animal to be treated.

### Recombinant Ca<sup>2+</sup> Receptors

Natural product screening has traditionally provided the lead structures for the development of diverse therapeutic molecules. However, high-throughput screening of natural product libraries or other molecule libraries for activity on the Ca<sup>2+</sup> receptor has not previously been possible. To achieve this capability, it is best to clone the Ca<sup>2+</sup> receptor cDNA and then create transfected cell lines suitable for high-throughput screening. The structure of the receptor can additionally be used to gain insight into the molecular geometry of the ligand binding site(s), and such information used to augment a rational drug design program as discussed above. Limited structure-activity studies and testing of selected natural product molecules will provide the initial structural data base necessary to guide rational natural product screening and drug design.

The bovine and human parathyroid cell Ca<sup>2+</sup> receptor cDNA can be cloned by functional expression in Xenopus oocytes. It is possible to monitor an increase in intracellular Ca<sup>2+</sup> in Xenopus oocytes indirectly by measuring current through the endogenous Ca<sup>2+</sup>-activated Cl<sup>-</sup> channel. The amplification of the response afforded by this signal transduction pathway enables the detection of receptor proteins encoded by mRNA at very low levels. This allows the detection of receptor-specific cDNA clones without the need for high affinity ligands, specific antisera, or protein or nucleic acid sequence information. An example of such a procedure follows.

Adult female Xenopus laevis were obtained from Xenopus I (Ann Arbor, MI) and maintained according to standard procedures. Lobes of ovary were excised from hypothermically-anesthetized toads. Clusters of oocytes were transferred into modified Barth's saline (MBS). Individual oocytes were obtained by incubation in MBS containing 2 mg/ml collagenase (Sigma, Type 1A) for 2h at 21°C and stage V-VI oocytes were selected for injection.

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Glass capillary tubes (1 mm diameter) were pulled to a fine tip and manually broken to achieve a tip diameter of about 15  $\mu$ M. A droplet of mRNA (1 ng/nl in diethylpyrocarbonate (DEPC)-treated water) was placed onto parafilm and drawn into the capillary tube by suction. The capillary tube was then connected to a picospritzer (WPI Instruments) and the volume of the air-pulsed droplets adjusted to deliver 50 ng of mRNA (typically 50 nl). A 35 mm culture dish with a patch of nylon stocking fixed to the bottom was used to secure the oocytes during injection of mRNA into the vegetal pole. The injected oocytes were placed into a 35 mm culture dish containing MBS, 100 ug/ml penicillin and 100 U/ml streptomycin and incubated at 18°C for 3 days.

Following incubation, an oocyte was placed into a 100  $\mu$ l plastic chamber and superfused with MBS at a flow rate of 0.5 ml/min using a peristaltic pump. Test molecules or inorganic polycations were added by rapidly moving the tubing into different buffers. Recording and current-passing electrodes were constructed from thin wall capillary tubing pulled to a resistance of 1-3 Mohms and filled with 3 M KCl. Oocytes were impaled (in the animal pole) with both electrodes under microscopic observation and connected to an Axon Instruments Axoclamp 2A voltage-clamp amplifier which was used to set the holding potential (-70 to -80 mV) and to measure the currents that were passed to maintain the holding potential. Currents were recorded directly onto a strip chart recorder.

For mRNA preparation, tissue was obtained from calves or patients with secondary HPT undergoing surgical removal of the parathyroid glands. Purified cells need not be prepared; whole pieces of gland were used to prepare mRNA that directs the expression of the  $\text{Ca}^{2+}$  receptor in Xenopus oocytes. Total cellular RNA was prepared by acid guanidinium thiocyanate/phenol extraction of homogenized glands. Oligo-dT cellulose chromatography was used to select poly(A)<sup>+</sup> mRNA by standard procedures. For size fraction-

ation of mRNA, centrifugation through glycerol gradients was used. The mRNA was denatured with 20 mM methylmercuric hydroxide and loaded (50-100 ug at a concentration of 1 mg/ml) onto a linear 15-30% glycerol gradient prepared in Beckman TLS55 tubes. Following centrifugation at 34,000 rpm for 16 hrs, 0.3 ml gradient fractions were collected and diluted in an equal volume of water containing 5mM beta-mercaptoethanol. mRNA was then recovered by two cycles of ethanol precipitation. If desired, the mRNA (50-100 ug of poly(A)<sup>+</sup>) may be separated on a 1.2% agarose/6.0 M urea preparative gel, along with a range of RNA size markers. Following visualization of the mRNA by ethidium bromide staining, gel slices containing RNA in 1.5-2.0 kb size steps are excised. mRNA is recovered from the agarose gel slices using RNAid binding matrix (according to the supplier's standard protocol; Stratagene, Inc.) and recovered mRNA fractions eluted into DEPC-treated water.

Amounts of recovered mRNA were quantified by UV absorbance measurement. The size range of mRNA contained within each fraction was determined by formaldehyde/agarose gel electrophoresis using a small quantity (0.5 ug) of each sample. The integrity of the mRNA was assessed by in vitro translation of each sample. Reticulocyte lysates (commercially available kits; BRL) were used to translate 0.05-0.5 ug of each mRNA fraction. The resulting <sup>35</sup>S-labelled proteins were analyzed by SDS-PAGE. The intact mRNA was capable of directing the synthesis of proteins of a complete size range, corresponding roughly to the sizes of the individual mRNA fractions.

A cDNA library was constructed in the vector λ ZAPII, following modifications of the technique of Gubler and Hoffman. RNA from the fraction(s) giving the best response in the oocyte assay was used as starting material. First-strand cDNA syntheses was primed with an oligo-dT/NotI primer-linker. Second-strand synthesis was by the RNase H/DNA Polymerase I self-priming method. Double-stranded cDNA was blunted with T4 DNA polymerase

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and EcoRI adaptors blunt-end ligated to the cDNA with T4 ligase. Following NotI digestion to cleave the linker, full-length cDNA was size-selected by exclusion chromatography on Sephacryl 500 HA. First-strand cDNA was radio-  
5 labeled with  $\alpha$ -<sup>32</sup>P-dATP, and all synthesis and recovery steps monitored by following the incorporation of radioactivity. Full-length cDNA recovered from the sizing column was ligated to EcoRI/NotI digested  $\lambda$  ZAPII arms. The ligation mix was test packaged with commercially  
10 available high efficiency packaging extract (Stratagene, Inc.) and plated on the appropriate host strain (XL1-blue). The percentage of recombinant phage was determined by the ratio of white to blue plaques when the library is plated on IPTG and X-gal.

15 The average insert size was determined from ten randomly selected clones. Phage DNA "mini-preps" were digested with EcoRI and NotI to release the insert, and the size determined by agarose gel electrophoresis. The library consisted of >90% recombinant phage, and the  
20 insert size ranged from 1.5 to 4.2 kb. The recombinant ligation was packaged in large scale to generate 800,000 primary clones. The packaging mix was titered and plated at 50,000 plaques per 15 cm plate. Each pool of 50,000 clones was eluted in SM buffer and stored individually.

25 Plate lysate stocks of each of the clone pools were used for small scale phage DNA preparation. Phage particles are concentrated by polyethylene glycol precipitation, and phage DNA purified by proteinase K digestion followed by phenol:chloroform extraction. Twenty micro-  
30 grams of DNA are digested with NotI, and used as template for in vitro transcription of sense-strand RNA. In vitro transcription is according to standard protocols, utilizing T7 RNA polymerase and 5' cap analog m<sup>7</sup>GpppG in a 50 $\mu$ l total reaction volume. Following Dnase I/Proteinase K  
35 digestion and phenol/chloroform extraction, the RNA is concentrated by ethanol precipitation and used for oocyte injection.

Oocytes were injected with synthetic mRNA (cRNA) from each of the 16 library subpools constituting 50,000 independent clones each. After incubation for 3 to 4 days, oocytes were assayed for the ability of 10 mM neomycin to  
5 elicit a  $\text{Ca}^{2+}$  dependent Cl-current. A pool designated 6 gave a positive signal and thus contains a cDNA clone encoding a functional calcium receptor. In order to decrease the complexity of pool 6 and thus proceed towards the purification of the calcium receptor clone contained  
10 within this pool, pool 6 phage were replated at ~20,000 plaques per plate and 12 plates harvested. DNA was prepared from each of these subpools and cRNA synthesized. Again, oocytes were injected with cRNA and assayed 3-4 days later for the ability of 10 mM neomycin to elicit a  
15  $\text{Ca}^{2+}$  dependent Cl-current. A subpool 6-3 was positive and this pool was subjected to a further round of plating reducing the complexity of pools to around 5,000 clones per pool. Pools were again assayed by preparation of cRNA and injection in oocytes. A subpool 6-3.4 was positive.  
20 In order to expedite further purification of the positive clone in pool 6-3.4, phage DNA from this pool was rescued as plasmid DNA by superinfection with the helper phage, ExAssist. Transfection of rescued plasmids into bacterial strain DH5alphaF' resulted in transformed bacterial colo-  
25 nies on ampicillin plates. These were harvested in pool of 900 clones each. Plasmid DNA was then prepared from each subpool and cRNA synthesized and assayed in the usual manner. Subpool 6-3.4.4 was positive. Bacteria contain-  
ing the plasmid subpool 6-3.4.4 were subsequently plated  
30 in subpools of ~50 clones each. Continuation of this process is expected to result in a single clone encoding a functional calcium receptor.

Initial experiments used Xenopus oocytes injected with water or poly(A)<sup>+</sup>-enriched mRNA (50 ng) from bovine  
35 parathyroid cells. After three days, the oocytes were examined for their ability to increase intracellular  $\text{Ca}^{2+}$  in response to increases in the concentration of extra-

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cellular di- and trivalent cations. The oocytes were impaled with recording and current-passing electrodes and  $[Ca^{2+}]_i$  was assessed indirectly by measuring currents through the endogenous  $Ca^{2+}$ -activated  $Cl^-$  channel. In  
5 oocytes injected with poly(A)<sup>+</sup>-enriched mRNA from bovine (or human, Fig. 26) parathyroid cells, increasing the concentration of extracellular  $Ca^{2+}$  from 0.7 to 3, 5 or 10 mM caused a rapid and transient increase in the  $Cl^-$  conductance which then oscillated around a higher basal conductance. Increasing the concentration of extracellular  $Mg^{2+}$   
10 from 1 to 10 mM likewise evoked oscillatory increases in  $Cl^-$  conductance. The  $Cl^-$  conductance response to extracellular  $Mg^{2+}$  persisted when the extracellular  $Ca^{2+}$  concentration was reduced to  $< 1 \mu M$  (Fig. 25).

15 The impermeant trivalent cation  $Gd^{3+}$  (600  $\mu M$ ) also caused oscillatory increases in the  $Cl^-$  conductance (Fig. 25). Such increases in the  $Cl^-$  conductance which oscillate and persist in the nominal absence of extracellular  $Ca^{2+}$  are noted when oocytes have been allowed to  
20 express other  $Ca^{2+}$ -mobilizing receptors and are stimulated with the appropriate ligand (e.g., substance K, Fig. 25). In these instances, the increase in  $Cl^-$  conductance reflects the mobilization of intracellular  $Ca^{2+}$ . These initial studies likewise show that extracellular polycations mobilize intracellular  $Ca^{2+}$  in parathyroid cell  
25 mRNA-injected oocytes.

Oocytes injected with water did not show any change in the  $Cl^-$  current when exposed to extracellular  $Ca^{2+}$  (10 mM) or  $Mg^{2+}$  (20 or 30 mM). In one series of experiments,  
30 oocytes were injected with the mRNA encoding the substance K receptor. In these oocytes, extracellular  $Mg^{2+}$  (20 mM) did not evoke any current but the cells responded vigorously to the addition of substance K (Fig. 25). These experiments indicate that there is no endogenous sensitivity of the oocyte to extracellular  $Ca^{2+}$  or  $Mg^{2+}$ .  
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Similar experiments were performed using oocytes injected with poly(A)<sup>+</sup>-enriched mRNA prepared from human parathyroid glands (hyperplastic tissue from a case of secondary HPT). In these oocytes, increasing the concentration of extracellular Ca<sup>2+</sup> caused a reversible increase in the Cl<sup>-</sup> conductance which oscillated (Fig. 26). The addition of 300 μM La<sup>3+</sup> likewise caused oscillatory increases in the Cl<sup>-</sup> conductance. Increasing the concentration of extracellular Mg<sup>2+</sup> from 1 to 10 mM evoked increases in the Cl<sup>-</sup> conductance that persisted in the absence of extracellular Ca<sup>2+</sup>. Additional experiments suggest that the response to extracellular Ca<sup>2+</sup> is concentration dependent. Thus, in three mRNA-injected oocytes, Cl<sup>-</sup> conductance increased to a maximum of 111 ± 22 nA at 3 mM and 233 ± 101 nA at 10 mM extracellular Ca<sup>2+</sup>.

The results obtained in Xenopus oocytes demonstrate the presence of a mRNA(s) in parathyroid cells encoding a protein(s) which can impart, in normally unresponsive cells, sensitivity to extracellular Ca<sup>2+</sup>. Moreover, the ability of extracellular Mg<sup>2+</sup> to evoke oscillatory increases in Cl<sup>-</sup> current in the absence of extracellular Ca<sup>2+</sup> demonstrates that the Cl<sup>-</sup> current depends on the mobilization of intracellular Ca<sup>2+</sup> rather than influx of extracellular Ca<sup>2+</sup>. The results obtained with La<sup>3+</sup> likewise show that the expressed protein(s) is linked to the mobilization of intracellular Ca<sup>2+</sup>. Together, these data show that the expressed protein(s) acts as a cell surface receptor rather than a channel. These studies provide compelling evidence for the existence of a Ca<sup>2+</sup> receptor protein on the surface of parathyroid cells and demonstrate the feasibility of using the Xenopus oocyte system to achieve the molecular cloning of the Ca<sup>2+</sup> receptor cDNA.

In another series of experiments, parathyroid cell mRNA, denatured with methylmercuric hydroxide, was size-fractionated by centrifugation through a glycerol gradient. Ten fractions were collected. Each group was

injected into Xenopus oocytes and after a three day incubation period the oocytes were assayed for expression of the  $\text{Ca}^{2+}$  receptor. Those oocytes injected with fractions 4-6 showed the largest and most consistent increases in  $\text{Cl}^-$  conductance in response to extracellular  $\text{Ca}^{2+}$  (Fig. 35). These results indicate that the  $\text{Ca}^{2+}$  receptor is encoded by mRNA in a size range of 2.5-3.5 kb. This indicates that a strategy using direct expression of RNA synthesized from a transcription vector cDNA library is feasible. Size-fractionation experiments of this sort were conducted and in each of three different fractionation experiments similar results were obtained.

The mRNA fractions obtained and characterized in the preceding experiments can be assayed by injection into oocytes. For each mRNA fraction, 10-20 oocytes are injected with 50 ng of RNA at a concentration of 1 ng/nl in water. Injected oocytes are maintained at 18° C for 48-72 h after which they are assessed for expression of the  $\text{Ca}^{2+}$  receptor using measurements of  $\text{Cl}^-$  current. For each group of injected oocytes the number positive for expression of the receptor, as well as the magnitude of the  $\text{Ca}^{2+}$ -dependent  $\text{Cl}^-$  current measured, is determined. As negative controls, oocytes are injected with rat liver poly(A)<sup>+</sup>-enriched mRNA, yeast RNA, or water.

It is expected that an mRNA in the range of 2.5 - 3.5 kb will encode the receptor. mRNA of a larger size may necessitate a cloning approach based on hybrid depletion of parathyroid mRNA prior to oocyte injection. This strategy is not dependent upon the generation of full-length cDNA clones for success. If receptor expression is not obtained with a single size fraction of mRNA, oocytes are injected with mixed size fractions to determine a combination that does give rise to a functional receptor. If it does appear that multiple subunits are necessary for the formation of a functional receptor, the hybrid depletion expression cloning strategy is used. In this approach, clones are selected on the basis of their abil-

ity to deplete a specific mRNA species from the total mRNA population. A clone encoding a single subunit is identified by its ability to prevent the formation of the active multi-subunit complex. By exhaustive screening it is possible to identify clones encoding all of the necessary subunits.

This approach permits the isolation of clones encoding individual subunits required to form a functional receptor complex. Synthetic RNA from pools of clones are assayed for their ability to induce expression of the  $\text{Ca}^{2+}$  receptor in Xenopus oocytes by the same techniques used to analyze the original mRNA fractions. Originally, 10 pools representing 100,000 primary clones each are examined. Pools of clones showing a positive response are screened at lower (typically 4 to 10 fold) complexity, and again positive pools further subdivided and screened. This process of library sub-fractionation is followed until individual positive clones are identified. As a negative control for the oocyte expression assay, anti-sense transcripts are generated by T7 transcription of those DNA templates that induce a positive response. Anti-sense transcripts are unable to give rise to an authentic receptor, and this will control any non-specific positive signal arising from injection of synthetic RNA. Another concern is the fact that synthetic RNA can occasionally "poison" translation in injected oocytes, by an undefined mechanism. To control for this possibility, synthetic RNAs giving a negative response are co-injected at various dilutions with parathyroid cell mRNA, to determine if they are non-specifically interfering with the expression of the  $\text{Ca}^{2+}$  receptor.

When an individual clone encoding the  $\text{Ca}^{2+}$  receptor is identified, the cDNA insert will be excised from the  $\lambda$  vector and used for large scale production of synthetic RNA. Oocyte injection of this single RNA species allows rigorous assessment of the characteristics of the expressed receptor.



If the size of the mRNA encoding the  $\text{Ca}^{2+}$  receptor is too large for cloning by direct transcription and expression, or if multiple subunits are involved, a hybrid-depletion technique of screening pools of clones is used.

5 cDNA insert DNA will be prepared from pools of clones from the size-selected parathyroid cell cDNA library. This DNA is hybridized to parathyroid cell mRNA under conditions that permit the formation of DNA/RNA duplexes. The unannealed, hybrid-depleted RNA is recovered and used for

10 oocyte injections. DNA from pools of clones containing sequences representing  $\text{Ca}^{2+}$  receptor mRNA is depleted from this mRNA from the total parathyroid cell mRNA population, and expression of the receptor is reduced or absent upon oocyte injection. A process of sub-fractionation is fol-

15 lowed on pools of clones of decreasing complexity, at each step assaying for cloned DNA that depletes  $\text{Ca}^{2+}$  receptor-encoding mRNA from the total parathyroid cell mRNA population. The use of an internal control during the hybrid depletion assays ensures that the hybrid-depleted RNA is

20 intact and capable of being translated in the oocyte.

Human parathyroid cells express a beta-adrenergic receptor coupled to adenylate cyclase. This receptor can be expressed in oocytes, where it is capable of agonist-induced activation of the endogenous adenylate cyclase.

25 During the hybrid-depletion screening for  $\text{Ca}^{2+}$  receptor clones, oocytes injected with hybrid depleted mRNA are assayed for isoproterenol-induced adenylate cyclase activation. A positive response in this assay serves to indicate that any observed inhibition of  $\text{Ca}^{2+}$  receptor response

30 is specific, and not due to a general inhibition of the total mRNA population.

The hybrid-depletion screening strategy can result in the isolation of clones that do not contain a complete protein coding region. Positive clones isolated by this

35 screening strategy are sequenced to determine their protein coding capacity. Northern blot analysis of human parathyroid gland RNA permits the determination of the

size of the complete mRNA corresponding to specific clones. If positive clones do not appear to be full-length, the cloned cDNA will be used as a hybridization probe to screen a parathyroid gland cDNA library for complete cDNAs.

A variety of cell lines are capable of coupling exogenously expressed receptors to endogenous functional responses. A number of these cell lines (e.g., NIH-3T3, HeLa, NG115, CHO, HEK, 293 and COS7) can be tested to confirm that they lack an endogenous  $\text{Ca}^{2+}$  receptor. Those lines lacking a response to external  $\text{Ca}^{2+}$  can be used to establish stably transfected cell lines expressing the cloned  $\text{Ca}^{2+}$  receptor.

Sequence analysis of  $\text{Ca}^{2+}$  receptor cDNA clones identified by expression cloning will delineate the open reading frame encoding the receptor protein. The coding region of the cDNA will be subcloned into multiple cloning site of the eukaryotic expression vector pMSG. This vector allows high level transcription driven by the mouse mammary tumor virus (MMTV) promoter, and is active in a wide variety of mammalian cells. The vector also contains a gpt gene for resistance to mycophenolic acid which is under the control of the SV40 early promoter, and sequences necessary for selection and growth in bacteria. Large quantities of the expression vector/receptor cDNA plasmid construct will be grown and purified from E. coli.

The most effective method for transfection of eukaryotic cell lines with plasmid DNA varies with the given cell type. The  $\text{Ca}^{2+}$  receptor expression construct will be introduced into cultured cells by the appropriate technique, either  $\text{Ca}^{2+}$  phosphate precipitation, DEAE-dextran transfection, lipofection or electroporation. Following the transfection procedure, cells are grown in the presence of the antibiotic G418 to select for cells expressing the neomycin resistance gene. Colonies of G418 resistant transfectants will be subcloned and established as individual cell lines. Expression of the  $\text{Ca}^{2+}$  receptor protein

in G418 resistant cells will be assessed by several methods. Southern blot and slot blot analysis will confirm the presence and the copy number of the receptor cDNA sequence. Northern blot analysis will be used to demonstrate that receptor mRNA is being transcribed from the plasmid construct. Functional expression of the receptor protein will be determined by measuring the mobilization of intracellular  $\text{Ca}^{2+}$  in response to externally applied  $\text{Ca}^{2+}$  receptor agonists.

Cloning the  $\text{Ca}^{2+}$  receptor enables both structural and functional studies of this novel receptor. Recombinantly produced receptor may be crystallized for structural studies. Stably transfected cell lines expressing the receptor can be used for high-throughput screening of natural product or other compound libraries. Molecules of the requisite potency and specificity can be labeled (radioactively or fluorescently). The ability of test molecules/extracts to displace such a labeled molecule will form the basis of a high-throughput assay for screening.

Given the appropriate cells or tissues expressing other calcium receptors, these receptors may be cloned in a manner analogous to that described above for the parathyroid cell calcium receptor. For example, mRNA from human osteoclastoma tissue encodes the osteoclast calcium receptor (Figure 34). Thus, to isolate a clone for the human osteoclast receptor, one need only isolate mRNA from osteoclastoma tissue, prepared a cDNA library and assay/fractionate subpools as described above. Furthermore, the preferred receptors for drug screening are of human origin. A clone encoding a calcium receptor from one species may be used to obtain the corresponding human cDNA clone by cross-hybridization as is well known by those skilled in the art. In addition, the clone of the parathyroid cell or other cell  $\text{Ca}^{2+}$  receptor allows isolation of genes encoding similar  $\text{Ca}^{2+}$ -sensing proteins in other cells, and expression of those proteins. This is achieved by a variety of approaches. Southern blot analysis of human geno-

mic DNA, utilizing the  $\text{Ca}^{2+}$  receptor cDNA as a hybridization probe, will give an indication of the number of related sequences encoded within the genome; hybridization at varying stringencies will give an indication of the degree of divergence among the related sequences. This will provide information about the potential number of genes encoding related receptor proteins. Northern blot analysis with  $\text{Ca}^{2+}$  receptor cDNA as probe will determine if the same or related transcripts are present in various tissues. If related transcripts homologous to the parathyroid cell  $\text{Ca}^{2+}$  receptor are detected, it is a relatively simple matter to obtain clones of these mRNAs, either by screening the appropriate cDNA libraries or by polymerase chain reaction (PCR) techniques. Novel receptor clones so obtained can be assessed functionally by expression, either in oocytes or in transfected cell lines. Transfected cell lines expressing a cell-specific  $\text{Ca}^{2+}$  receptor can then provide a means of high-throughput screening for molecules that act specifically on the  $\text{Ca}^{2+}$ -sensing mechanism of, for example, osteoclasts or juxtaglomerular cells.

In an alternative method, the calcium receptor can be cloned by expression in eukaryotic cells. For example, a cDNA library can be prepared from parathyroid mRNA and cloned into the eukaryotic expression vector, pCDNA1. Subpools from this library can be transfected into eukaryotic cells such as COS7 or HEK293 cells resulting in relatively high level transient expression of encoded cDNA sequences. Cells transfected with a function calcium receptor clone will express the calcium receptor which can then be activated by calcium, neomycin or other calcimimetic compounds. If cells are first loaded with a fluorometric indicator for  $[\text{Ca}^{2+}]_i$ , activation of the calcium receptor results in increased fluorescence. Thus library subpools containing the calcium receptor are identified by their ability, upon transfection into eukaryotic cells, to induce a calcium or calcimimetic-specific increase in

fluorescence. This fluorescence can be detected using either a fluorimeter or a fluorescence activated cell sorter (FACS).

In an alternative method, the  $\text{Ca}^{2+}$  receptor can be  
5 cloned by use of a monoclonal antibody generated against the receptor. Monoclonal antibodies provide powerful tools for the immunoaffinity purification of specific proteins. Once purified, limited amino acid sequence data can be obtained from the protein of interest, and used to  
10 design oligonucleotide sequence probes to screen for clones of the complete cDNA sequence.

For production of hybridomas, whole bovine parathyroid gland cells are used as the immunogen. Purified, dispersed cells are obtained, and live cell preparations  
15 are injected intraperitoneally into the appropriate mouse strain, according to established procedures. Standard protocols are followed for immunization schedules and for the production of hybridomas. A two-step screening procedure is used to identify hybridomas secreting monoclonal  
20 antibodies that recognize the  $\text{Ca}^{2+}$  receptor. The initial screen will identify those monoclonals that recognize parathyroid cell surface antigens. Immunohistochemical techniques are then used to screen hybridoma supernatants for the presence of mouse antibodies that bind to the  
25 surface of parathyroid cells. This screen can be performed on fixed sections of parathyroid gland tissue, or on dispersed cells in primary culture. The techniques for this assay are well established in the literature.

This screen will identify hybridomas producing monoclonal antibodies to a variety of cell surface determinants, and monoclonals specific for the  $\text{Ca}^{2+}$  receptor would be expected to comprise only a small subset of these. To  
30 identify monoclonal antibodies that bind to the  $\text{Ca}^{2+}$  receptor, hybridoma supernatants that test positive in the initial screen are assayed for their ability to block the response of cultured parathyroid cells to  $\text{Ca}^{2+}$  receptor agonists. Some antibodies that bind to the extracellular  
35

domain of the receptor are expected to inhibit or activate ligand binding or to otherwise interfere with or affect receptor activation.

Monoclonal antibodies positive in both screens are characterized through Western blotting, immunoprecipitation and immunohistochemistry. This permits the determination of the size of the antigen that is recognized and its tissue distribution. The appropriate monoclonal antibody is then used for purification of the  $\text{Ca}^{2+}$  receptor protein by immunoaffinity chromatography, following standard techniques. Sufficient quantities of protein are obtained to allow limited amino acid sequence determination. Degenerate oligonucleotide probes are then designed on the basis of the peptide sequence information. These probes are then used to screen parathyroid gland cDNA libraries for full length clones of the  $\text{Ca}^{2+}$  receptor. Clones obtained are characterized by DNA sequencing and by functional expression in the oocyte system and in cultured mammalian cell lines.

Alternatively, the antibodies can be used to screen expression libraries, e.g., cDNA libraries in  $\lambda$ gt11 or its equivalent, to determine those clones expressing antigenically reactive protein. Such clones can then be sequenced to determine whether they encode a protein that might be a  $\text{Ca}^{2+}$  receptor.

It will also be appreciated by those skilled in the art that phage display libraries can be used to clone and analyze calcium receptors in place of monoclonal antibodies. In these libraries, antibody variable regions or random peptides are shotgun cloned into phage expression vectors such that the antibody regions or peptides are displayed on the surface of the phage particle. Phage which display antibody regions or peptides capable of high specific binding to calcium receptors will bind to cells which display these receptors (e.g. parathyroid cells, C-cells, osteoclasts, etc.). Millions of such phage can be panned against these cell types selecting only those

phage which can bind to these cells (which includes those phage binding to calcium receptors). In this manner, the complexity of the library can be vastly reduced. Subsequently, the screens described above for monoclonal antibodies can be used to isolate phage which display calcium receptor-binding antibody or peptide regions, and these phage can be used to isolate the calcium receptor for purposes of structural identification and cloning. Kits to prepare such phage display libraries are commercially available (e.g. Stratacyte, or Cambridge Antibody Technology Limited). Recombinant phage endowed with such calcium receptor-binding properties can also be used in lieu of monoclonal antibodies in the various analyses of calcium receptors. Such phage can also be used in high throughput binding competition screens to identify organic compounds capable of functional binding to calcium receptors which can serve as structural leads for the development of human therapeutics acting at the calcium receptor.

In another alternative, affinity cross-linking of radioligands to their receptors can be used to isolate the receptor protein as described by Pilch & Czech, 1 Receptor Biochem. Methodol. 161, 1984. Covalent attachment of a radioligand allows extensive washing to remove non-specific binding. For example, a high affinity molecule, e.g., a random copolymer of arginine and tyrosine (MW = 22K; argtyr ratio = 4:1) which mobilizes intracellular  $\text{Ca}^{2+}$  with an  $\text{EC}_{50}$  of about 100 nM or less, is iodinated with  $^{125}\text{I}$ , and cross-linked. Protamines, because of their much smaller size, may be preferable in cross-linking studies and can be reductively alkylated as described by Dottavio-Martin & Ravel, 87 Analyt. Biochem. 562, 1978.

Nonspecific labelling is kept to a minimum by cross-linking in the presence of unlabeled polycations and di- and trivalent cations. At high concentrations of these molecules nonspecific interactions of the label with the cell surface might be reduced.

### Uses

Primary hyperparathyroidism (HPT) is characterized by hypercalcemia and elevated levels of circulating PTH. One of the major defects in HPT appears to be a diminished sensitivity of parathyroid cells to negative feedback regulation by extracellular  $\text{Ca}^{2+}$ . Thus, in tissue from patients with primary HPT, the "set-point" for extracellular  $\text{Ca}^{2+}$  is shifted to the right so that higher than normal concentrations of extracellular  $\text{Ca}^{2+}$  are required to depress PTH secretion. Moreover, in primary HPT, even high concentrations of extracellular  $\text{Ca}^{2+}$  often depress PTH secretion only partially. In secondary (uremic) HPT, a similar increase in the set-point for extracellular  $\text{Ca}^{2+}$  is observed even though the degree to which  $\text{Ca}^{2+}$  suppresses PTH secretion is normal. The changes in PTH secretion are paralleled by changes in  $[\text{Ca}^{2+}]_i$ ; the set-point for extracellular  $\text{Ca}^{2+}$ -induced increases in  $[\text{Ca}^{2+}]_i$  is shifted to the right and the magnitude of such increases is reduced. Moreover, staining of tissue with a monoclonal antibody that appears to recognize the  $\text{Ca}^{2+}$  receptor is diminished in adenomatous and hyperplastic parathyroid cells.

The  $\text{Ca}^{2+}$  receptor constitutes a discrete molecular entity for pharmacological intervention. Molecules that mimic or antagonize the action of extracellular  $\text{Ca}^{2+}$  are beneficial in the long-term management of both primary and secondary HPT. Such molecules provide the added impetus required to suppress PTH secretion which the hypercalcemic condition alone cannot achieve. Such molecules with greater efficacy than extracellular  $\text{Ca}^{2+}$  may overcome the apparent nonsuppressible component of PTH secretion which is particularly troublesome in adenomatous tissue. Alternatively or additionally, such molecules can depress synthesis of PTH, as prolonged hypercalcemia has been shown to depress the levels of preproPTH mRNA in bovine and human adenomatous parathyroid tissue. Prolonged hypercalcemia also depresses parathyroid cell proliferation in vitro, so calcimimetics can also be effective in limiting



the parathyroid cell hyperplasia characteristic of secondary HPT.

Other cells in the body can respond directly to physiological changes in the concentration of extracellular  $\text{Ca}^{2+}$ . Calcitonin secretion from parafollicular cells in the thyroid (C-cells) is regulated by changes in the concentration of extracellular  $\text{Ca}^{2+}$ . Renin secretion from juxtaglomerular cells in the kidney, like PTH secretion, is depressed by increased concentrations of extracellular  $\text{Ca}^{2+}$ . Extracellular  $\text{Ca}^{2+}$  causes the mobilization of intracellular  $\text{Ca}^{2+}$  in these cells. Isolated osteoclasts respond to increases in the concentration of extracellular  $\text{Ca}^{2+}$  with corresponding increases in  $[\text{Ca}^{2+}]_i$  that arise partly from the mobilization of intracellular  $\text{Ca}^{2+}$ . Increases in  $[\text{Ca}^{2+}]_i$  in osteoclasts are associated with an inhibition of functional responses (bone resorption) analogous to PTH secretion in parathyroid cells. Thus, there are sufficient indications to suggest that  $\text{Ca}^{2+}$ , in addition to its ubiquitous role as an intracellular signal, also functions as an extracellular signal to regulate the responses of certain specialized cells. Molecules of this invention can be used in the treatment of diseases associated with disrupted  $\text{Ca}^{2+}$  responses in these cells.

Cloning the  $\text{Ca}^{2+}$  receptor on parathyroid cells and other cells will allow the presence of homologous proteins in other cells to be directly assessed. A family of structurally homologous  $\text{Ca}^{2+}$  receptor proteins can thus be obtained. Such receptors will allow understanding of how these cells detect extracellular  $\text{Ca}^{2+}$  and enable evaluation of the mechanism(s) as a site of action for the therapeutics described herein effective in the treatment of HPT, osteoporosis, and hypertension, and novel therapies for other bone and mineral-related diseases.

Other uses are discussed above. For example, recombinant  $\text{Ca}^{2+}$  receptor proteins may be used in therapy, and introduced by standard methods, e.g., by transfection of nucleic acid encoding that protein. In addition, such

protein is useful in assays for calcimimetic molecules of this invention.

The following examples illustrate the invention but do not limit its scope.

## 5 Examples

In the studies described herein, a variety of organic molecules were found to mobilize intracellular  $\text{Ca}^{2+}$  and depress PTH secretion in parathyroid cells. These molecules are structurally diverse but most have a net positive charge at physiological pH. The cationic nature of the organic molecules plays an important role but is not the sole factor determining activity.

### Example 1: Screening Calcimimetic Molecules on Bovine Parathyroid cells

15 Dissociated bovine parathyroid cells were purified on gradients of Percoll and cultured overnight in serum-free medium. The cells were subsequently loaded with fura-2 and the concentration of free intracellular  $\text{Ca}^{2+}$  measured fluorimetricly. Changes in  $[\text{Ca}^{2+}]_i$  were used to screen for  
20 molecules active at the  $\text{Ca}^{2+}$  receptor. To be considered a calcimimetic, a molecule was required to show the normal effects caused by increasing extracellular  $\text{Ca}^{2+}$  and triggered by the activation of the  $\text{Ca}^{2+}$  receptor. That is,

1) The molecule must elicit an increase in  $[\text{Ca}^{2+}]_i$ ;  
25 that persists in the absence of extracellular  $\text{Ca}^{2+}$  (demonstrating the mobilization of intracellular  $\text{Ca}^{2+}$ );

2) The molecule must cause a decrease in isoproterenol-stimulated cyclic AMP formation which is blocked by pertussis toxin;

30 3) The molecule must inhibit PTH secretion over the same range of concentrations that cause the increase in  $[\text{Ca}^{2+}]_i$ ; and

4) The concentration-response curves for  $\text{Ca}^{2+}$  mobilization and PTH secretion by the molecule must be

shifted to the right by a PKC activator, such as phorbol myristate acetate (PMA).

Several structurally different classes of molecules were tested: polyamines, aminoglycoside antibiotics, protamine, and polymers of lysine or arginine. The structures of these molecules are depicted in Figure 1. Included in Figure 1 are the net positive charge of the molecules and their  $EC_{50}$ 's for evoking the mobilization of intracellular  $Ca^{2+}$  in bovine parathyroid cells.

10 In general, the greater the net positive charge on the molecule, the greater its potency in causing the mobilization of intracellular  $Ca^{2+}$ . However, some striking exceptions to this apparent rule have been found as discussed below.

15 As can be seen from the figures, spermine, neomycin B, and protamine evoked rapid and transient increases in  $[Ca^{2+}]_i$  in fura-2-loaded bovine parathyroid cells (Figs. 6, 7, 11). They did not, however, cause sustained, steady-state increases in  $[Ca^{2+}]_i$  in bovine parathyroid cells (Fig. 6, 11), although they did in human parathyroid cells (Fig. 19). In this respect, they resembled the cytosolic  $Ca^{2+}$  response elicited by extracellular  $Mg^{2+}$ , which causes the mobilization of intracellular  $Ca^{2+}$  unaccompanied by an influx of extracellular  $Ca^{2+}$  in bovine cells (Fig. 11b).  
20 Transient increases in  $[Ca^{2+}]_i$  elicited by spermine, neomycin B, or protamine were not blocked by low concentrations ( $1 \mu M$ ) of  $La^{3+}$  or  $Gd^{3+}$  (Fig. 11f,g). Cytosolic  $Ca^{2+}$  transients elicited by the molecular polycations persisted in the absence of extracellular  $Ca^{2+}$  but were blocked when  
25 cellular stores of  $Ca^{2+}$  were depleted by pretreatment with ionomycin (Figs. 7; 11h,i). All these molecules therefore cause the mobilization of intracellular  $Ca^{2+}$  in parathyroid cells.

It was additionally shown that the molecular polycations mobilized the same pool of intracellular  $Ca^{2+}$  as that  
35 used by extracellular  $Ca^{2+}$ . Thus, increasing the concentration of extracellular  $Ca^{2+}$  progressively inhibited the

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transient increases in  $[Ca^{2+}]_i$  evoked by spermine (Fig. 6). Conversely, a maximally effective concentration of spermine or neomycin B (Fig. 12) blocked transient, but not steady-state increases in  $[Ca^{2+}]_i$  evoked by extracellular  $Ca^{2+}$ .

Significantly, spermine, neomycin B, and protamine inhibited PTH secretion to the same extent as extracellular  $Ca^{2+}$ . These inhibitory effects on secretion were obtained at concentrations that caused the mobilization of intracellular  $Ca^{2+}$  (Figs. 8, 13). These findings are relevant to understanding the mechanisms contributing to the regulation of PTH secretion by extracellular  $Ca^{2+}$ . Because a variety of inorganic polycations all inhibit secretion, yet only extracellular  $Ca^{2+}$  causes sustained, steady-state increases in  $[Ca^{2+}]_i$ , such increases in  $[Ca^{2+}]_i$  cannot be importantly involved in the regulation of secretion. Mobilization of intracellular  $Ca^{2+}$ , rather than the influx of extracellular  $Ca^{2+}$ , is the essential mechanism associated with the inhibition of PTH secretion. This is important because it defines the sufficient mechanism to be affected if a molecule is to affect PTH secretion; molecules stimulating selectively the influx of extracellular  $Ca^{2+}$  will be relatively ineffective in suppressing PTH secretion. In contrast, molecules causing solely the mobilization of intracellular  $Ca^{2+}$  should be just as efficacious as extracellular  $Ca^{2+}$  in suppressing PTH secretion.

Like the mobilization of intracellular  $Ca^{2+}$  elicited by extracellular  $Ca^{2+}$ , that elicited by molecular polycations was depressed by PMA. A representative experiment showing the preferential inhibitory effects of PMA on cytosolic  $Ca^{2+}$  transients elicited by spermine is shown in Fig. 14. Cytosolic  $Ca^{2+}$  transients evoked by ATP were unaffected, even when a submaximal concentration of ATP was used. The effect of PMA on cytosolic  $Ca^{2+}$  transients elicited by the molecular polycations paralleled its effect on responses to extracellular  $Ca^{2+}$ ; in both cases

there was a shift to the right in the concentration-response curve (Fig. 15). The depressive effects of PMA on  $[Ca^{2+}]$ ; were accompanied by potentiating effects on secretion which were overcome at higher concentrations of the organic polycations (Fig. 16).

The mobilization of intracellular  $Ca^{2+}$  elicited by molecular polycations was associated with increases in the formation of inositol phosphates. For example, protamine caused a rapid (within 30 s) increase in the formation of  $IP_3$ , which was accompanied by a rise in levels of  $IP_1$ . Both these effects were dependent on the concentration of extracellular protamine (Fig. 17). Moreover, pretreatment with PMA blunted the formation of inositol phosphates elicited by molecular polycations. Representative results obtained with spermine are presented in Fig. 18.

Spermine, neomycin B, and protamine depressed isoproterenol-induced increases in cyclic AMP. Like the inhibitory effects of extracellular  $Ca^{2+}$  on cyclic AMP formation, those caused by molecular polycations were blocked by pretreatment with pertussis toxin (Table 2).

Table 2

		cyclic AMP (% of control)	
		control	+PTx
25	0.5 mM $Ca^{2+}$	100	106 ± 8
	2.0 mM $Ca^{2+}$	19 ± 4	94 ± 2
	0.5 mM $Ca^{2+}$ , 200 $\mu$ M Spermine	23 ± 5	93 ± 6
	0.5 mM $Ca^{2+}$ , 30 $\mu$ M Neomycin B	28 ± 8	87 ± 6
	0.5 mM $Ca^{2+}$ , 2 $\mu$ g/ml Protamine	20 ± 4	89 ± 9
30	Pertussis toxin (PTx) blocks the inhibitory effects of extracellular $Ca^{2+}$ and molecular polycations on cyclic AMP formation. Bovine parathyroid cells were cultured for 16 h with or without 100 ng/ml pertussis toxin. The cells were subsequently washed and incubated for 15 min with 10		
35	$\mu$ M isoproterenol with or without the indicated concentrations of extracellular $Ca^{2+}$ or molecular polycations.		

Total cyclic AMP (cells + supernatant) was determined by RIA and the results are expressed as a percentage of the levels obtained in 0.5 mM  $\text{Ca}^{2+}$  ( $112 \pm 17$  pmole/ $10^6$  cells). Each value is the mean  $\pm$  SEM of three experiments.

5

In human parathyroid cells, extracellular  $\text{Mg}^{2+}$  elicited a sustained, steady-state increase in  $[\text{Ca}^{2+}]_i$  in addition to a rapid transient increase (Fig. 10). As in bovine parathyroid cells responding to extracellular  $\text{Ca}^{2+}$ , the steady-state increase in  $[\text{Ca}^{2+}]_i$  evoked by  $\text{Mg}^{2+}$  in human parathyroid cells results from  $\text{Ca}^{2+}$  influx through voltage-insensitive channels (Fig. 10a). This effect of  $\text{Mg}^{2+}$  on steady-state  $[\text{Ca}^{2+}]_i$  in human parathyroid cells is seen in both adenomatous and hyperplastic tissue.

15 Neomycin B and spermine were tested for effects on  $[\text{Ca}^{2+}]_i$  in human parathyroid cells prepared from adenomatous tissue. Representative results with neomycin B are shown in Fig. 19. Neomycin B caused not only a transient but additionally a steady-state increase in  $[\text{Ca}^{2+}]_i$  in human parathyroid cells (Fig. 19a). Thus, in human cells, the pattern of change in  $[\text{Ca}^{2+}]_i$  evoked by extracellular  $\text{Ca}^{2+}$ ,  $\text{Mg}^{2+}$  or neomycin B is very similar.

25 Cytosolic  $\text{Ca}^{2+}$  transients elicited by neomycin B persisted in the presence of  $\text{La}^{3+}$  ( $1\mu\text{M}$ ) and absence of extracellular  $\text{Ca}^{2+}$ . Neomycin B therefore causes the mobilization of intracellular  $\text{Ca}^{2+}$  in human parathyroid cells. Neomycin B inhibited PTH secretion from human parathyroid cells at concentrations that caused the mobilization of intracellular  $\text{Ca}^{2+}$  (Fig. 13). There were, however, some differences in the responses of human and bovine parathyroid cells to neomycin B. The  $\text{EC}_{50}$  of neomycin B for the mobilization of intracellular  $\text{Ca}^{2+}$  was  $40\mu\text{M}$  in bovine and  $20\mu\text{M}$  in human parathyroid cells (cf. Figs. 13 and 15), whereas the potency of spermine was similar in bovine and human parathyroid cells ( $\text{EC}_{50} = 150$

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$\mu\text{M}$ ). Thus, although bovine cells can be used for initial studies to screen test molecules for activity, it is important to perform follow-up studies using human parathyroid cells.

5 To assess the effects of molecular polycations on C-cells, a neoplastic cell line, derived from a rat medullary thyroid carcinoma (rMTC 6-23 cells) was used. Both spermine (10 mM) and neomycin B (5 mM) were without effect on basal  $[\text{Ca}^{2+}]_i$  in these cells. Nor did either molecule  
10 affect the response to the subsequent addition of extracellular  $\text{Ca}^{2+}$ . Representative results documenting the lack of effect of neomycin B are shown in Fig 21. Neomycin B (1 mM) or spermine (1 or 5 mM) failed to evoke any increase in  $[\text{Ca}^{2+}]_i$  in osteoclasts (Fig. 23). In the trace  
15 shown, there appeared to be some potentiation of the response to a subsequent increase in the concentration of extracellular  $\text{Ca}^{2+}$ , although this was not a consistent finding. In two other cells, spermine (5 mM) was again without effect on basal  $[\text{Ca}^{2+}]_i$  and caused a small inhibi-  
20 tion (about 15%) of the extracellular  $\text{Ca}^{2+}$ -induced increase in  $[\text{Ca}^{2+}]_i$ . In a third cell, neomycin B (5 mM) was without effect on basal  $[\text{Ca}^{2+}]_i$  and did not affect increases in  $[\text{Ca}^{2+}]_i$  elicited by extracellular  $\text{Ca}^{2+}$ . The overall picture that develops from these studies is that spermine and neo-  
25 mycin B are without effect on basal or stimulated levels of cytosolic  $\text{Ca}^{2+}$  in osteoclasts.

The failure of the molecular polycations to affect the  $\text{Ca}^{2+}$ -sensing mechanisms of C-cells or osteoclasts demonstrates the ability to discover or design novel lead  
30 molecules that act specifically on the parathyroid cell  $\text{Ca}^{2+}$  receptor or otherwise modulate one or more functions of the parathyroid cell's normal response to  $[\text{Ca}^{2+}]$ .

Screening of various other molecules is described in detail below and the results summarized in Table 1.

Example 2: Polyamine Screening

Straight chain polyamines (spermine, spermidine, TETA, TEPA, and PEHA) and two derivatives thereof (NPS 381 and NPS 382) were screened as in Example 1. These molecules were all found to mobilize intracellular  $\text{Ca}^{2+}$  in bovine parathyroid cells. Their order of potency is as follows, with the net positive charge listed in parentheses:

<u>Table 3</u>		
	<u>Molecule</u>	<u>EC<sub>50</sub> (in <math>\mu\text{M}</math>)</u>
10	NPS 382 (+8)	50
	NPS 381 (+10)	100
	spermine (+4)	150
	PEHA (+6)	500
15	spermidine (+3)	2000
	TEPA (+5)	2500
	TETA (+4)	8000

Putrescine (+2) and cadaverine (+2) were inactive at a concentration of 2mM.

Another straight-chain polyamine, DADD, behaved somewhat differently from the other polyamines and is described in Example 7.

Example 3: Cyclic Polyamine Screening

Two cyclic polyamines, hexacyclen and NPS 383, were screened as in Example 1. Hexacyclen (+6, EC<sub>50</sub> = 20  $\mu\text{M}$ ) is 7-fold more potent than NPS 383 (+8, EC<sub>50</sub> = 150  $\mu\text{M}$ ). The converse would be expected based solely on net positive charge as the structural characteristic for  $\text{Ca}^{2+}$  receptor activity.

30 Example 4: Aminoglycoside Antibiotic Screening

Six antibiotics were screened as in Example 1. The resulting EC<sub>50</sub>'s for the mobilization of intracellular  $\text{Ca}^{2+}$ , in rank order of potency, were:



Table 4

	<u>Antibiotic</u>	<u>EC<sub>50</sub> (in <math>\mu</math>M)</u>
	neomycin (+6)	10
	gentamicin (+5)	150
5	bekanamycin (+5)	200
	streptomycin (+3)	600

Kanamycin (+4.5) and lincomycin (+1) were without effect at a concentration of 500  $\mu$ M. Within the aminoglycoside series, there is a correlation between net positive charge and potency. However, neomycin is considerably more potent than various polyamines (NPS 381, NPS 382, NPS 383, PEHA) that have an equal or greater positive charge.

Example 5: Peptide and Polyamino Acid Screening

Protamine and polymers of lysine or arginine varying in peptide length were screened for their ability to mobilize intracellular  $\text{Ca}^{2+}$  as in Example 1. The resulting EC<sub>50</sub>'s for the mobilization of intracellular  $\text{Ca}^{2+}$ , in rank order of potency, were:

Table 5

	<u>Peptide (MW in kD)</u>	<u>EC<sub>50</sub> (in mM)</u>
20	polyArg (100)	4
	polyArg (40)	15
	polyLys (27)	30
	protamine (4.8)	75
25	polyArgTyr (22)	200
	polyLys (14)	1000
	polyLys (3.8)	3000

The net positive charge of these polymers increases as the MW increases. Thus, as for the aminoglycosides, there is a direct correlation between net charge and potency among this series of polyamino acids. Protamine is essentially polyArg with a net positive charge of +21.

Example 6: Arylalkylamine Screening

Molecules selected from the class of arylalkylamine toxins derived from the venoms of wasps and spiders were screened as in Example 1.

5 Philanthotoxin-433 (+3) was without effect at a concentration of 500  $\mu\text{M}$ . It is similar in structure to the argiotoxins described below.

Argiotoxin-636 (400  $\mu\text{M}$ ) did not elicit increases in  $[\text{Ca}^{2+}]_i$  but it did potentiate cytosolic  $\text{Ca}^{2+}$  responses to  
10 the subsequent addition of extracellular  $\text{Ca}^{2+}$ . This is a feature common to all molecules that activate the  $\text{Ca}^{2+}$  receptor and is also seen with a variety of extracellular divalent cations. This is considered in more detail in Example 7.

15 In contrast to argiotoxin-636, argiotoxin-659 elicited increases in  $[\text{Ca}^{2+}]_i$  with an  $\text{EC}_{50}$  of 300  $\mu\text{M}$ . Argiotoxin-659 differs from argiotoxin-636 in having a hydroxylated indole moiety rather than a dihydroxyphenyl group. This  
is the only difference in the structure of these two  
20 molecules. Thus, the difference in potency lies in the nature of the aromatic group, not in the polyamine chain which carries the positive charge.

Example 7: Screening of  $\text{Ca}^{2+}$  Channel Blockers

$\text{Ca}^{2+}$  channel blockers, i.e., those molecules which  
25 block influx of extracellular  $\text{Ca}^{2+}$  through voltage-sensitive  $\text{Ca}^{2+}$  channels, were screened as in Example 1. There are three structural classes of  $\text{Ca}^{2+}$  channel blockers: (1) dihydropyridines, (2) phenylalkylamines, and (3) benzothiazipines.

30 None of the dihydropyridines tested (nifedipine, nitrendipine, BAY K 8644, and (-) 202-791 and (+) 202-791) had any effect on basal  $[\text{Ca}^{2+}]_i$  or increases in  $[\text{Ca}^{2+}]_i$  evoked by extracellular  $\text{Ca}^{2+}$  when they were tested at 1  $\mu\text{M}$ . Previous studies showed that parathyroid cells lack  
35 voltage-sensitive  $\text{Ca}^{2+}$  channels, but do have voltage-

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insensitive  $\text{Ca}^{2+}$  channels that are regulated by the  $\text{Ca}^{2+}$  receptor.

The phenylalkylamines examined were verapamil, D-600 (a methoxy-derivative of verapamil), TMB-8, and an analog of TMB-8, NPS 384. The first three molecules were tested at a concentration of 100  $\mu\text{M}$ . The phenylalkylamines behaved differently from other molecules examined. They evoked no change in  $[\text{Ca}^{2+}]_i$  when added to cells bathed in buffer containing a low concentration of extracellular  $\text{Ca}^{2+}$  (0.5 mM). However, verapamil, D-600, and TMB-8 potentiated the mobilization of intracellular  $\text{Ca}^{2+}$  elicited by extracellular divalent cations and they additionally blocked the influx of extracellular  $\text{Ca}^{2+}$ . At intermediate levels of extracellular  $\text{Ca}^{2+}$  (1-1.5 mM), these molecules were capable of evoking a small but robust increase in  $[\text{Ca}^{2+}]_i$  that arose from the mobilization of intracellular  $\text{Ca}^{2+}$ .

The phenylalkylamines act differently than organic polycations like neomycin. The data suggest that verapamil, D-600 and TMB-8 are partial agonists at the  $\text{Ca}^{2+}$  receptor, in contrast to the other molecules examined which are full agonists.

Molecule NPS 384, at a concentration of 300  $\mu\text{M}$ , did not evoke an increase in  $[\text{Ca}^{2+}]_i$  but it blocked influx of extracellular  $\text{Ca}^{2+}$ . Testing at higher concentrations may reveal an ability of this molecule to cause the mobilization of intracellular  $\text{Ca}^{2+}$ .

While the ability of these molecules to block influx is intriguing and not entirely unexpected, it is the ability of these molecules to evoke transient increases in  $[\text{Ca}^{2+}]_i$  (arising from intracellular  $\text{Ca}^{2+}$  mobilization) that is important. Considerable experience with measurements of  $[\text{Ca}^{2+}]_i$  in parathyroid cells shows that transient increases in  $[\text{Ca}^{2+}]_i$  almost invariably result from the mobilization of intracellular  $\text{Ca}^{2+}$  and therefore reflects activation of the  $\text{Ca}^{2+}$  receptor.

The benzothiazepine examined, diltiazem, was similar in all respects to verapamil and D-600 and was also effective at 100  $\mu$ M.

It should be mentioned that with the exception of the phenylalkylamines, all the active molecules tested above evoke increase in  $[Ca^{2+}]_i$  that are of magnitude similar to that evoked by a maximally effective concentration of extracellular  $Ca^{2+}$ . This shows that these molecules are equally efficacious as extracellular divalent cations. This contrasts with the activity of phenylalkylamines, which seem to act only as partial agonists.

Amongst the phenylalkylamines, some interesting structure-activity relationships emerge. Significant is the different potencies of molecules like TMB-8 and NPS 384. TMB-8 potentiated transient increases in  $[Ca^{2+}]_i$  at 100  $\mu$ M whereas NPS 384 fails to do so even at 300  $\mu$ M, yet these molecules carry the same net positive charge. It follows that some other structural feature, unrelated to net charge, imparts greater potency to TMB-8.

#### 20 Example 8: Molecule Screening on Human Parathyroid Cells

Spermine and neomycin were tested for effects on  $[Ca^{2+}]_i$  in human parathyroid cells obtained from glands removed by surgery and prepared as in Example 1. In human parathyroid cells, spermine was found to cause only a small increase in  $[Ca^{2+}]_i$  when tested at a concentration of 300  $\mu$ M.

Neomycin, on the other hand, evoked a large increase in  $[Ca^{2+}]_i$  in human parathyroid cells when tested at a concentration of 20  $\mu$ M. The magnitude of the response elicited by neomycin was equal to that evoked by a maximally effective concentration of extracellular  $Ca^{2+}$ .

#### Example 9: Molecule Screening on Xenopus Oocytes

Oocytes injected with mRNA from human parathyroid cells express the  $Ca^{2+}$  receptor and mobilize intracellular  $Ca^{2+}$  in response to a variety of extracellular inorganic

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di- and trivalent cations. Using this screen allows one to test for an action directly on the  $\text{Ca}^{2+}$  receptor. Oocytes expressing the  $\text{Ca}^{2+}$  receptor also responded to several molecules active on intact parathyroid cells when  
5 screened as follows. Hexacyclen caused the mobilization of intracellular  $\text{Ca}^{2+}$  at a concentration of 135  $\mu\text{M}$ . Neomycin (100  $\mu\text{M}$ ) and NPS 382 (5 mM) were also effective. This offers rather compelling evidence showing that these molecules act on the  $\text{Ca}^{2+}$  receptor or on some other protein  
10 intimately associated with its function.

For example, we have been able to detect  $\text{Ca}^{2+}$  receptor expression in oocytes by measuring  $^{45}\text{Ca}^{2+}$  mobilization. In these experiments, oocytes were injected with bovine parathyroid mRNA or water and, after 72 hours exposed to serum  
15 or 10 mM neomycin. Prior to being stimulated, oocytes were loaded with  $^{45}\text{Ca}^{2+}$ . Stimulation with serum for 20 min resulted in intracellular  $^{45}\text{Ca}^{2+}$  release representing a 45% increase compared to mock challenge with buffer. Challenge with 10 mM neomycin for 20 min. resulted in a 76%  
20 increase in  $^{45}\text{Ca}^{2+}$  release. The assay is sensitive enough for use in cloning the  $\text{Ca}^{2+}$  receptor, and has the advantage of a higher throughput than the electrophysiological measurement of  $\text{Ca}^{2+}$  activated  $\text{Cl}^-$  current.

In another example, human osteoclastoma tissue was  
25 obtained from bone biopsy tissue. Oocytes injected with mRNA isolated from this tissue were challenged with 30 mM  $\text{Ca}^{2+}$ . Controls did not respond while 8 of 12 oocytes injected with osteoclastoma mRNA responded appropriately (Fig. 34). These experiments provide the first evidence  
30 that the  $\text{Ca}^{2+}$  response of osteoclasts to extracellular  $\text{Ca}^{2+}$  is in fact genetically encoded. The results also indicate that the osteoclast  $\text{Ca}^{2+}$  receptor may be cloned by expression in Xenopus oocytes.

#### Example 10: Molecule Screening on Rat Osteoclasts

35 However, the different sensitivities of parathyroid cells and osteoclasts to extracellular  $\text{Ca}^{2+}$  suggest that

their  $\text{Ca}^{2+}$  receptors are different. While parathyroid cells respond to extracellular  $\text{Ca}^{2+}$  concentrations between 0.5 and 3 mM, osteoclasts respond only when the level of extracellular  $\text{Ca}^{2+}$  increases beyond 5 mM. This rather high concentration of  $\text{Ca}^{2+}$  is nonetheless physiological for osteoclasts; as they resorb bone, the local concentration of extracellular  $\text{Ca}^{2+}$  may reach levels as high as 30 mM.

Molecule screening with osteoclasts was performed as follows. Osteoclasts were obtained from the long bones of neonatal rats.  $[\text{Ca}^{2+}]_i$  was measured in single cells using the fluorimetric indicator indo-1. Spermine, spermidine, neomycin, and verapamil were tested, and none of these caused any large increase in  $[\text{Ca}^{2+}]_i$  in osteoclasts (although small responses were detected).

At a concentration of 1 mM, spermidine caused a small increase in  $[\text{Ca}^{2+}]_i$  (about 10% of that evoked by a maximal concentration of extracellular  $\text{Ca}^{2+}$ ). Neither Neomycin (10 mM) nor Spermine (10 or 20 mM) caused increases in  $[\text{Ca}^{2+}]_i$  in rat osteoclasts. Neomycin (10 mM) did not block the increase in  $[\text{Ca}^{2+}]_i$  elicited by the subsequent addition of 25 mM extracellular  $\text{Ca}^{2+}$ . Pretreatment with spermine (20 mM) however, did depress the response to extracellular  $\text{Ca}^{2+}$ . Verapamil (100  $\mu\text{M}$ ) caused no detectable increase in  $[\text{Ca}^{2+}]_i$  but it did block the response to extracellular  $\text{Ca}^{2+}$ .

Comparisons between osteoclasts and parathyroid cells show that molecules active on the latter are relatively ineffective in osteoclasts. This demonstrates that drugs that target a specific  $\text{Ca}^{2+}$  receptor without affecting those receptor types present on other  $\text{Ca}^{2+}$ -sensing cells are readily developed. Similarly, drugs active at two or more such  $\text{Ca}^{2+}$  receptors may also be developed.

#### Other $\text{Ca}^{2+}$ Receptor Examples

The following examples demonstrate that, just as there are subtypes of receptors for molecular ligands, so too do there appear to be subtypes of  $\text{Ca}^{2+}$  receptors that

can be differentially affected by drugs. The parathyroid cell  $\text{Ca}^{2+}$  receptor senses levels of extracellular  $\text{Ca}^{2+}$  around 1.5 mM whereas the  $\text{Ca}^{2+}$  receptor on the osteoclast responds to levels around 10 mM (Fig. 22). Neomycin or spermine, which activate the parathyroid cell  $\text{Ca}^{2+}$  receptor, fail to affect the  $\text{Ca}^{2+}$  receptors on C-cells or osteoclasts (Figs. 21 and 23). These data constitute the first evidence for pharmacologically distinct subtypes of  $\text{Ca}^{2+}$  receptors and these data are being used to design and develop drugs that act selectively on a particular type of  $\text{Ca}^{2+}$  receptor. Indeed, testing of lead molecules demonstrate such cell-specific effects. For example, NPS 449, which elicits increases in  $[\text{Ca}^{2+}]_i$  in osteoclasts is without effect on  $[\text{Ca}^{2+}]_i$  in parathyroid cells. Conversely, NPS 447, which activates the parathyroid cell  $\text{Ca}^{2+}$  receptor, is effective in activating the osteoclast  $\text{Ca}^{2+}$  receptor only at concentrations 10-fold higher. Finally, agatoxin 489, although not very potent in activating the C-cell  $\text{Ca}^{2+}$  receptor ( $\text{EC}_{50} = 150 \mu\text{M}$ ), is a quite potent activator of the parathyroid cell  $\text{Ca}^{2+}$  receptor ( $\text{EC}_{50} = 3 \mu\text{M}$ ). The lead molecules presently under development will affect selectively the activity of a specific type of  $\text{Ca}^{2+}$ -sensing cell in vivo.

Drugs with less specificity might not necessarily be therapeutically undesirable. Thus, depressing osteoclast activity and stimulating calcitonin secretion are two different approaches to inhibiting bone resorption. Drugs that target the  $\text{Ca}^{2+}$  receptors on both of these cells might be very effective therapies for osteoporosis. Because PTH is also involved in regulating bone metabolism, drugs acting on the parathyroid cell  $\text{Ca}^{2+}$  receptor may also be useful in the treatment and/or prevention of osteoporosis.

Results of some test molecules are shown below. In Table 6, the comparative activity of calcimimetic molecules is shown. Bovine parathyroid cells and C-cells (rMT 6-23 cells) were loaded with fura-2, and rat osteoclasts with indo-1 and the potency of the indicated molecules to

mobilize intracellular  $\text{Ca}^{2+}$  determined by constructing cumulative concentration-response curves. Molecules listed as "inactive" did not alter  $[\text{Ca}^{2+}]$ ; when tested at a concentration of 1 mM.

Table 6

COMPOUND	EC <sub>50</sub> ( $\mu\text{M}$ )		
	PARATHYROID	OSTEOCLAST	C-CELL
NPS 568 (EE)	0.78	200	>300
NPS 568 (LE)	30	—	—
NPS 467 (EE)	2	>100	—
NPS 467 (LE)	>30	—	—
NPS 017	6	inactive	150
NPS 447	9	150	—
NPS 456*	15	200	>100
NPS 015	22	—	inactive
NPS 109	40	>300	5
NPS 449	inactive	150	—
NPS 468*	30	250	—
spermine	150	inactive	inactive
neomycin	40	inactive	inactive

\*racemic mixture; "inactive" is defined as causing no increase in cytosolic  $\text{Ca}^{2+}$  at a concentration of 1-5 mM; EE is early eluting; LE is late eluting.

#### Example 11: Lead Molecules for Parathyroid $\text{Ca}^{2+}$ Receptor

Structure-activity studies using polyamines and arylalkylamines led to the testing of molecules structurally akin to NPS 456. NPS 456 is a potent activator of the parathyroid cell  $\text{Ca}^{2+}$  receptor. This molecule is notable because it possess only one positive charge yet is much more potent than many polybasic molecules. Brief (2 min) pretreatment with PMA shifts the concentration-response curve for NPS 456 to the right. This indicates that NPS 456 acts through the same mechanism used by extracellular  $\text{Ca}^{2+}$ . NPS 456 evokes the mobilization of intracellular  $\text{Ca}^{2+}$  in Xenopus oocytes expressing the

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parathyroid cell  $\text{Ca}^{2+}$  receptor, which demonstrates a direct action on the  $\text{Ca}^{2+}$  receptor (Fig. 33). Moreover, NPS 456 contains a chiral carbon, and therefore exists in two isomeric forms. Both isomers have been synthesized and examined for activity. The R-isomer, NPS 447, is 12-times more potent than the S-isomer, NPS 448 (Fig. 28). This is the first demonstration that a  $\text{Ca}^{2+}$  receptor can recognize an organic molecule in a stereospecific manner.

Because NPS 447 is a structurally simple molecule with selective and potent effects on the parathyroid cell  $\text{Ca}^{2+}$  receptor, structure-activity studies around this lead molecule are simple. The aim of these studies is to generate an array of related molecules with various characteristics from which the final development candidate can be selected. This effort has already revealed some of the structural domains of NPS 447 that contribute to activity and potency. For example, the novel compound NPS 459 is an analog of NPS 447 that is smaller ( $\text{MW} < 240$ ) yet nearly as potent as the parent molecule, whereas several other analogs are relatively inactive. The most interesting molecules from this analog project can be put into in vivo testing for effects on PTH secretion and serum  $\text{Ca}^{2+}$  levels (see Examples 15, 16, 17, 18 and 23).

The novel compound NPS 467 is an even smaller molecule than NPS 447 yet the former is about 3-fold more potent than the latter in causing the mobilization of intracellular  $\text{Ca}^{2+}$  in parathyroid cells. Like NPS 456, NPS 467 is a racemic mixture. It is anticipated that Resolution of NPS 467 into its enantiomers provides an isomer of even greater potency than the racemic mixture (see Example 16). NPS 551 is another novel compound as potent as NPS 467 in causing the mobilization of intracellular  $\text{Ca}^{2+}$  in parathyroid cells. NPS 551 is a racemic mixture and it is anticipated that the resolution of NPS 551 into its enantiomers will result in an isomer that is more potent than the racemic mixture. Further structure-activity studies on molecules related to NPS 447, NPS 467, NPS 551 and NPS

568 are expected to yield pure isomers with greater potency than these molecules in their racemate forms.

Results obtained with NPS 456 (Fig. 33) show that it elicits oscillatory increases in  $\text{Cl}^-$  current at concentrations of 100  $\mu\text{M}$ . NPS 456 is the most potent molecule activate on Xenopus oocytes expressing the parathyroid cell  $\text{Ca}^{2+}$  receptor. The results obtained in this expression system with neomycin and NPS 456 demonstrate that these molecules act directly on the  $\text{Ca}^{2+}$  receptor.

10 Example 12: Osteoclast  $\text{Ca}^{2+}$  Receptor Lead Molecules

The strategy used for elucidating the mechanism of action of extracellular  $\text{Ca}^{2+}$  on the osteoclast was similar to that proven effective in parathyroid cells. The first experiments examined the effects of  $\text{La}^{3+}$  on  $[\text{Ca}^{2+}]_i$  in single rat osteoclasts loaded with the fluorimetric indicator indo-1. As described above, trivalent cations like  $\text{La}^{3+}$  are impermeant and block  $\text{Ca}^{2+}$  influx. Low micromolar concentrations of  $\text{La}^{3+}$  partially depressed extracellular  $\text{Ca}^{2+}$ -induced increases in  $[\text{Ca}^{2+}]_i$  (Fig. 29). The demonstration of a  $\text{La}^{3+}$ -resistant increase in  $[\text{Ca}^{2+}]_i$  provides evidence for the mobilization of intracellular  $\text{Ca}^{2+}$ . The results of these experiments parallel those obtained in parathyroid cells and suggest that similar mechanisms are used by extracellular  $\text{Ca}^{2+}$  to regulate  $[\text{Ca}^{2+}]_i$  in both cell types.

Another series of experiments showed that extracellular  $\text{Mn}^{2+}$  evoked transient increases in  $[\text{Ca}^{2+}]_i$  (Fig. 30(a)) that persisted in the absence of extracellular  $\text{Ca}^{2+}$  (Fig. 30B). These results are likewise indicative of the mobilization of intracellular  $\text{Ca}^{2+}$ . Although  $\text{Mn}^{2+}$  can enter some cells, it is unlikely to do so in the osteoclast because  $\text{Mn}^{2+}$  quenches the fluorescence of indo-1. Thus, if  $\text{Mn}^{2+}$  penetrated intracellularly, a decrease, not an increase in the fluorescent signal would be observed.

35 The results obtained with a variety of di- and tri-valent cations are all consistent with the presence of a

Ca<sup>2+</sup> receptor on the surface of the osteoclast that is coupled to the mobilization of intracellular Ca<sup>2+</sup> and influx of extracellular Ca<sup>2+</sup> through voltage-insensitive channels. Results show evidence for genetic material in human osteoclasts that encodes a Ca<sup>2+</sup> receptor protein (see below). Transient increases in [Ca<sup>2+</sup>]<sub>i</sub> resulting from the mobilization of intracellular Ca<sup>2+</sup>, are sufficient to inhibit osteoclastic bone resorption in vitro. Thus, as with the parathyroid cell, activation of the Ca<sup>2+</sup> receptor appears to be a viable means of inhibiting the activity of osteoclasts.

NPS 449 is presently the lead molecule for calcimimetic drugs on this receptor. It is a small molecule (MW < 425) and it mobilizes intracellular Ca<sup>2+</sup> in rat osteoclasts with a EC<sub>50</sub> of 200 μM (Figs. 31A and 31B). Although the potency of NPS 449 is relatively low, it has a simple structure with only one positive charge and is expected to have desirable pharmacodynamic and pharmacokinetic properties.

NPS 449 was examined for its ability to inhibit bone resorption in vitro. This was done by morphometric analysis of pit formation on thin slices of bovine cortical bone using scanning electron microscopy. Rat osteoclasts were incubated for 24 hours in slices of bone in the presence or absence of various concentrations of NPS 449. NPS 449 caused a concentration-dependent inhibition of bone resorption with an IC<sub>50</sub> of 10 μM. The anticipated results provide the first demonstration that molecules acting at this novel site can inhibit osteoclastic bone resorption. More potent analogs of NPS 449 will be generated using synthetic chemistry and will be tested and assayed using the methods described herein.

#### Example 13: C-Cell Ca<sup>2+</sup> Receptor Lead Molecules

Activation of the C-cell Ca<sup>2+</sup> receptor stimulates the secretion of calcitonin which then acts on osteoclasts to inhibit bone resorption. Calcimimetic drugs selectively

affecting C-cells are useful in the treatment of osteoporosis.

The mobilization of intracellular  $\text{Ca}^{2+}$  is used as a functional index of  $\text{Ca}^{2+}$  receptor activity. The screening effort in C-cells is facilitated by the availability of cultured cell lines expressing the C-cell phenotype (*e.g.*, rat medullary thyroid carcinoma cells; rMTC 6-23 cells). Selected for initial study were three arylalkylamine molecules. Two are naturally occurring (agatoxin 489 and agatoxin 505) and the other (NPS 019) is a synthetic agatoxin analog. Agatoxin 505 was found to block extracellular  $\text{Ca}^{2+}$ -induced increases in  $[\text{Ca}^{2+}]_i$ , with an  $\text{IC}_{50}$  of 3  $\mu\text{M}$ . The inhibitory effect resulted from a block of the L-type voltage-sensitive  $\text{Ca}^{2+}$  channel present in these cells. In contrast, agatoxin 489 was found to mobilize intracellular  $\text{Ca}^{2+}$  in rMTC cells with an  $\text{EC}_{50}$  of 150  $\mu\text{M}$ . This was the first organic molecule discovered that was found to activate the C-cell  $\text{Ca}^{2+}$  receptor. The synthetic analog, NPS 019, was even more potent and mobilized intracellular  $\text{Ca}^{2+}$  with an  $\text{EC}_{50}$  of 5  $\mu\text{M}$  (Fig. 32). It is significant that the only structural difference between NPS 019 and agatoxin 489 is the presence or absence of an hydroxyl group. The fact that such subtle differences in structure affect profoundly the potency of molecules indicates a structurally specific binding site on the  $\text{Ca}^{2+}$  receptor. This, in turn, encourages the view that very potent and selective activators of  $\text{Ca}^{2+}$  receptors can be developed.

NPS 019, which is a small molecule ( $\text{MW} < 500$ ), is a lead molecule for development of calcimimetics of the C-cell  $\text{Ca}^{2+}$  receptor and can be tested for its ability to stimulate calcitonin secretion *in vitro*. Subsequent *in vivo* testing will then determine the ability of this molecule to stimulate calcitonin secretion and inhibit bone resorption. These *in vivo* studies will be performed in rats. The results obtained in these studies, which are anticipated to be positive, will provide the first evi-

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dence showing that a small organic molecule acting on a novel receptor can stimulate calcitonin secretion and depress bone resorption.

Example 14: Calcilytic Activity of NPS 021 on Parathyroid  
5 Cells

For a compound to be considered a calcilytic, it must block the effects of extracellular  $\text{Ca}^{2+}$  or a calcimimetic compound on an extracellular  $\text{Ca}^{2+}$ -sensing cell. An example of a calcilytic compound is NPS 021, the structure of which is provided in Fig. 1. In bovine parathyroid cells loaded with fura-2, NPS 021 blocks increases in  $[\text{Ca}^{2+}]_i$  elicited by extracellular  $\text{Ca}^{2+}$ . The  $\text{IC}_{50}$  of NPS 021 for blocking this response is about 200  $\mu\text{M}$  and, at concentrations around 500  $\mu\text{M}$ , the increase in  $[\text{Ca}^{2+}]_i$  evoked by extracellular  $\text{Ca}^{2+}$  is abolished. Significantly, NPS 021 does not by itself cause any change in  $[\text{Ca}^{2+}]_i$  when tested at low  $[\text{Ca}^{2+}]$  (0.5 mM; Fig. 37).

Example 15: NPS 467 Lowers Serum Ionized Calcium

Compounds shown to activate the bovine parathyroid cell  $\text{Ca}^{2+}$  receptor in vitro were tested for hypocalcemic activity in vivo. Male Sprague-Dawley rats (200 g) were maintained on a low calcium diet for one week prior to receiving test substance or vehicle as control. Blood was collected from the tail vein three hours after the intraperitoneal administration of NPS 467. Ionized  $\text{Ca}^{2+}$  in whole blood or serum was measured with a Ciba-Corning 634 Analyzer according to the instructions provided with the instrument. Serum total calcium, albumin and phosphate were measured by techniques well-known in the art.

NPS 467 caused a dose-dependent reduction in serum or whole blood  $\text{Ca}^{2+}$  (Fig. 38). The fall in blood  $\text{Ca}^{2+}$  at this time was paralleled by a proportional fall in the levels of blood total calcium. There was no change in serum albumin or phosphate levels at any of the doses examined. In preliminary studies, NPS 467, at doses effective in

lowering blood  $\text{Ca}^{2+}$ , caused a dose-dependent reduction in circulating levels of PTH (Fig. 39). The hypocalcemic effect of NPS 467 was maximal within three hours and returned toward control levels after 24 hours (Fig. 40).

5 NPS 467 (the EE isomer; see Example 16) was also effective in lowering serum ionized  $\text{Ca}^{2+}$  in rats maintained on a normal, calcium-replete diet. A single dose of NPS 467 (EE isomer, 10 mg/kg i.p.) caused a rapid fall in serum levels of ionized  $\text{Ca}^{2+}$  which were maximal by 1 hr  
10 (22% decrease from the control level) and remained depressed at or near this level for up to 6 hours.

Example 16: NPS 467 Lowers Serum Ionized Calcium in a Stereospecific Manner

NPS 467 is a racemic mixture. Resolution of NPS 467  
15 into its two enantiomers was achieved by separation on a chiral column. The EE-isomer (for "early eluting", see Example 21) was about 100-fold more potent than the LE-isomer (for "late-eluting") in activating the bovine parathyroid cell  $\text{Ca}^{2+}$  receptor in vitro as assessed by the  
20 ability of the enantiomers to evoke increases in the  $[\text{Ca}^{2+}]_i$  in parathyroid cells (Fig. 41). Likewise, similar resolution of the novel compound NPS 568 into its enantiomers showed that the EE-isomer was 40-fold more potent than the LE-isomer in causing the mobilization of intracellular  $\text{Ca}^{2+}$   
25 in bovine parathyroid cells (see Table 6, supra).

The isomers of NPS 467 were examined for effects on serum  $\text{Ca}^{2+}$  as in Example 15. Consistent with the in vitro results, the EE-isomer or NPS 467 proved to be more potent than the LE-isomer in lowering serum  $\text{Ca}^{2+}$  in vivo (Fig. 42;  
30 each compound was tested at a concentration of 5 mg/kg body weight).

Example 17: NPS 467 Lowers Serum Ionized Calcium in an in vivo Model of Secondary Hyperparathyroidism

An accepted and widely used animal model of secondary  
35 hyperparathyroidism arising from chronic renal failure is

the 5/6 nephrectomized rat. Animals receiving such surgery become initially hypocalcemic and, to maintain serum  $\text{Ca}^{2+}$  levels, there is a compensatory hyperplasia of the parathyroid glands and elevated levels of circulating PTH.

- 5 Male Sprague-Dawley rats (250 g) received a 5/6 nephrectomy and were allowed to recover for 2 weeks. At this time they were normocalcemic (due to elevated levels of serum PTH). The administration of NPS 467 (EE isomer; 10 mg/kg i.p.) caused a rapid (within 2 hours) fall in serum  
10 ionized  $\text{Ca}^{2+}$  levels to 83% of controls in an animal model of secondary hyperparathyroidism. This suggests that compounds of this sort will effectively depress PTH secretion in patients with secondary hyperparathyroidism and hyperplastic parathyroid glands.

15 Example 18: NPS 467 Fails to Lower Serum Ionized Calcium Levels in Parathyroidectomized Animals

- To determine the primary target tissue upon which NPS 467 acts to cause a hypocalcemic response, the parathyroid glands in rats were surgically removed. Animals receiving  
20 a total parathyroidectomy become hypocalcemic and are largely dependent upon dietary calcium to maintain serum  $\text{Ca}^{2+}$  homeostasis. Parathyroidectomized animals had serum ionized  $\text{Ca}^{2+}$  levels of 0.92 mM which fell gradually to 0.76 mM after 6 hours of fasting. The administration of a  
25 single dose of NPS 467 EE (10 mg/kg i.p.) did not cause any change in serum ionized  $\text{Ca}^{2+}$  levels over a period of 6 hours. These results demonstrate that intact parathyroid glands are required for the hypocalcemic effects of NPS 467 EE. The data additionally demonstrate that NPS  
30 467 EE can target the parathyroid glands in vivo. The results are consistent with the view that NPS 467 EE acts on the parathyroid cell  $\text{Ca}^{2+}$  receptor in vivo to depress secretion of PTH and thereby cause serum levels of ionized  $\text{Ca}^{2+}$  to fall.

Example 19: NPS 467 Increases Intracellular Calcium in Human Parathyroid Glands

Dissociated parathyroid cells were prepared from a parathyroid adenoma obtained by surgery from a patient with primary hyperparathyroidism. The cells were loaded with fura-2 and  $[Ca^{2+}]_i$  measured as described above. Both NPS 467 EE and NPS 568 EE caused concentration-dependent increase in  $[Ca^{2+}]_i$ . The  $EC_{50}$ 's for NPS 467 EE and NPS 568 EE were 20 and 3  $\mu M$ , respectively. Both these compounds are thus able to increase  $[Ca^{2+}]_i$  in pathological human tissue and would thus be expected to decrease serum levels of PTH and  $Ca^{2+}$  in patients with primary hyperparathyroidism.

Example 20: Mechanism of Action of NPS 467 at the Parathyroid Cell Calcium Receptor

Dissociated bovine parathyroid cells were used to further explore the mechanism of action of NPS 467 at the receptor level. In the presence of 0.5 mM extracellular  $Ca^{2+}$ , NPS 467 EE caused a rapid and transient increase in  $[Ca^{2+}]_i$  which persisted in the presence of 1  $\mu M$   $La^{3+}$  and was partially depressed by pretreatment with PMA (100 nM for 2 min.). Moreover, NPS 467 (EE isomer, 30  $\mu M$ ) caused a rapid increase in  $Cl^-$  conductance in Xenopus oocytes injected with parathyroid cell mRNA. All these results are consistent with an action of NPS 467 on the  $Ca^{2+}$  receptor. However, the cytosolic  $Ca^{2+}$  response to NPS 467 was abolished when parathyroid cells were suspended in  $Ca^{2+}$ -free buffer. This suggests that NPS 467 cannot, by itself, cause the mobilization of intracellular  $Ca^{2+}$ . It does, however, elicit responses in parathyroid cells and in oocytes when a small amount of extracellular  $Ca^{2+}$  is present. This suggests that partial occupancy of the  $Ca^{2+}$ -binding site is required for NPS 467 to elicit a response. To test this hypothesis, parathyroid cells were suspended in  $Ca^{2+}$ -free buffer and exposed to a submaximal concentration of neomycin. Neomycin was used because it mimics, in



nearly all respects, the effects of extracellular  $\text{Ca}^{2+}$  on parathyroid cells and on Xenopus oocytes expressing the parathyroid cell  $\text{Ca}^{2+}$  receptor. The addition of 10  $\mu\text{M}$  neomycin did not by itself cause an increase in  $[\text{Ca}^{2+}]_i$  under these conditions. However the subsequent addition of NPS 467 EE (30  $\mu\text{M}$ ) now elicited a transient increase in  $[\text{Ca}^{2+}]_i$  which, because there was no extracellular  $\text{Ca}^{2+}$  present, must have come from the mobilization of intracellular  $\text{Ca}^{2+}$ . When cells bathed in  $\text{Ca}^{2+}$ -free buffer were exposed to 30  $\mu\text{M}$  NPS 467 there was no increase in  $[\text{Ca}^{2+}]_i$ . This concentration of NPS 467 is maximally effective in increasing  $[\text{Ca}^{2+}]_i$  when extracellular  $\text{Ca}^{2+}$  (0.5 mM) is present. However, the subsequent addition of 10  $\mu\text{M}$  neomycin now evoked a transient increase in  $[\text{Ca}^{2+}]_i$ . Presumably, neomycin binds to the same site as extracellular  $\text{Ca}^{2+}$  and can functionally substitute for it. Using a submaximal concentration, which by itself causes no response, achieves partial occupancy of the  $\text{Ca}^{2+}$ -binding site and allows activation of the  $\text{Ca}^{2+}$  receptor by NPS 467.

Additional studies to further define the mechanism of action of NPS 467 were performed. The cells were once again suspended in  $\text{Ca}^{2+}$ -free buffer to insure that any observed increase in  $[\text{Ca}^{2+}]_i$  resulted from the mobilization of intracellular  $\text{Ca}^{2+}$ . In these experiments, however, a maximally effective concentration (100  $\mu\text{M}$ ) of neomycin was used. In the absence of extracellular  $\text{Ca}^{2+}$ , 100  $\mu\text{M}$  neomycin evoked a rapid and transient increase in  $[\text{Ca}^{2+}]_i$ . The subsequent addition of 30  $\mu\text{M}$  NPS 467 EE did not cause an increase in  $[\text{Ca}^{2+}]_i$ . In the converse experiment, 30  $\mu\text{M}$  NPS 467 EE was added before 100  $\mu\text{M}$  neomycin. As expected, NPS 467 EE did not cause any increase in  $[\text{Ca}^{2+}]_i$ . It did not, however, affect the increase in  $[\text{Ca}^{2+}]_i$  evoked by the subsequent addition of 100  $\mu\text{M}$  neomycin. These results, obtained with maximally effective concentrations of NPS 467 and neomycin, suggest that these two compounds do not act at the same site. Rather, the results can be suffi-

ciently explained by postulating two separate sites on the  $\text{Ca}^{2+}$  receptor, one to which extracellular  $\text{Ca}^{2+}$  and neomycin bind, and another to which NPS 467 and structurally related compounds (such as NPS 568) bind. Ligand binding to the former site can result in full activation of the  $\text{Ca}^{2+}$  receptor whereas ligand binding to the latter site can only occur and/or be functionally relevant when the extracellular  $\text{Ca}^{2+}$ -binding site is occupied to some as yet undefined degree. It is possible that ligand binding to the extracellular  $\text{Ca}^{2+}$ -binding site exposes a previously occluded binding site for NPS 467. It appears that the NPS 467-binding site is an allosteric site that augments receptor activation in response to ligand binding at the extracellular  $\text{Ca}^{2+}$ -binding site.

The data demonstrate that the parathyroid cell  $\text{Ca}^{2+}$  receptor possesses at least two distinct sites for organic ligands. One site binds the physiological ligand, extracellular  $\text{Ca}^{2+}$ , and certain organic polycations like neomycin. Binding to this site result in full activation of the  $\text{Ca}^{2+}$  receptor, an increase in  $[\text{Ca}^{2+}]_i$ , and the inhibition of PTH secretion. NPS 467 defines a previously unrecognized binding site on the  $\text{Ca}^{2+}$  receptor. Binding to this site can only occur and/or results in full activation of the  $\text{Ca}^{2+}$  receptor when the extracellular  $\text{Ca}^{2+}$ -binding site is partially occupied. Ligands acting at either site are effective in suppressing serum  $\text{Ca}^{2+}$  levels in vivo.

#### Example 21: Preparation of NPS 467

In a 250 ml round bottom flask, 10.0 g (100 mmoles) 3'-methoxy acetophenone and 13.5 g (100 mmoles) 3-phenylpropylamine were mixed and treated with 125 mmoles (35.5 g) titanium(IV) isopropoxide. The reaction mixture was stirred 30 minutes at room temperature under a nitrogen atmosphere. After this time 6.3 g (100 mmoles) sodium cyanoborohydride in 100 ml ethanol was added dropwise over the course of 2 minutes. The reaction was stirred room temperature under nitrogen for 16 hours. After this time

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the reaction mixture was transferred to a 2 L separatory funnel with 1.5 L ethyl ether and 0.5 L water. The phases were equilibrated and the ether layer removed. The remaining aqueous phase was thoroughly extracted with four  
5 1 L portions of ether. The washes were combined, dried over anhydrous potassium carbonate and reduced to a clear, light amber oil.

TLC analysis of this material on silica using chloroform-methanol-isopropylamine (100:5:1) showed product at  
10 Rf 0.65 with traces of the two starting materials at Rf 0.99 (3' methoxy acetophenone) and Rf 0.0 (3-phenylpropylamine).

The reaction mixture was chromatographed through silica (48 x 4.6 cm) using a gradient of chloroform-methanol-isopropylamine (99:1:0.1) to (90:10:0.1) which  
15 yielded 13.66 g of purified NPS 467. This material was dissolved in Hexane-isopropanol (99:1) containing 0.1% diethylamine to yield a solution with a concentration of 50 mg/ml. Chiral resolution was accomplished by chromatography of 4 ml of this solution (200 mg, maximum to  
20 achieve separation) through ChiralCel OD (25 x 2 cm) using 0.7% isopropylamine, 0.07% diethylamine in hexane at 100 ml/min, monitoring optical density at 260 nm. Under these conditions (with injections of 100 mg material) the early  
25 eluting isomer (NPS 467EE) began to emerge from the column at ~ 26 min, the late eluting isomer (NPS 467LE) began to emerge at ~34 minutes. Baseline resolution was accomplished with under these conditions. Each optical isomer (free base) was converted to the corresponding hydrochloride salt by dissolving 3 g of the free base in 100 ml  
30 ethanol and treating with it with 100 ml water containing 10 molar equivalents HCl. Lyophilization of this solution yielded a white solid.

#### Example 22: Preparation of NPS 568

35 NPS 568 was prepared using the methods described in Example 21 substituting an equivalent amount of 3-(2-

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chlorophenyl)propylamine for 3-phenylpropylamine. It was found that allowing the mixture of 3'-methoxyacetophenone, 3-(2-chlorophenyl)propylamine and titanium(IV) isopropoxide to stir for 5 hours prior to treatment with  $\text{NaCNBH}_3/\text{EtOH}$  resulted in significantly greater yield (98%).

Example 23: NPS 467 Lowers Serum Ionized Calcium When Administered Orally

Rats (male, Sprague-Dawley, 250-300 g) were fed standard rat chow and fasted overnight prior to the experiment. NPS 467 (EE isomer) was suspended in corn oil and administered as a single oral dose through a gavage needle. Three hours later a sample of blood was taken from the tail vein and assessed for ionized  $\text{Ca}^{2+}$  levels. Fig. 44 shows that NPS 467 EE caused a dose-dependent reduction in serum levels of ionized  $\text{Ca}^{2+}$  when administered orally.

Other embodiments are within the following claims.

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Claims:

1. A pharmaceutical composition comprising a molecule which mimics the activity of extracellular  $\text{Ca}^{2+}$  by evoking an increase in  $[\text{Ca}^{2+}]_i$  in a cell, or blocks an  
5 increase in  $[\text{Ca}^{2+}]_i$  evoked by extracellular  $\text{Ca}^{2+}$ , said molecule having an  $\text{EC}_{50}$  of less than or equal to 5  $\mu\text{M}$ , wherein said molecule is not protamine.
2. The pharmaceutical composition of claim 1 wherein said increase in  $[\text{Ca}^{2+}]_i$  in a cell is transient, having a  
10 duration of less than thirty seconds.
3. The pharmaceutical composition of claim 1 or 2 wherein said increase in  $[\text{Ca}^{2+}]_i$  is rapid, occurring within thirty seconds.
4. The pharmaceutical composition of claim 1 wherein  
15 said molecule evokes a sustained increase in  $[\text{Ca}^{2+}]_i$ , having a duration greater than thirty seconds.
5. The pharmaceutical composition of claim 1 wherein said molecule further evokes an increase in inositol-1,4,5-trisphosphate or diacylglycerol.
- 20 6. The pharmaceutical composition of claim 5 wherein said increase in said inositol-1,4,5-trisphosphate or diacylglycerol occurs within 60 seconds.
7. The pharmaceutical composition of claim 2 wherein said transient increase is diminished by pretreatment of  
25 said cell for less than 10 minutes with an activator of protein kinase C.
8. The pharmaceutical composition of claim 7 wherein said activator of protein kinase C is selected from the group consisting of phorbol myristate acetate, mezerein  
30 and (-)-indolactam V.

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9. The pharmaceutical composition of claim 1 wherein said molecule inhibits dopamine- or isoproterenol-stimulated cyclic AMP formation.

10. The pharmaceutical composition of claim 2  
5 wherein said transient increase in  $[Ca^{2+}]_i$  is reduced by pretreatment of said cell for ten minutes with 10 mM sodium fluoride.

11. The pharmaceutical composition of claim 1  
10 wherein said cell is a parathyroid cell and said molecule inhibits parathyroid hormone secretion from said cell.

12. The pharmaceutical composition of claim 1  
wherein said molecule elicits an increase in  $Cl^-$  conductance in a Xenopus oocyte injected with mRNA from a cell  
15 selected from the group consisting of parathyroid cells, bone osteoclasts, juxtaglomerular kidney cells, proximal tubule kidney cells, keratinocytes, parafollicular thyroid cells and placental trophoblasts.

13. The pharmaceutical composition of claim 1  
20 wherein said molecule mobilizes intracellular  $Ca^{2+}$  to cause said increase in  $[Ca^{2+}]_i$ .

14. The pharmaceutical composition of claim 1  
wherein said cell is a C-cell or an osteoclast and said molecule inhibits bone resorption in vivo.

15. The pharmaceutical composition of claim 1  
25 wherein said cell is a C-cell and said molecule stimulates calcitonin secretion in vitro or in vivo.

16. The pharmaceutical composition of claim 1  
wherein said molecule is either a calcimimetic or calcilytic having an  $EC_{50}$  or  $IC_{50}$  at a  $Ca^{2+}$  receptor of less than  
30 or equal to 5  $\mu M$ .

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17. The pharmaceutical composition of claim 1 wherein said molecule has an  $EC_{50}$  less than or equal to 5  $\mu M$  at one or more but not all cells chosen from the group consisting of parathyroid cells, bone osteoclasts, juxta-glomerular kidney cells, proximal tubule kidney cells, keratinocytes, parafollicular thyroid cells and placental trophoblasts.

18. The pharmaceutical composition of claim 1 or 16 or 17 wherein said molecule has an  $EC_{50}$  or  $IC_{50}$  less than or equal to 1  $\mu M$ .

19. The pharmaceutical composition of claim 1 or 16, or 17 wherein said molecule has an  $EC_{50}$  or  $IC_{50}$  less than or equal to 100 nanomolar.

20. The pharmaceutical composition of claim 1 or 16, or 17 wherein said molecule has an  $EC_{50}$  or  $IC_{50}$  less than or equal to 10 nanomolar.

21. The pharmaceutical composition of claim 1 or 16, wherein said molecule has an  $EC_{50}$  or  $IC_{50}$  less than or equal to 1 nanomolar.

22. The pharmaceutical composition of claim 1, 16 or 17 wherein said molecule is positively charged at physiological pH.

23. The pharmaceutical composition of claim 22 wherein said molecule is selected from the group consisting of branched or cyclic polyamines, positively charged polyamino acids, and arylalkylamines.

24. The pharmaceutical composition of claim 23 wherein said branched polyamine has the formula

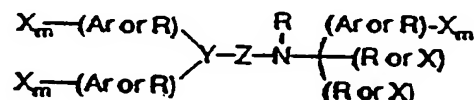


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wherein k is an integer from 1 to 10,  
each j is the same or different and is an integer  
from 2 to 20, and

each R<sub>i</sub> is the same or different and is selected from  
the group consisting of hydrogen and -(CH<sub>2</sub>)<sub>j</sub>-NH<sub>2</sub>, wherein j  
is as defined above, and wherein at least one R<sub>i</sub> is not  
hydrogen.

25. The pharmaceutical composition of claim 1, 16 or  
17 wherein said molecule has the formula

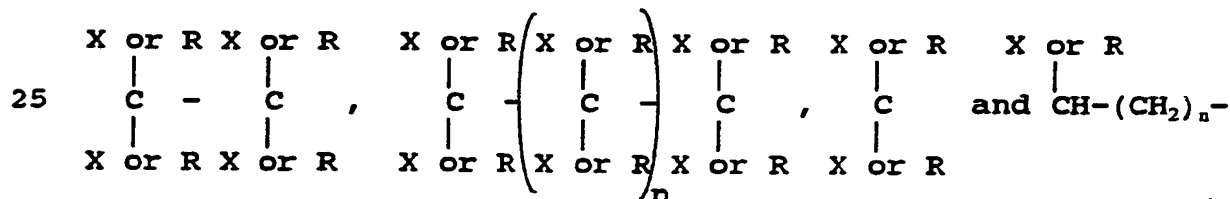


10 wherein each X is selected (independently) from the group  
consisting of H, CH<sub>3</sub>, CH<sub>3</sub>O, CH<sub>3</sub>CH<sub>2</sub>O, Br, Cl, F, CF<sub>3</sub>, CHF<sub>2</sub>,  
CH<sub>2</sub>F, CF<sub>3</sub>O, CH<sub>3</sub>S, OH, CH<sub>2</sub>OH, CONH<sub>2</sub>, CN-, NO<sub>2</sub>, and CH<sub>3</sub>CH<sub>2</sub>; Ar  
is a hydrophobic entity;

each R independently is selected from the group con-  
sisting of hydrogen, methyl, ethyl, propyl, isopropyl,  
15 butyl, isobutyl, cyclopentyl, cyclohexyl, cycloheptyl,  
cyclooctyl, indenyl, indanyl, dihydroindolyl, thiodi-  
hydroindolyl, and 2-, 3-, or 4-piperid(in)yl;

Y is selected from the group consisting of CH, nitro-  
20 gen and an unsaturated carbon;

Z is selected from the group consisting of oxygen,  
nitrogen, sulfur,



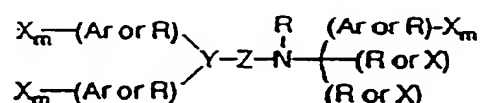
where each n is independently between 1 and 4 inclusive,  
30 and each m is independently between 0 and 5 inclusive;

wherein said molecule is either a calcimimetic or  
calcilytic.



26. The pharmaceutical composition of claim 25 wherein said hydrophobic entity is selected from the group consisting of phenyl, 2-, 3-, or 4-pyridyl, 1- or 2-naphthyl, 1- or 2-quinoliny, 2- or 3-indolyl, benzyl, and  
5 phenoxy.

27. A pharmaceutical composition comprising a molecule having the formula

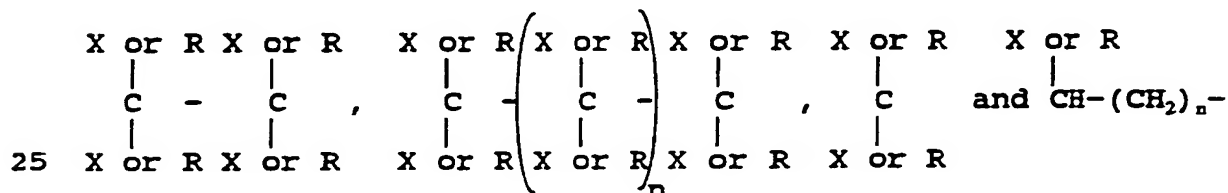


wherein each X independently is selected from the group consisting of H, CH<sub>3</sub>, CH<sub>3</sub>O, CH<sub>3</sub>CH<sub>2</sub>O, Br, Cl, F, CF<sub>3</sub>, CHF<sub>2</sub>,  
10 CH<sub>2</sub>F, CF<sub>3</sub>O, CH<sub>3</sub>S, OH, CH<sub>2</sub>OH, CONH<sub>2</sub>, CN, NO<sub>2</sub>, and CH<sub>3</sub>CH<sub>2</sub>; Ar is a hydrophobic entity;

each R independently is selected from the group consisting of hydrogen, methyl, ethyl, propyl, isopropyl, butyl, isobutyl, cyclopentyl, cyclohexyl, cycloheptyl,  
15 cyclooctyl, indenyl, indanyl, dihydroindolyl, thiodihydroindolyl, and 2-, 3-, or 4-piperid(in)yl;

Y is selected from the group consisting of CH, nitrogen and an unsaturated carbon;

Z is selected from the group consisting of oxygen,  
20 nitrogen sulfur,



where each n is independently between 1 and 4 inclusive, and each m is independently between 0 and 5 inclusive;

wherein said molecule is not NPS 447 or NPS 449.

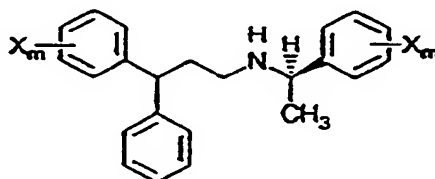
30 28. The pharmaceutical composition of claim 27 wherein said molecule is NPS 467.

29. The pharmaceutical composition of claim 25 wherein said molecule is NPS 459, NPS 467, NPS 551 or NPS 568.

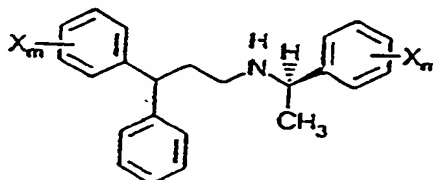
30. The pharmaceutical composition of claim 25, wherein said molecule is an R-diphenylpropyl- $\alpha$ -phenethylamine derivative.

31. The pharmaceutical composition of claim 27, wherein said molecule is an R-diphenylpropyl- $\alpha$ -phenethylamine derivative.

32. The pharmaceutical composition of claim 30 wherein said molecule has the formula:



33. The pharmaceutical composition of claim 31 wherein said molecule has the formula:



34. The pharmaceutical composition of claim 32 wherein each X is independently selected from the group consisting of Cl, F, CF<sub>3</sub>, CH<sub>3</sub>, and CH<sub>3</sub>O.

35. The pharmaceutical composition of claim 33 wherein each X is independently selected from the group consisting of Cl, F, CF<sub>3</sub>, CH<sub>3</sub>, and CH<sub>3</sub>O.

36. A method for treating a patient having a disease characterized by an abnormal  $[Ca^{2+}]_i$  or  $[Ca^{2+}]_o$  in one or more cells or in the blood or plasma, comprising the step of administering to said patient a therapeutically effective amount of a molecule which mimics the activity of extracellular  $Ca^{2+}$  by evoking an increase in  $[Ca^{2+}]_i$  in a cell, or blocks an increase in  $[Ca^{2+}]_i$  evoked by extracellular  $Ca^{2+}$ .

37. The method of claim 36 wherein said molecule has an  $EC_{50}$  of less than or equal to  $5 \mu M$ .

38. The method of claim 36 wherein said molecule is not protamine.

39. The method of claim 36 wherein said molecule interacts at a  $Ca^{2+}$  receptor as a calcimimetic or calcilytic.

40. The method of claim 36 wherein said increase in  $[Ca^{2+}]_i$  in a cell is transient, having a duration of less than thirty seconds.

41. The method of claim 36 or 40 wherein said increase in  $[Ca^{2+}]_i$  is rapid, occurring within thirty seconds.

42. The method of claim 36 wherein said molecule evokes a sustained increase in  $[Ca^{2+}]_i$ , having a duration greater than thirty seconds.

43. The method of claim 36 wherein said molecule further evokes an increase in inositol-1,4,5-trisphosphate or diacylglycerol.

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44. The method of claim 43 wherein said increase in said inositol-1,4,5-trisphosphate or diacylglycerol occurs within less than 60 seconds.

45. The method of claim 40 wherein said transient increase is diminished by pretreatment of said cell with an activator of protein kinase C.

46. The method of claim 45 wherein said activator of protein kinase C is selected from the group consisting of phorbol myristate acetate, mezerein and (-)-indolactam V.

47. The method of claim 36 wherein said molecule inhibits dopamine- or isoproterenol-stimulated cyclic AMP formation.

48. The method of claim 40 wherein said transient increase in  $[Ca^{2+}]_i$  is reduced by pretreatment of said cell for ten minutes with 10 mM sodium fluoride.

49. The method of claim 36 wherein said cell is a parathyroid cell and said molecule inhibits parathyroid hormone secretion from said cell.

50. The method of claim 36 wherein said molecule elicits an increase in  $Cl^-$  conductance in a Xenopus oocyte injected with mRNA from a cell selected from the group consisting of parathyroid cells, bone osteoclasts, juxtaglomerular kidney cells, proximal tubule kidney cells, keratinocytes, parafollicular thyroid cells and placental trophoblasts.

51. The method of claim 36 wherein said molecule mobilizes  $Ca^{2+}$  within the cell to cause said increase in  $[Ca^{2+}]_i$ .

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52. The method of claim 36 wherein said cell is a C-cell or an osteoclast and said molecule inhibits bone resorption in vivo.

53. The method of claim 36 wherein said cell is a  
5 C-cell and said molecule stimulates calcitonin secretion in vitro or in vivo.

54. The method of claim 36 wherein said molecule is either a calcimimetic or calcilytic having an  $EC_{50}$  or  $IC_{50}$  at a  $Ca^{2+}$  receptor of less than or equal to 5  $\mu M$ .

10 55. The method of claim 36 wherein said molecule has an  $EC_{50}$  less than or equal to 5  $\mu M$  at one or more but not all cells chosen from the group consisting of parathyroid cells, bone osteoclasts, juxtaglomerular kidney cells, proximal tubule kidney cells, keratinocytes, parafollicu-  
15 lar thyroid cells and placental trophoblasts.

56. The method of claim 36, 54 or 55 wherein said molecule has an  $EC_{50}$  or  $IC_{50}$  less than or equal to 1  $\mu M$ .

57. The method of claim 36, 54 or 55 wherein said molecule has an  $EC_{50}$  or  $IC_{50}$  less than or equal to 100  
20 nanomolar.

58. The method of claim 36, 54, or 55 wherein said molecule has an  $EC_{50}$  or  $IC_{50}$  less than or equal to 10 nanomolar.

59. The method of claim 38, 54 or 55, wherein said  
25 molecule has an  $EC_{50}$  or  $IC_{50}$  less than or equal to 1 nanomolar.

60. The method of claim 36, 54 or 55 wherein said molecule is positively charged at physiological pH.

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61. The method of claim 60 wherein said molecule is selected from the group consisting of branched or cyclic polyalkylamines, positively charged polyamino acids, and arylamines.

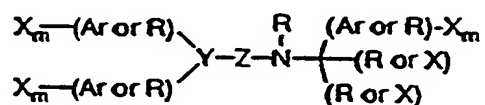
- 5 62. The method of claim 61 wherein said branched polyamine has the formula



wherein k is an integer from 1 to 10,  
each j is the same or different and is an integer  
10 from 2 to 20, and

each  $R_i$  is the same or different and is selected from the group consisting of hydrogen and  $-(\text{CH}_2)_j-\text{NH}_2$ , wherein j is as defined above, and wherein at least one  $R_i$  is not hydrogen.

- 15 63. The method of claim 36, 54 or 55 wherein said molecule has the formula



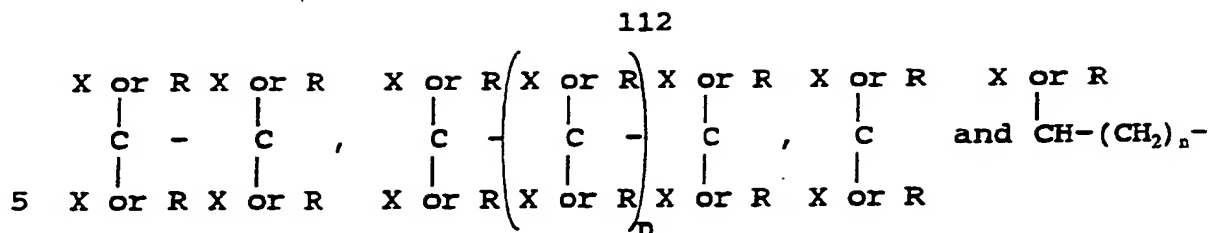
wherein each X independently is selected from the group consisting of H,  $\text{CH}_3$ ,  $\text{CH}_3\text{O}$ ,  $\text{CH}_3\text{CH}_2\text{O}$ , Br, Cl, F,  $\text{CF}_3$ ,  $\text{CHF}_2$ ,  $\text{CH}_2\text{F}$ ,  $\text{CF}_3\text{O}$ ,  $\text{CH}_3\text{S}$ , OH,  $\text{CH}_2\text{OH}$ ,  $\text{CONH}_2$ , CN,  $\text{NO}_2$ , and  $\text{CH}_3\text{CH}_2$ ;

20 Ar is a hydrophobic entity;

each R independently is selected from the group consisting of hydrogen, methyl, ethyl, propyl, isopropyl, butyl, isobutyl, cyclopentyl, cyclohexyl, cycloheptyl, cyclooctyl, indenyl, indanyl, dihydroindolyl, thiodi-  
25 hydroindolyl, and 2-, 3-, or 4-piperid(in)yl;

Y is selected from the group consisting of CH, nitrogen and an unsaturated carbon;

Z is selected from the group consisting of oxygen, nitrogen, sulfur,

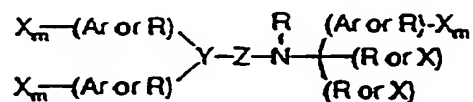


where each n is independently between 1 and 4 inclusive,  
and each m is independently between 0 and 5 inclusive;

wherein said molecule is either a calcimimetic or  
calcilytic.

64. The method of claim 63 wherein said hydrophobic entity is selected from the group consisting of phenyl, 2-3-, or 4-pyridyl, 1- or 2-naphthyl, 1- or 2-quinoliny, 2- or 3-indolyl, benzyl, and phenoxy.

65. The method of claim 36, 54 or 55 wherein said molecule has the formula



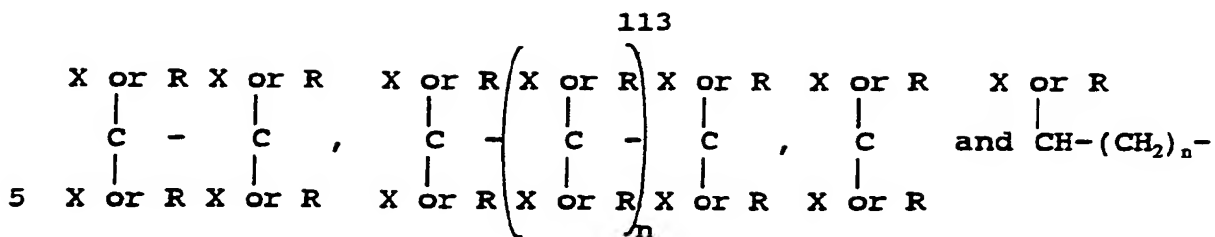
wherein each X independently is selected from the group consisting of H, CH<sub>3</sub>, CH<sub>3</sub>O, CH<sub>3</sub>CH<sub>2</sub>O, Br, Cl, F, CF<sub>3</sub>, CHF<sub>2</sub>, CH<sub>2</sub>F, CF<sub>3</sub>O, CH<sub>3</sub>S, OH, CH<sub>2</sub>OH, CONH<sub>2</sub>, CN, NO<sub>2</sub>, and CH<sub>3</sub>CH<sub>2</sub>;

Ar is a hydrophobic entity;

each R independently is selected from the group consisting of hydrogen, methyl, ethyl, propyl, isopropyl, butyl, isobutyl, cyclopentyl, cyclohexyl, cycloheptyl, cyclooctyl, indenyl, indanyl, dihydroindolyl, thiodihydroindolyl, and 2-,3-, or 4-piperid(in)yl;

Y is selected from the group consisting of CH, nitrogen and an unsaturated carbon;

Z is selected from the group consisting of oxygen, nitrogen, sulfur,



where each n is independently between 1 and 4 inclusive,  
 and each m is independently between 0 and 5 inclusive;  
 wherein said molecule is not NPS 447 or NPS 449.

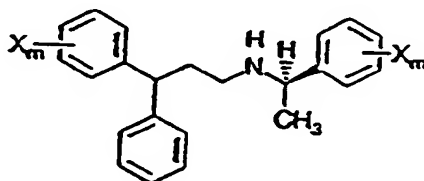
66. The method of claim 65 wherein said molecule is  
 NPS 467 or NPS 019.

67. The method of claim 63 wherein said molecule is  
 NPS 467, NPS 019 or NPS 456.

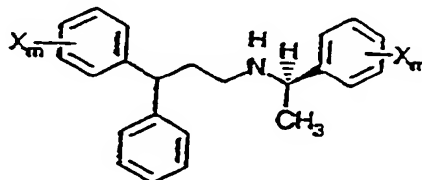
68. The method of claim 63, wherein said molecule is  
 an R-diphenylpropyl- $\alpha$ -phenethylamine derivative.

69. The method of claim 65, wherein said molecule is  
 an R-diphenylpropyl- $\alpha$ -phenethylamine derivative.

70. The method of claim 68 wherein said molecule has  
 the formula:



71. The method of claim 69 wherein said molecule has  
 the formula:





72. The method of claim 70 wherein each X is independently selected from the group consisting of Cl, F, CF<sub>3</sub>, CH<sub>3</sub>, and CH<sub>3</sub>O.

73. The method of claim 71 wherein each X is independently selected from the group consisting of Cl, F, CF<sub>3</sub>, CH<sub>3</sub>, and CH<sub>3</sub>O.

74. The method of claim 36 wherein said patient has a disease characterized by an abnormal level of one or more ions or substances the level of which is regulated or affected by activity of one or more Ca<sup>2+</sup> receptors.

75. The method of claim 74, wherein said molecule is active on a Ca<sup>2+</sup> receptor of a cell selected from the group consisting of parathyroid cells, bone osteoclasts, juxtaglomerular kidney cells, proximal tubule kidney cells, keratinocytes, parafollicular thyroid cells, and placental trophoblasts.

76. The method of claim 36, wherein said patient has a disease selected from the group consisting of primary and secondary hyperparathyroidism, Paget's disease, hypercalcemia malignancy, osteoporosis and hypertension.

77. The method of claim 75 wherein said molecule is active only at one or more said Ca<sup>2+</sup> receptors and not all said Ca<sup>2+</sup> receptors.

78. The method of claim 36 wherein said molecule reduces the level of parathyroid hormone in the serum of said patient.

79. The method of claim 78, wherein said level of parathyroid hormone is reduced to that level present in a normal individual.

80. The method of claim 78 wherein said level is reduced to a degree sufficient to cause a decrease in plasma  $\text{Ca}^{2+}$ .

81. The method of claim 78 wherein said molecule is  
5 provided in an amount sufficient to have a therapeutically relevant effect on said patient.

82. The method of claim 36 wherein said molecule blocks an increase in  $[\text{Ca}^{2+}]_i$  within the cell evoked by extracellular  $[\text{Ca}^{2+}]$ .

10 83. A method for diagnosis of a disease or condition in a patient, comprising the steps of identifying the number and/or location of one or more  $\text{Ca}^{2+}$  receptors within said patient and comparing said number and/or location with that observed in normal patients as an indication of  
15 the presence of a said disease or condition.

84. The method of claim 83 wherein said method is an immunoassay in which an antibody to a  $\text{Ca}^{2+}$  receptor is used to identify the number or location of said  $\text{Ca}^{2+}$  receptors.

85. The method of claim 83 wherein said assay com-  
20 prises providing a labelled calcimimetic or calcilytic molecule which binds to a  $\text{Ca}^{2+}$  receptor.

86. The method of claim 83 wherein said disease is a cancer.

87. The method of claim 86 wherein said cancer is an  
25 ectopic tumor in the parathyroid.

88. The method of claim 83 wherein said condition is characterized by an increased number or activity of osteoclasts in bone.

89. A method for treatment of osteoporosis, hyperparathyroidism or hypertension, comprising the steps of administering a therapeutically effective amount of a molecule which mimics the activity of extracellular  $\text{Ca}^{2+}$  by evoking an increase in  $[\text{Ca}^{2+}]_i$  in a cell, or blocks an increase in  $[\text{Ca}^{2+}]_i$  (elicited by extracellular  $\text{Ca}^{2+}$ ).

90. A method for identifying a molecule useful as a therapeutic molecule, comprising the steps of screening a potentially useful molecule for its ability to mimic the activity of extracellular  $\text{Ca}^{2+}$  in a cell, or to block an increase in  $[\text{Ca}^{2+}]_i$  (elicited by extracellular  $\text{Ca}^{2+}$ ), and determining whether said molecule has an  $\text{EC}_{50}$  of less than or equal to 5  $\mu\text{M}$ .

91. A recombinant  $\text{Ca}^{2+}$  receptor.

92. A cell comprising a recombinant  $\text{Ca}^{2+}$  receptor.

93. A method for identifying a useful calcimimetic molecule comprising the step of identifying a molecule which mimics one or more activities of  $\text{Ca}^{2+}$  at a first  $\text{Ca}^{2+}$  receptor but not at a second  $\text{Ca}^{2+}$  receptor.

94. A method for identifying a useful calcilytic molecule comprising the step of identifying a molecule which blocks one or more activities of  $\text{Ca}^{2+}$  at a first  $\text{Ca}^{2+}$  receptor but not at a second  $\text{Ca}^{2+}$  receptor.

95. The method of claim 93 or 94 wherein said method includes providing a recombinant  $\text{Ca}^{2+}$  receptor useful in said identifying step.

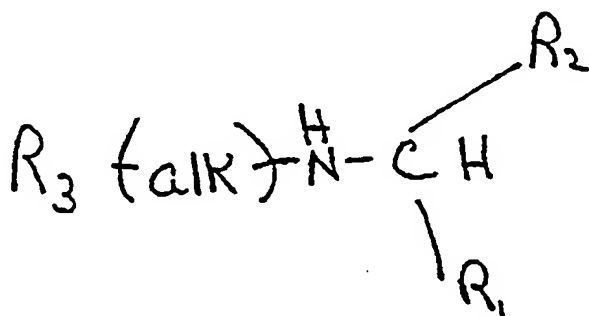
96. Purified nucleic acid encoding a  $\text{Ca}^{2+}$  receptor.

97. The molecules NPS 459, NPS 467, NPS 544, NPS 551 and NPS 568.

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98. The pharmaceutical composition of claim 25 wherein said molecule is NPS 447, NPS 448, NPS 449 or NPS 456.

99. A compound of the formula



5 wherein alk is straight or branched-chain alkylene of from 1 to 6 carbon atoms; R<sub>1</sub> is lower alkyl of from 1 to 3 carbon atoms or lower haloalkyl of from 1 to 3 carbon atoms substituted with from 1 to 7 halogen atoms; and R<sub>2</sub> and R<sub>3</sub> are independently selected monocyclic or bicyclic  
10 carbocyclic aryl or cycloalkyl groups, having 5- or 6-membered rings optionally substituted with 1 to 5 substituents independently selected from lower alkyl of 1 to 3 carbon atoms, lower haloalkyl of 1 to 3 carbon atoms substituted with 1 to 7 halogen atoms, lower alkoxy of 1 to  
15 3 carbon atoms, halogen, nitro, amino, alkylamino, amido, lower alkylamido of 1 to 3 carbon atoms, cyano, hydroxy, acyl of 2 to 4 carbon atoms or lower thioalkyl of 1 to 3 carbon atoms and pharmaceutically acceptable salts and acid addition salts thereof.

20 100. A compound according to claim 99 wherein alk is n-propylene.

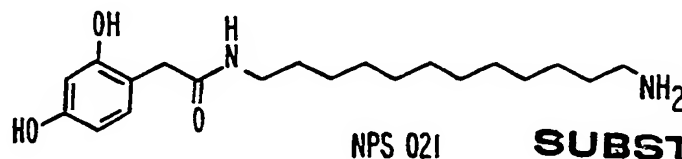
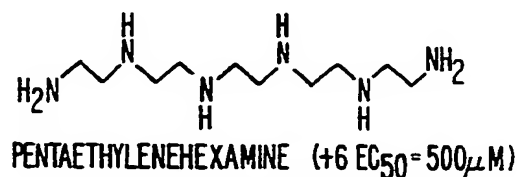
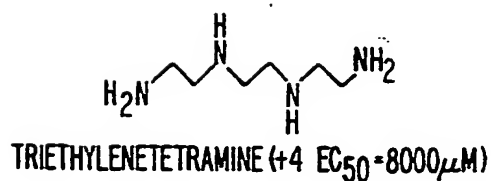
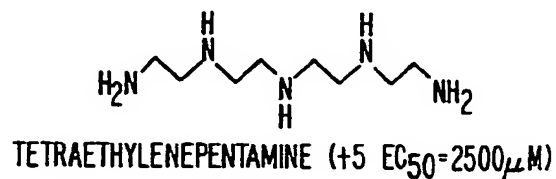
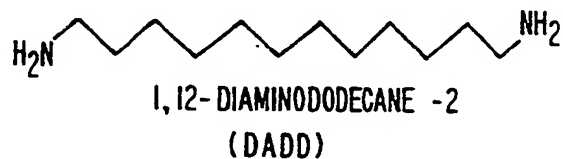
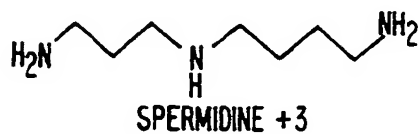
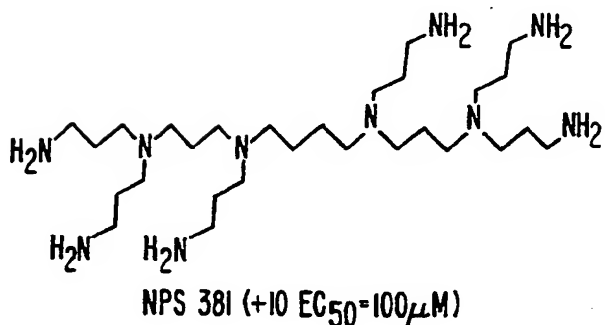
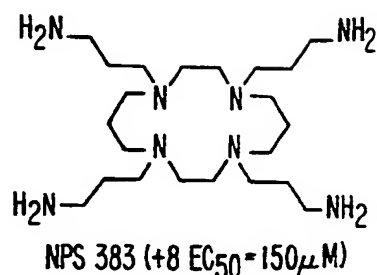
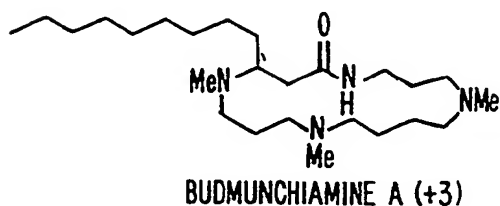
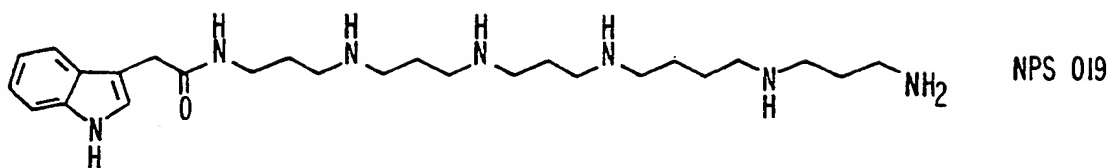
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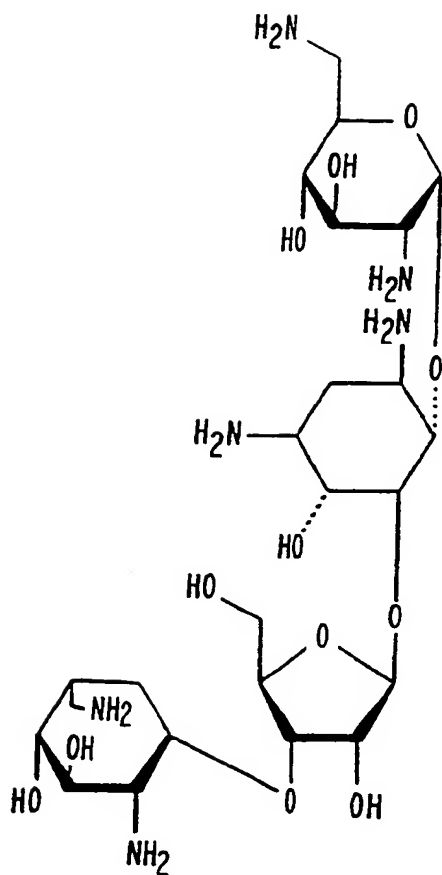
101. A compound according to claim 100 wherein  $R_1$  is methyl.

102. A compound according to claim 101 wherein  $R_2$  and  $R_3$  are independently selected optionally substituted phenyl  
5 groups.

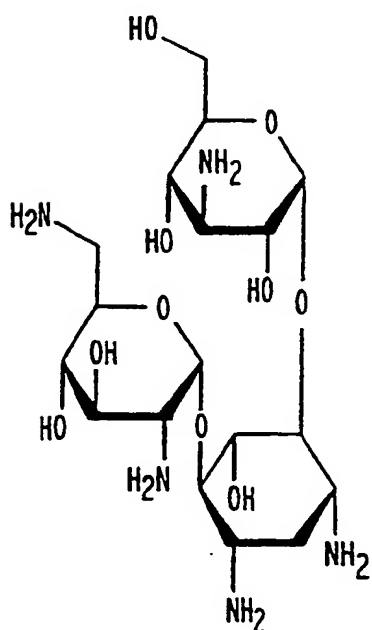
FIG. 1a.



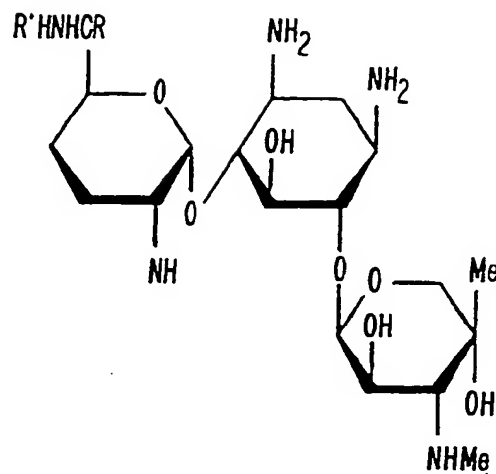
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Neomycin B<sub>2</sub> (+6)



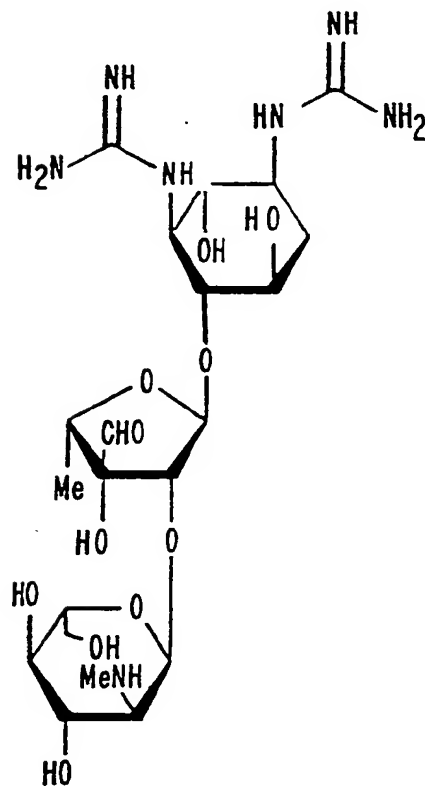
Bekanamycin (+5)



### Gentamicin (Complex +5)

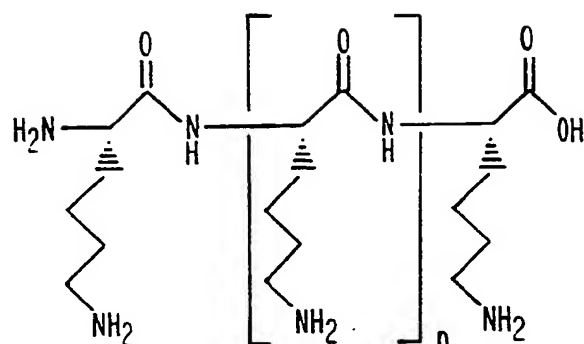
C1  $R = R' = \text{Me}$

C2 R = Me; R' = H

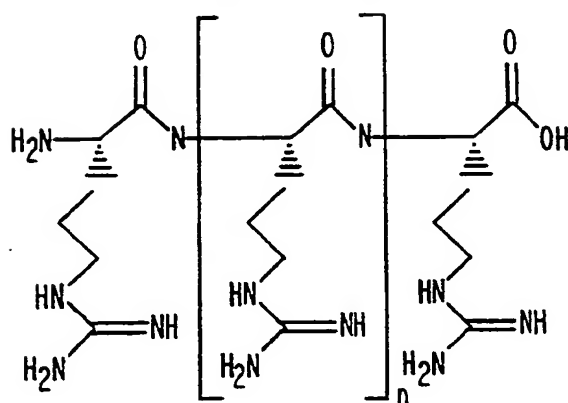
$$C1aR = R' = H$$


Streptomycin (+3)

FIG. 1b.



poly-Lys (27 kD) n= 210 + 210  
 poly-Lys (14 kD) n= 110 + 110  
 poly-Lys (38 kD) n= 55 + 55



poly-Arg (100 kD) n= 640 + 640  
 poly-Arg (40 kD) n= 256 + 256

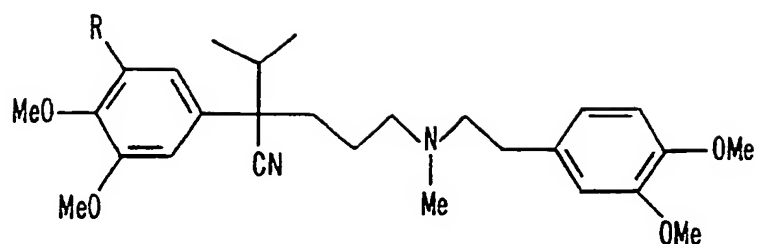
PROTAMINE + 21

H N-Pro-Arg-Arg-Arg-Arg-Ser-Ser-Ser-Arg-Pro-Val-Arg-Arg-Arg-Arg-  
 Pro-Arg-Val-Ser-Arg-Arg-Arg-Arg-Arg-Gly-Gly-Arg-Arg-Arg-OH

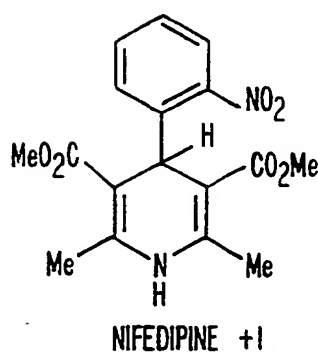
FIG. 1c.

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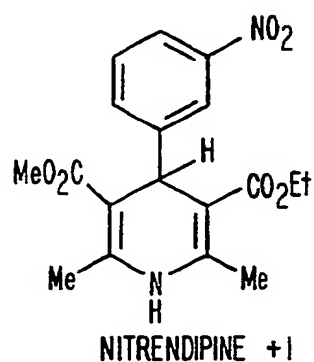




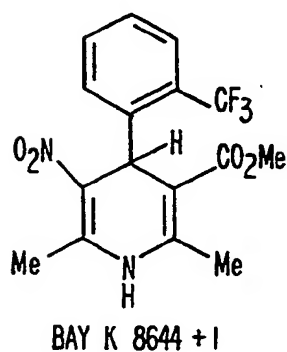
VERAPAMIL R=H +I  
D-600 R=OMe +I



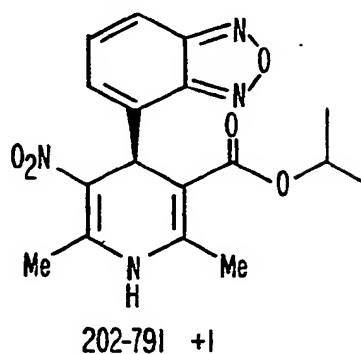
NIFEDIPINE +I



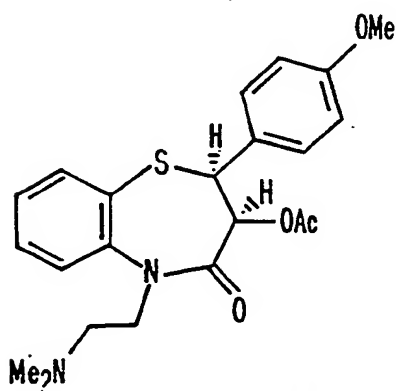
NITRENDIPINE +I



BAY K 8644 +I



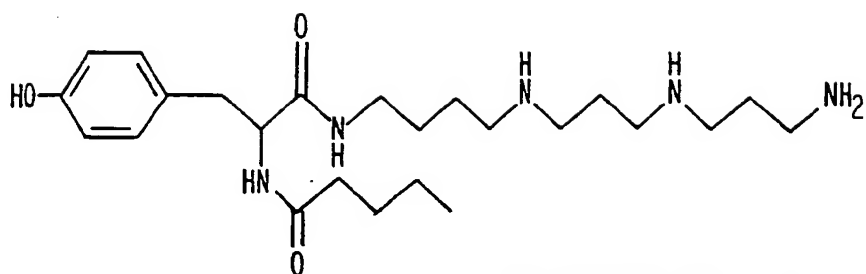
202-791 +I



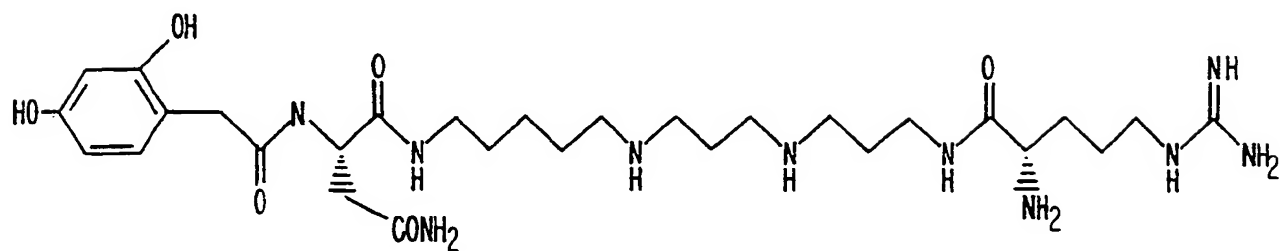
DILTIAZEM +I

FIG. 1d.

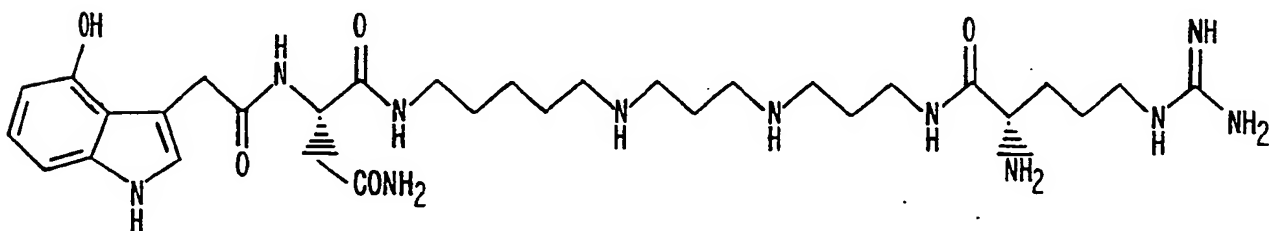
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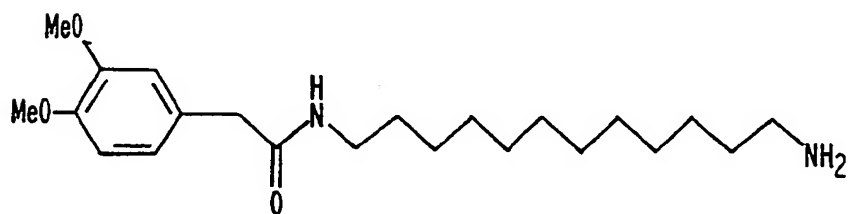
PHILANTHOTOXIN 433 +3



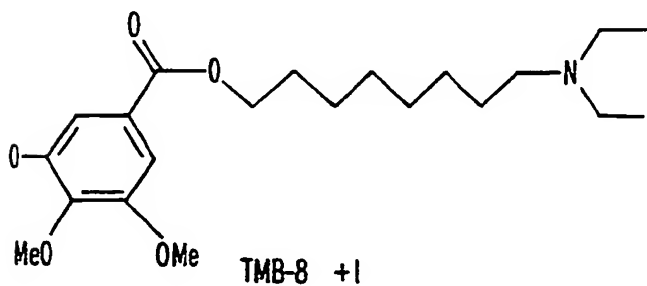
ARGIOTOXIN 636 +4



ARGIOTOXIN 659 +4



NPS 384



TMB-8 +1

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FIG. 1e.

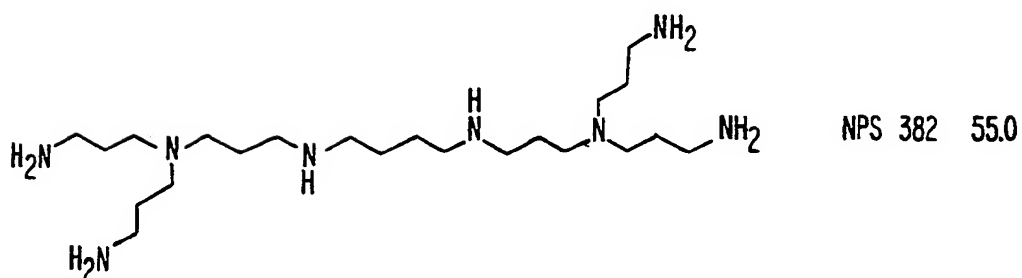
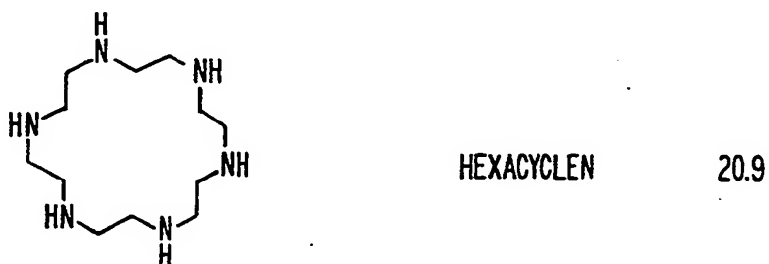
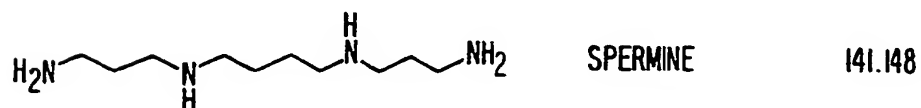
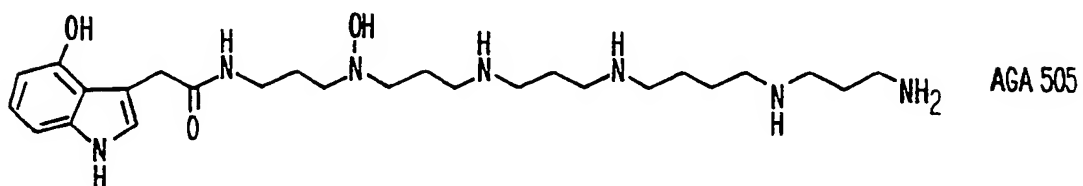
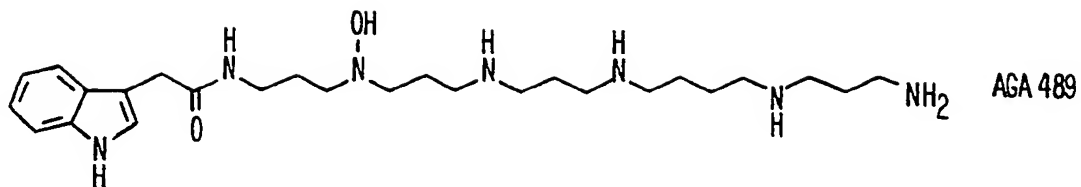
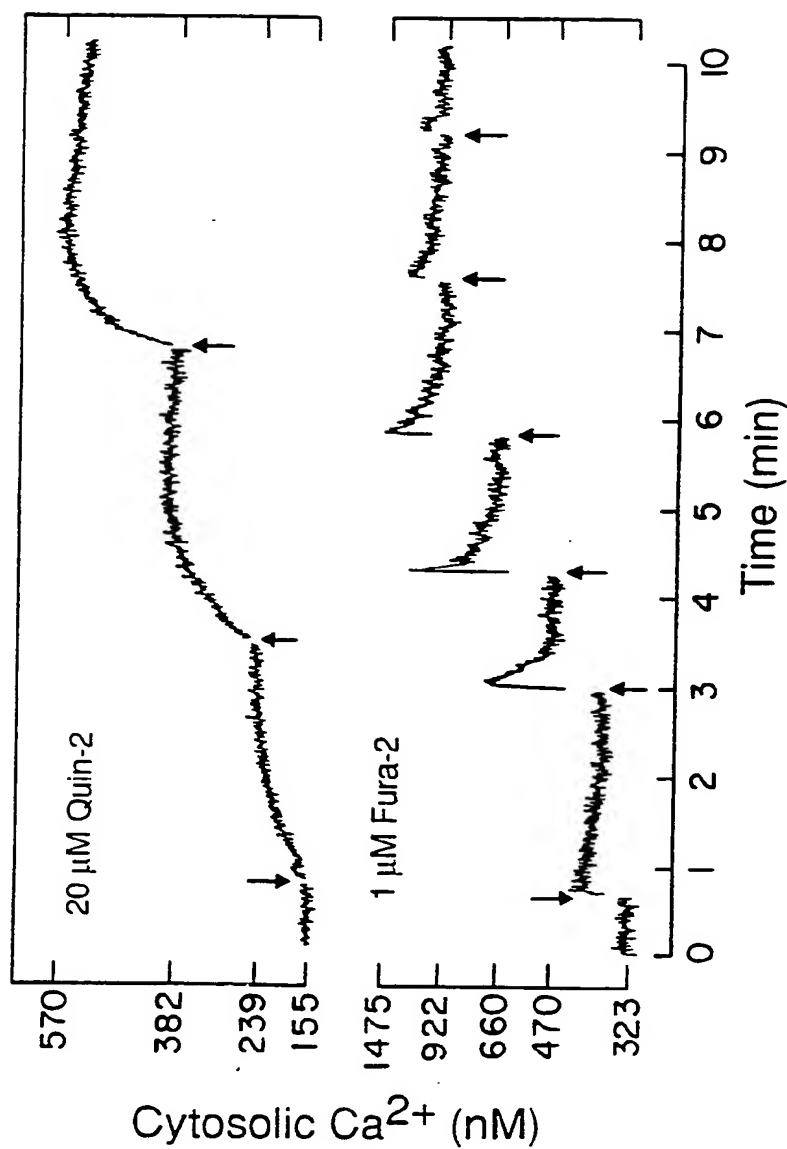


FIG. 1f.

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FIG. 2.



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FIG. 3a.

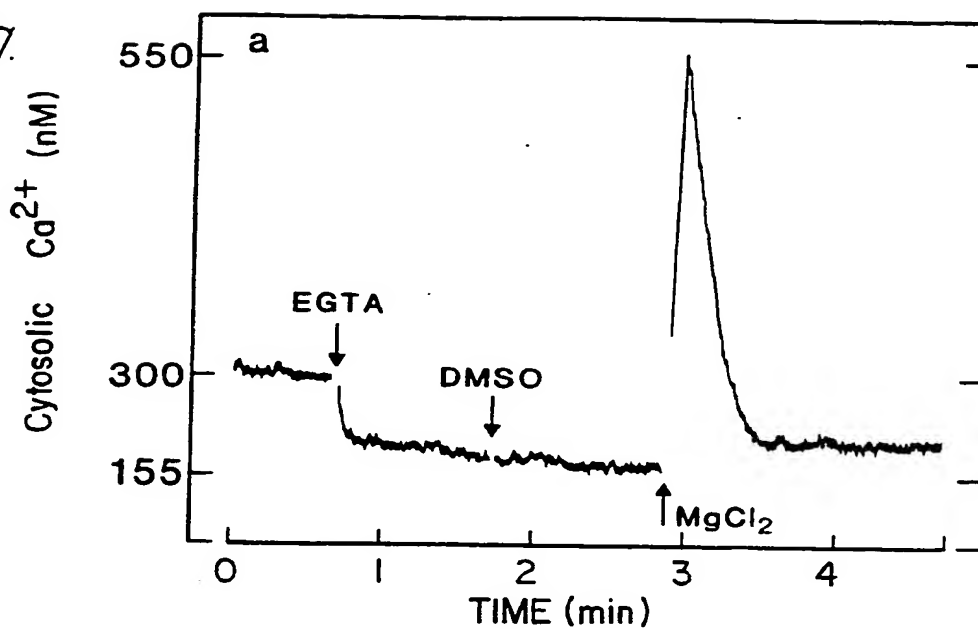


FIG. 3b.

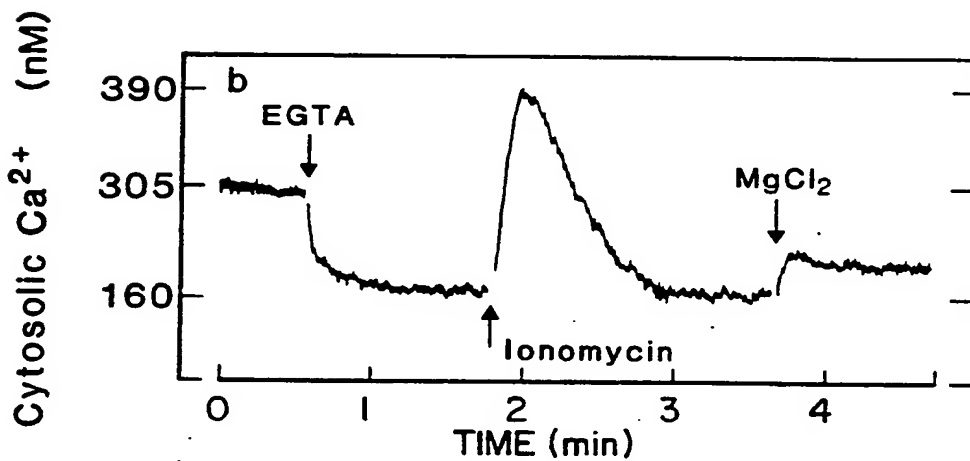


FIG. 3c.

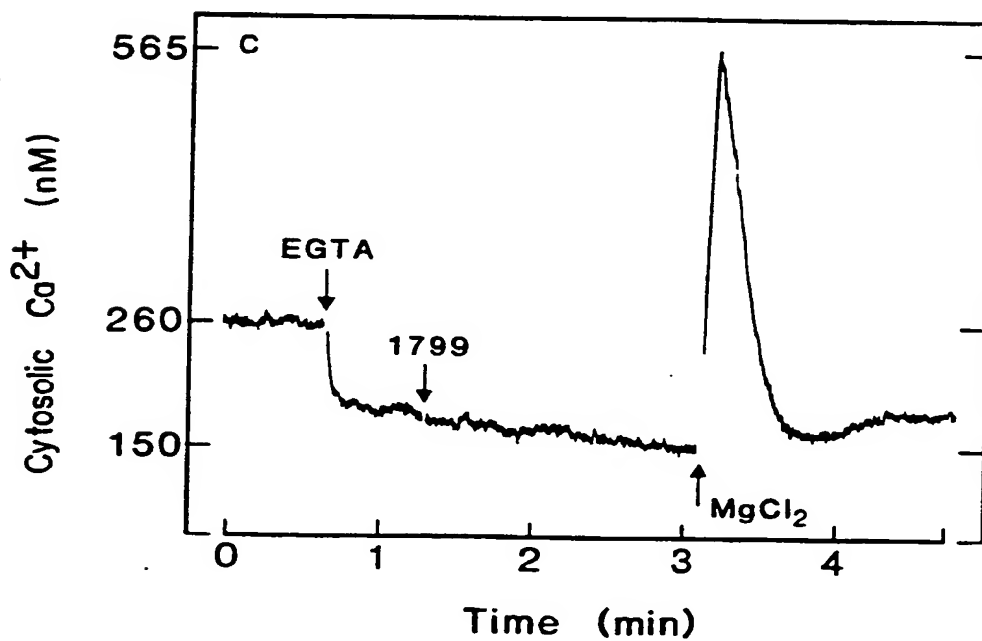


FIG. 4a.

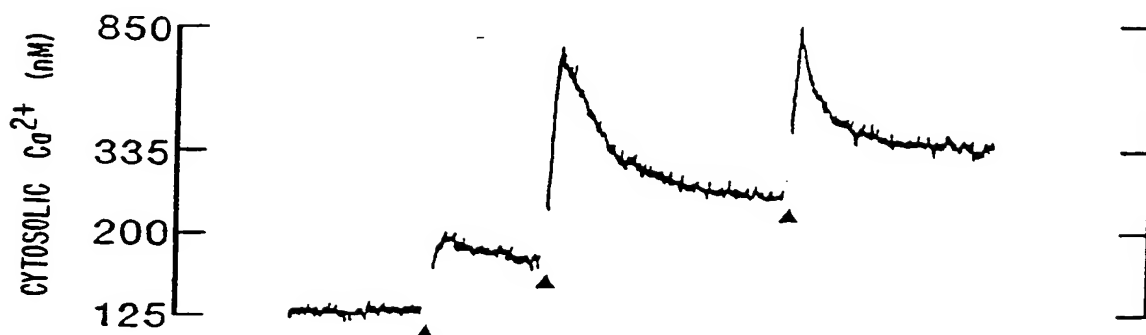


FIG. 4b.

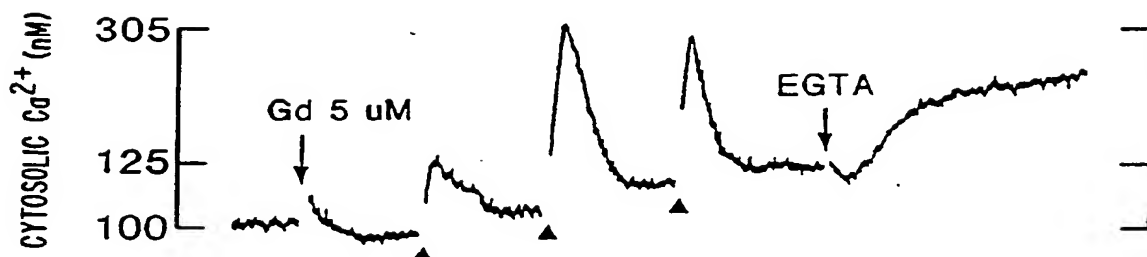
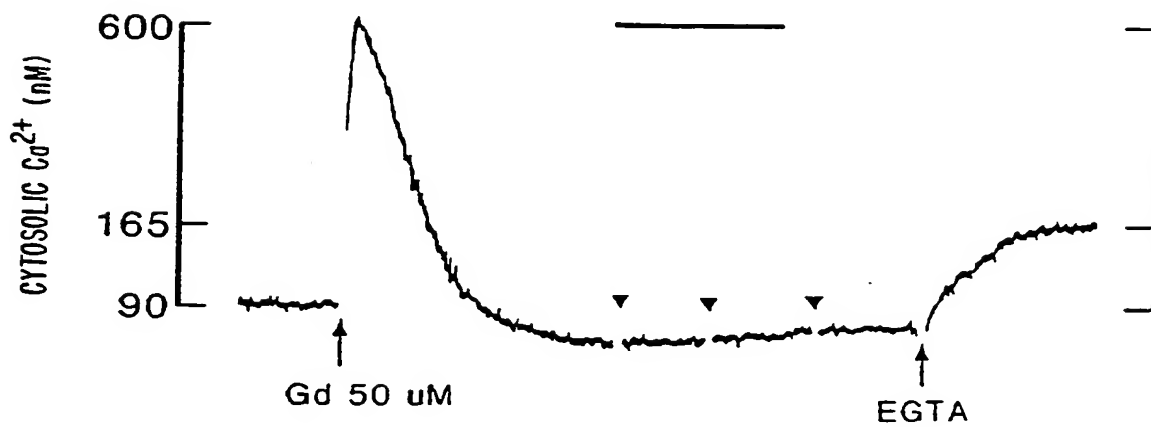


FIG. 4c.



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FIG. 5c.

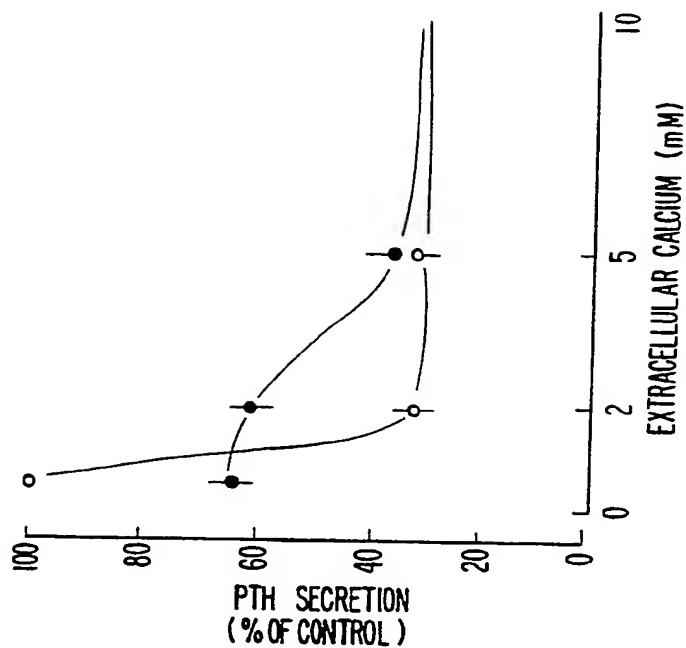


FIG. 5b.

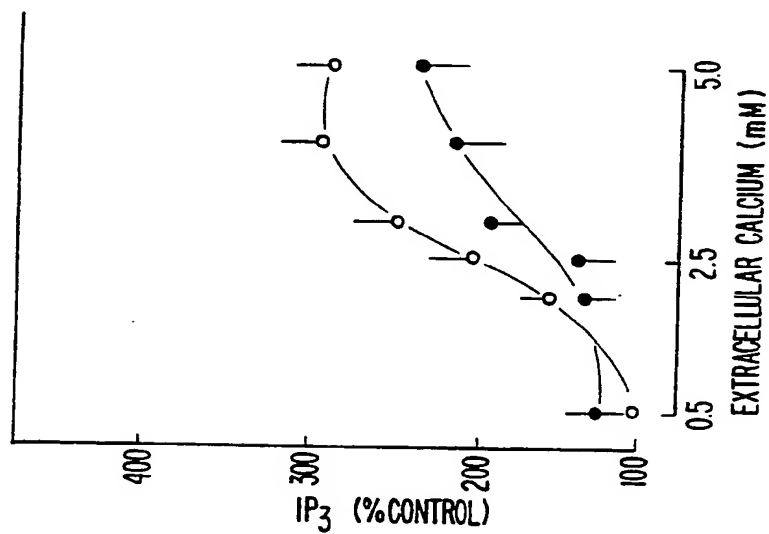
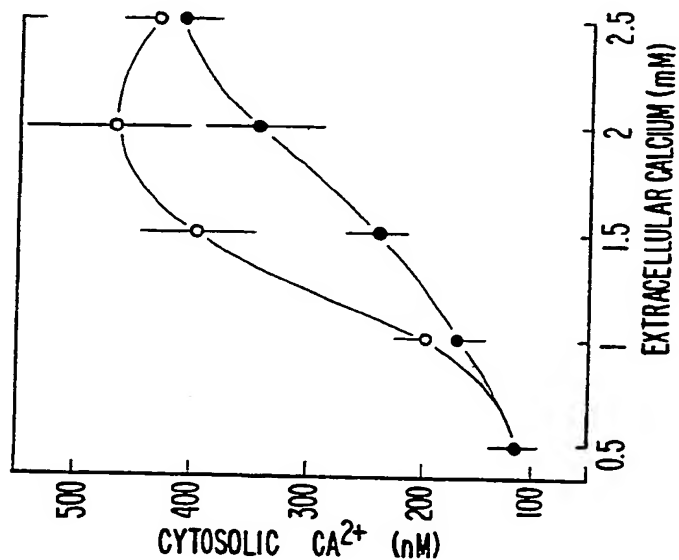


FIG. 5a.



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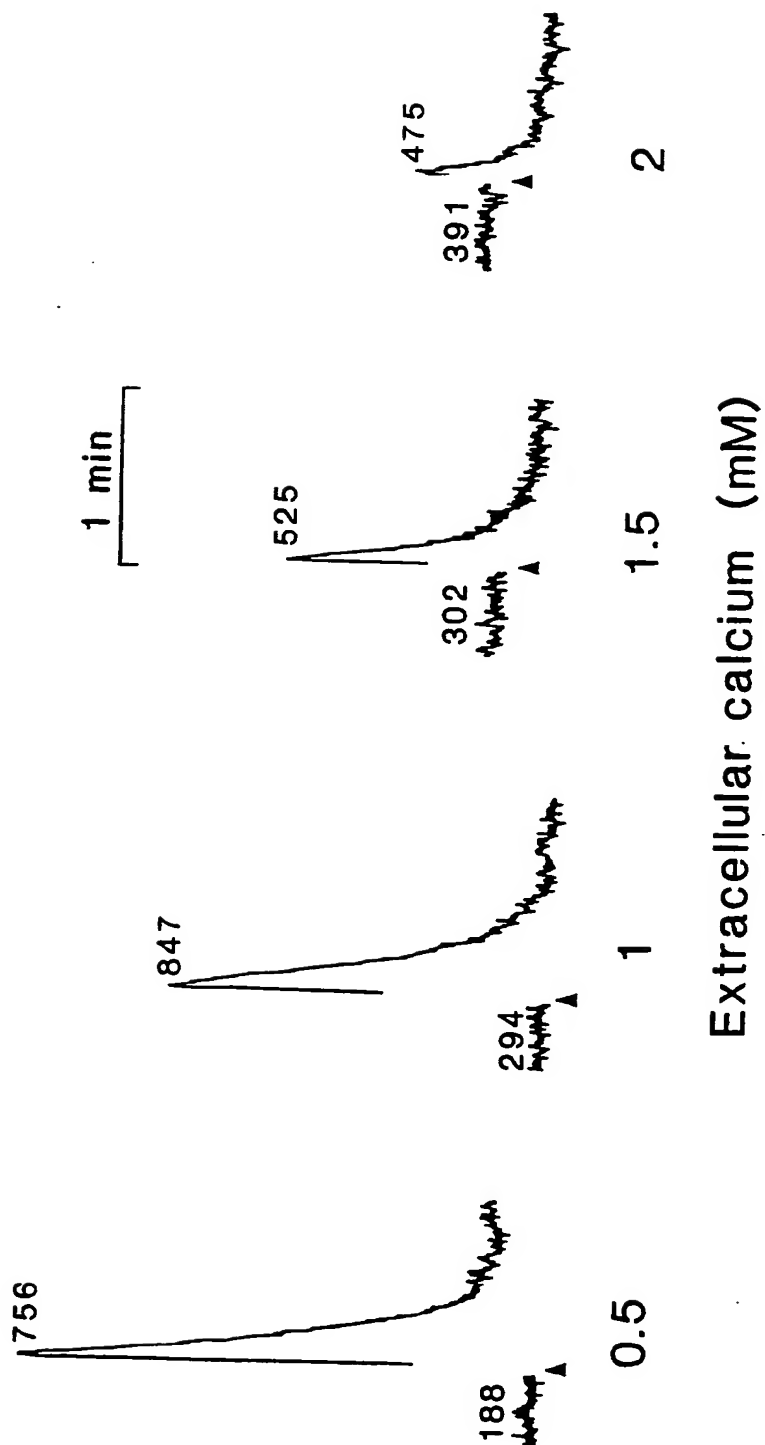


FIG. 6.

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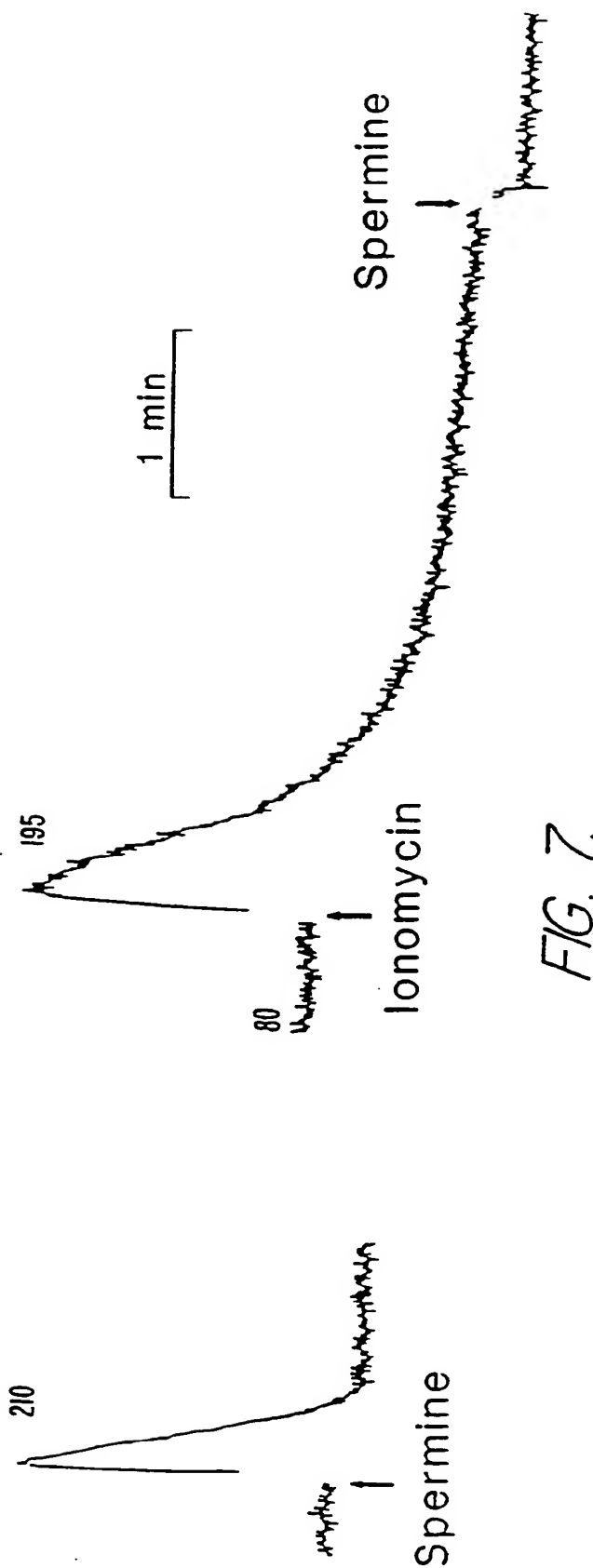


FIG. 7.

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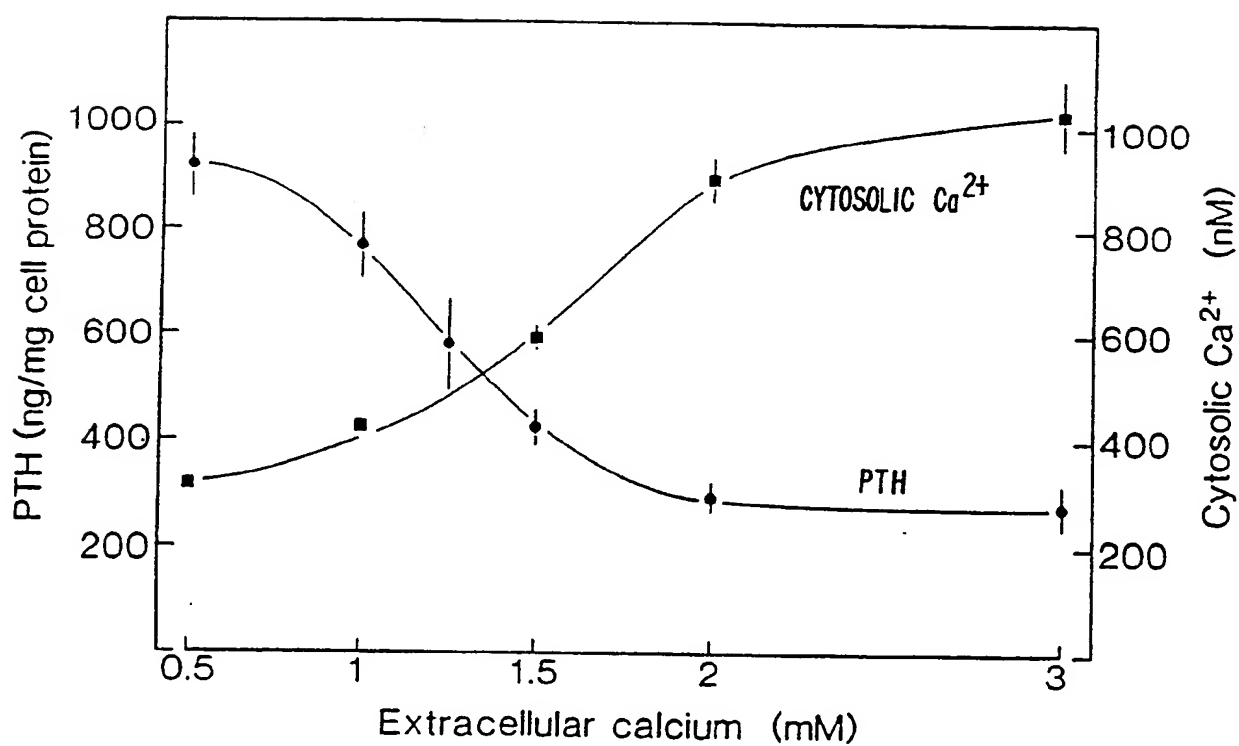
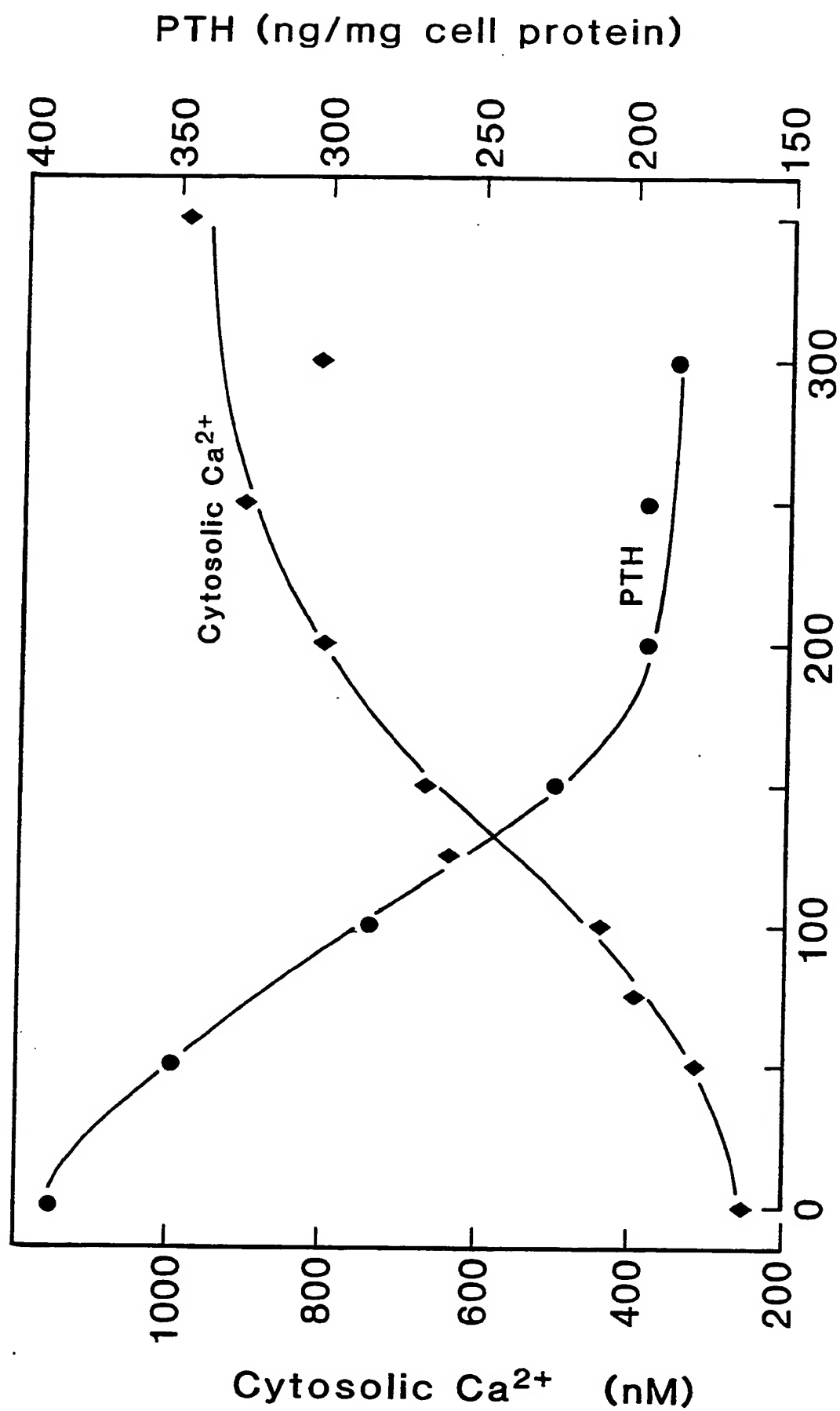


FIG. 8a.



Spermine ( $\mu\text{M}$ )  
FIG. 8b.

FIG. 9a.

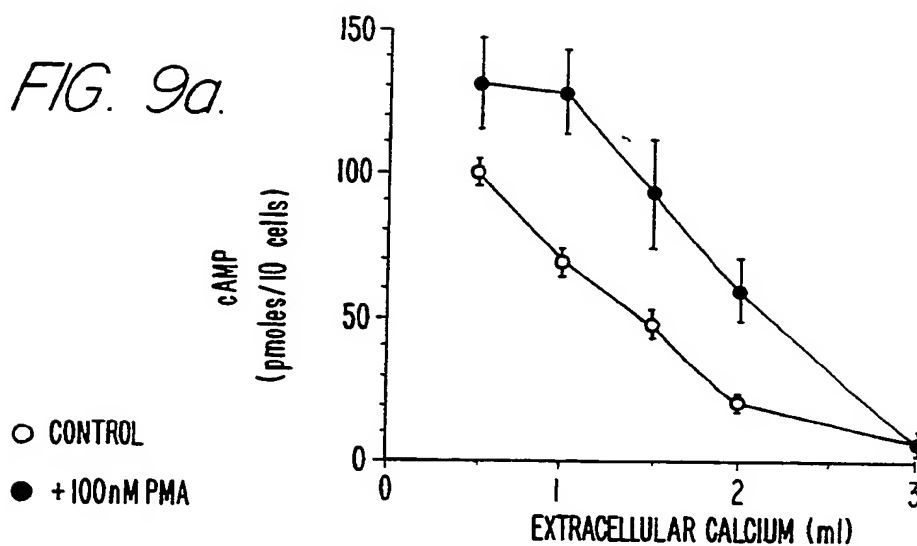


FIG. 9b.

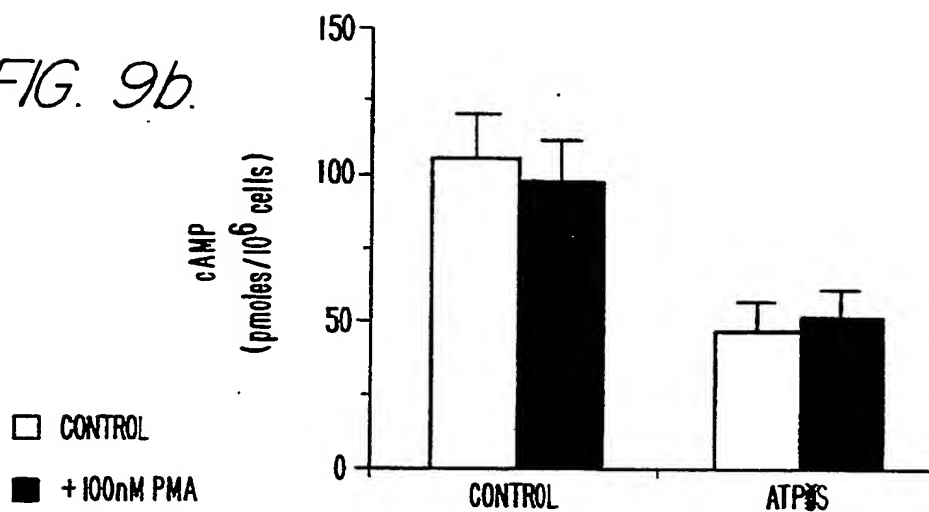
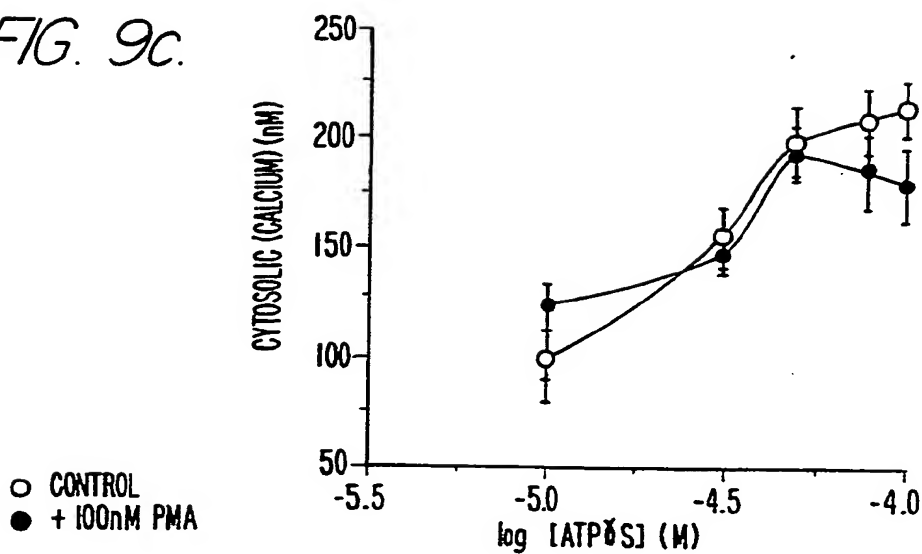
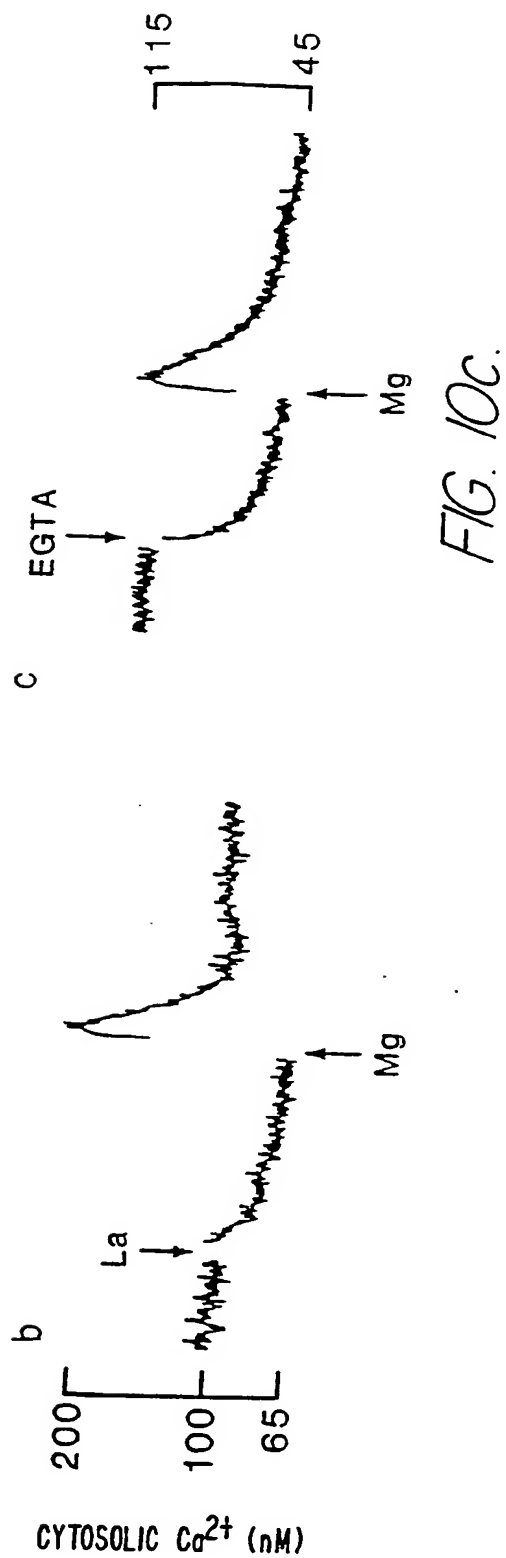
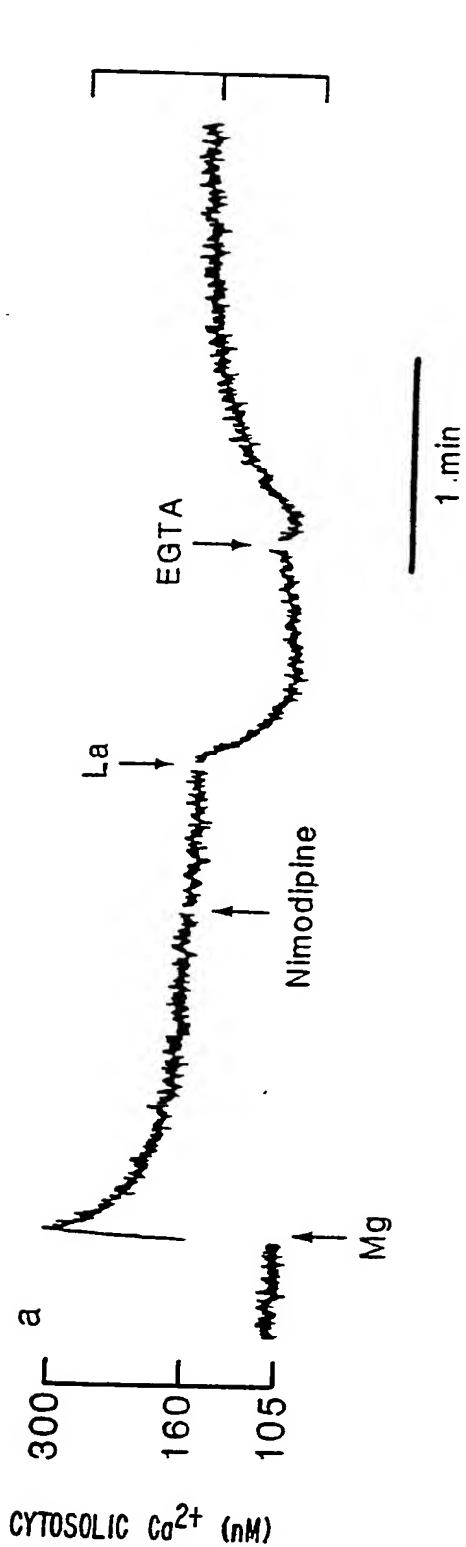


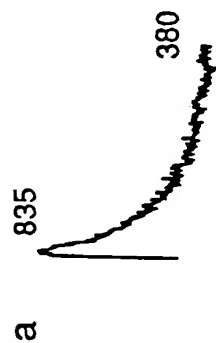
FIG. 9c.





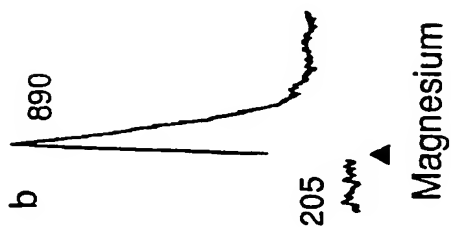
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FIG. 11a.



Calcium

FIG. 11b.



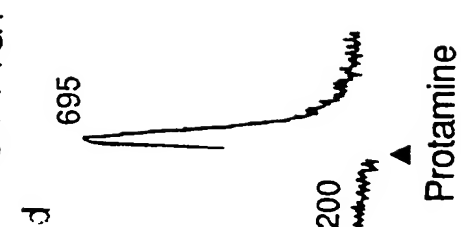
Magnesium

FIG. 11c.



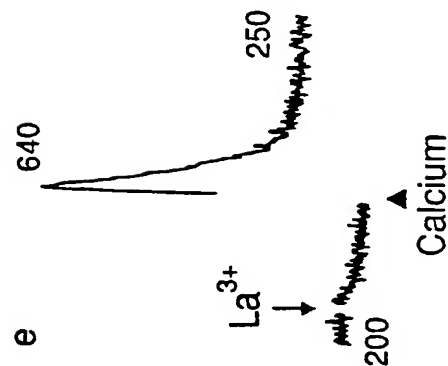
Neomycin

FIG. 11d.



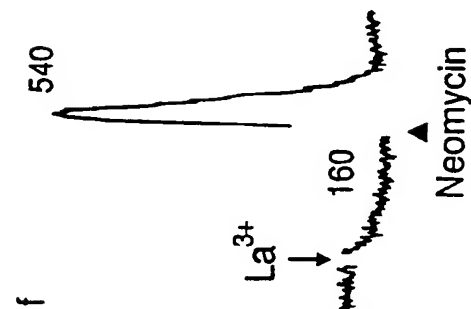
Protamine

FIG. 11e.



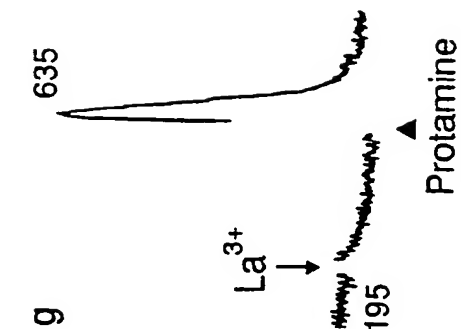
Calcium

FIG. 11f.



Neomycin

FIG. 11g.



Protamine

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FIG. 11h.

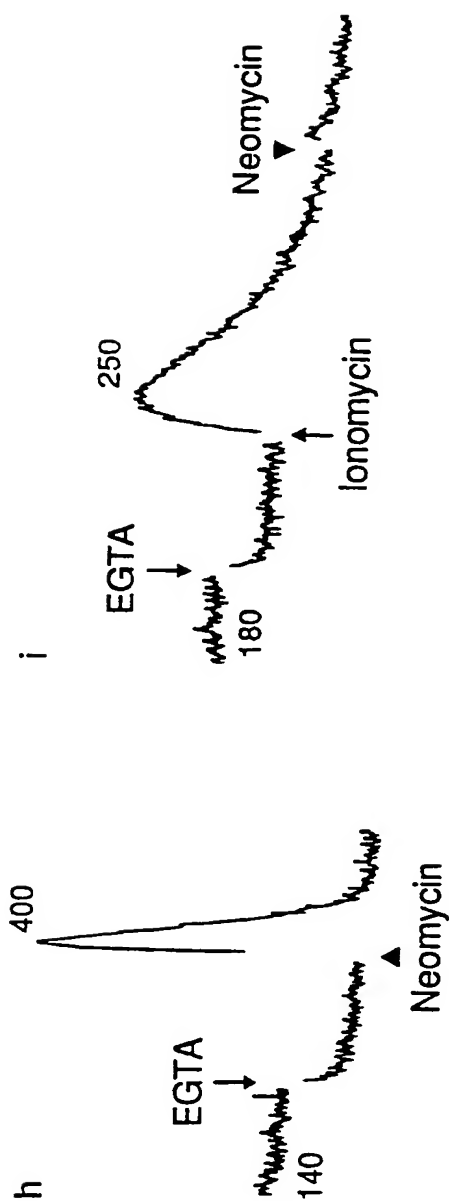
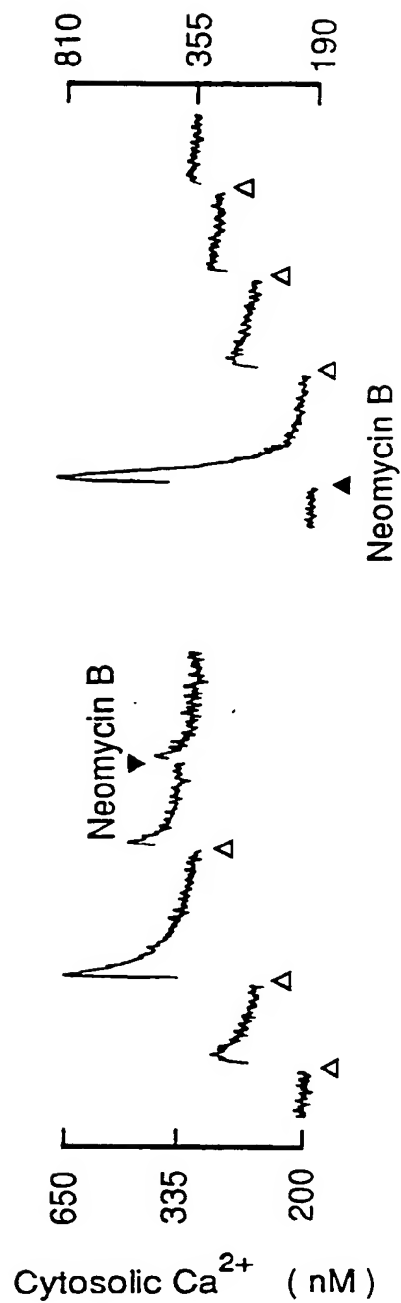


FIG. 11i.

FIG. 12.



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FIG. 13a.

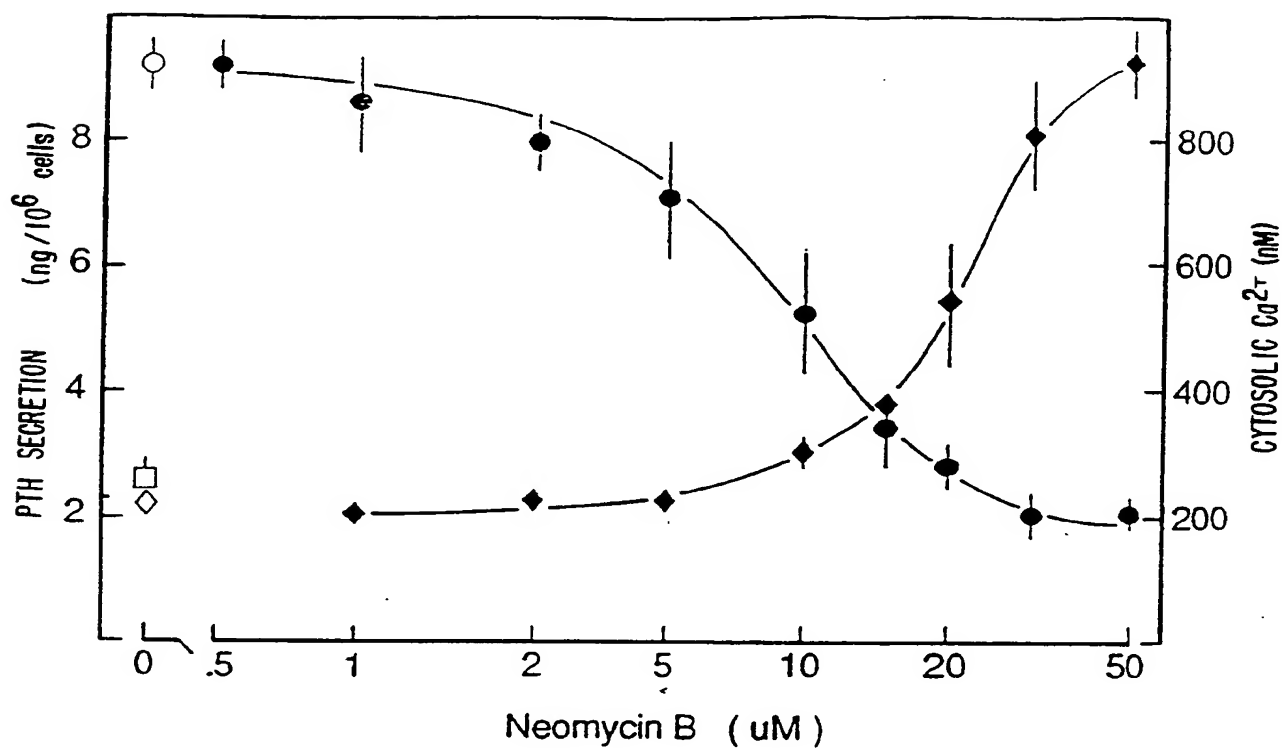
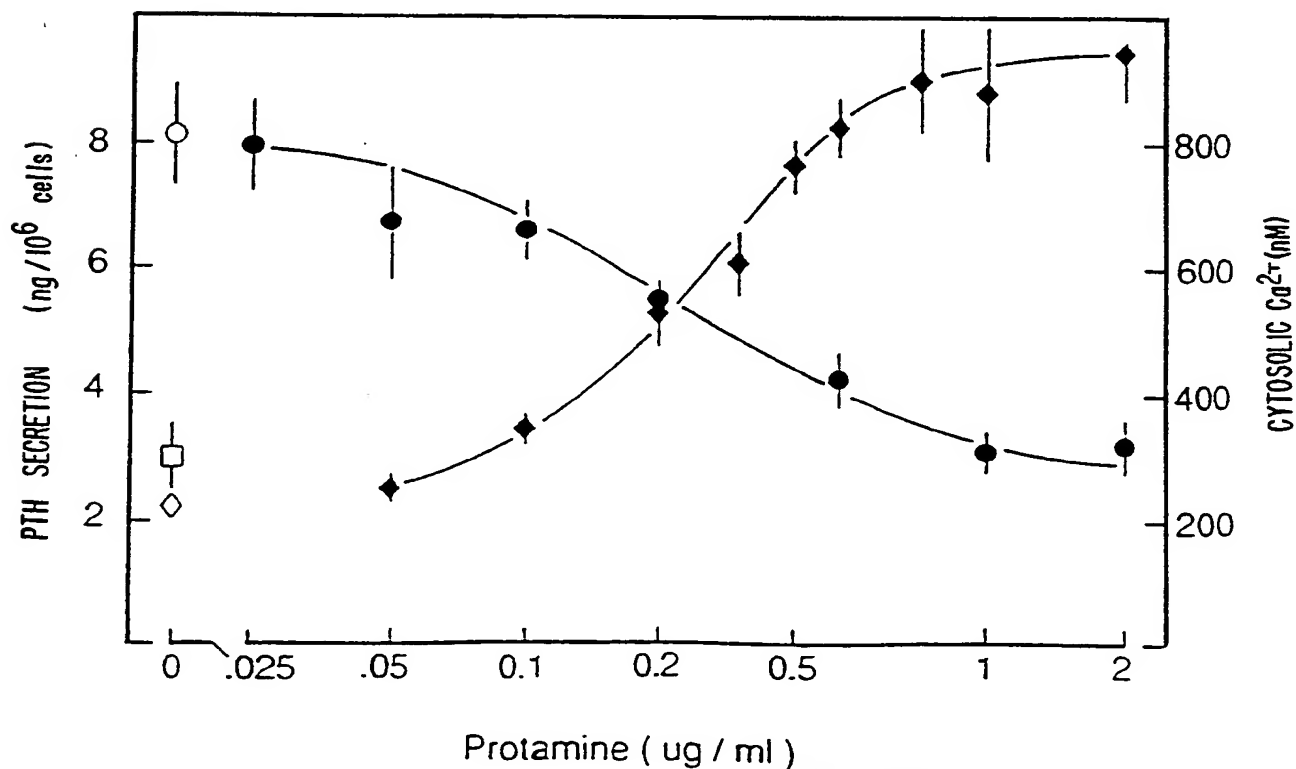


FIG. 13b.

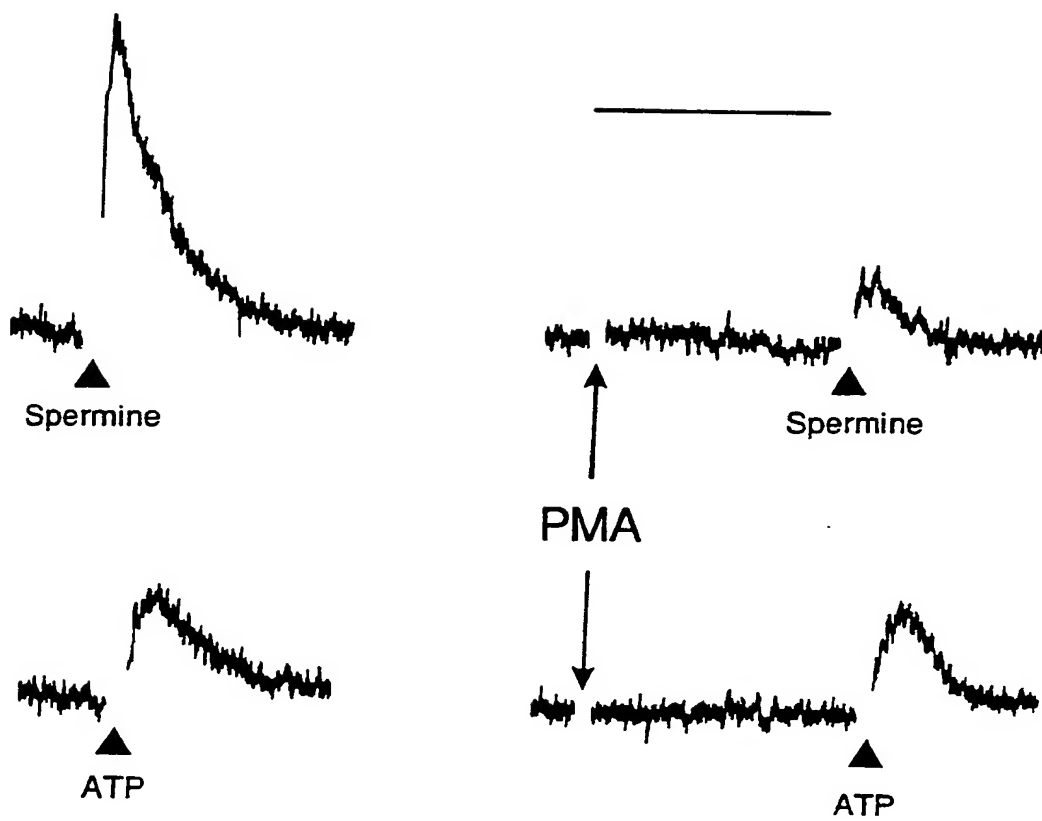


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FIG. 14.



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FIG. 15a.

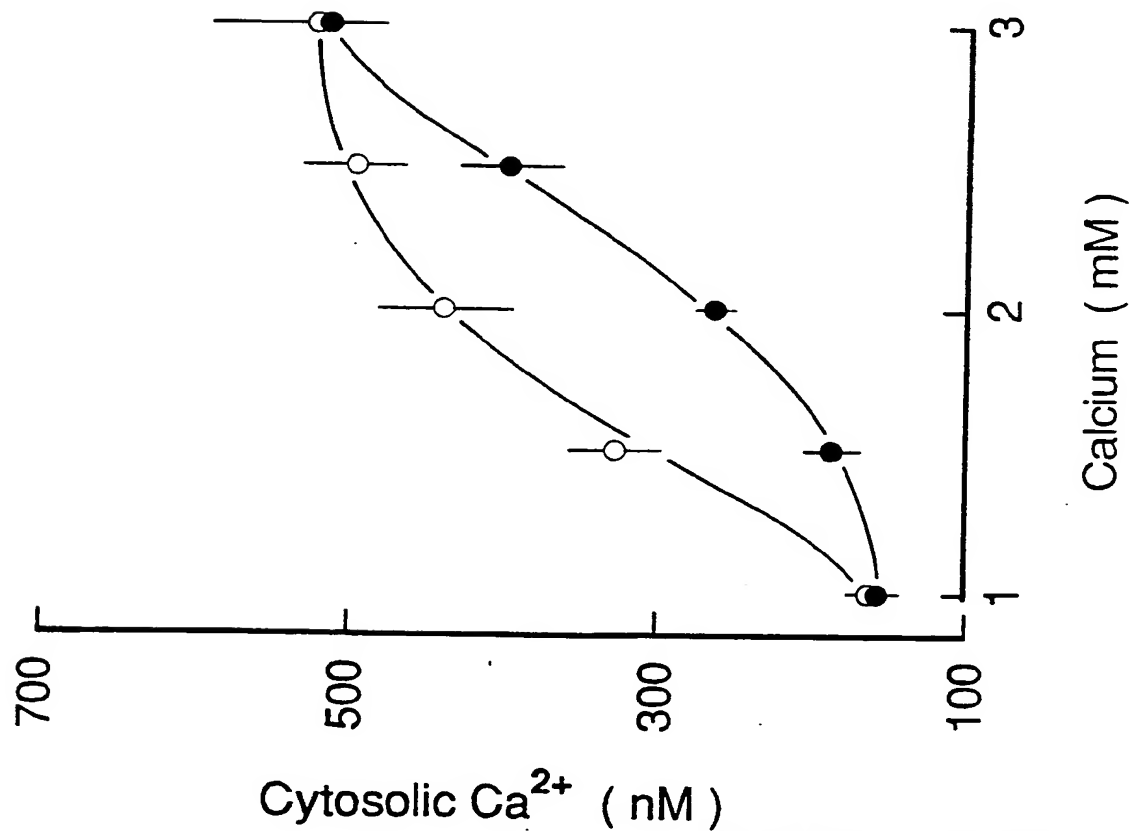
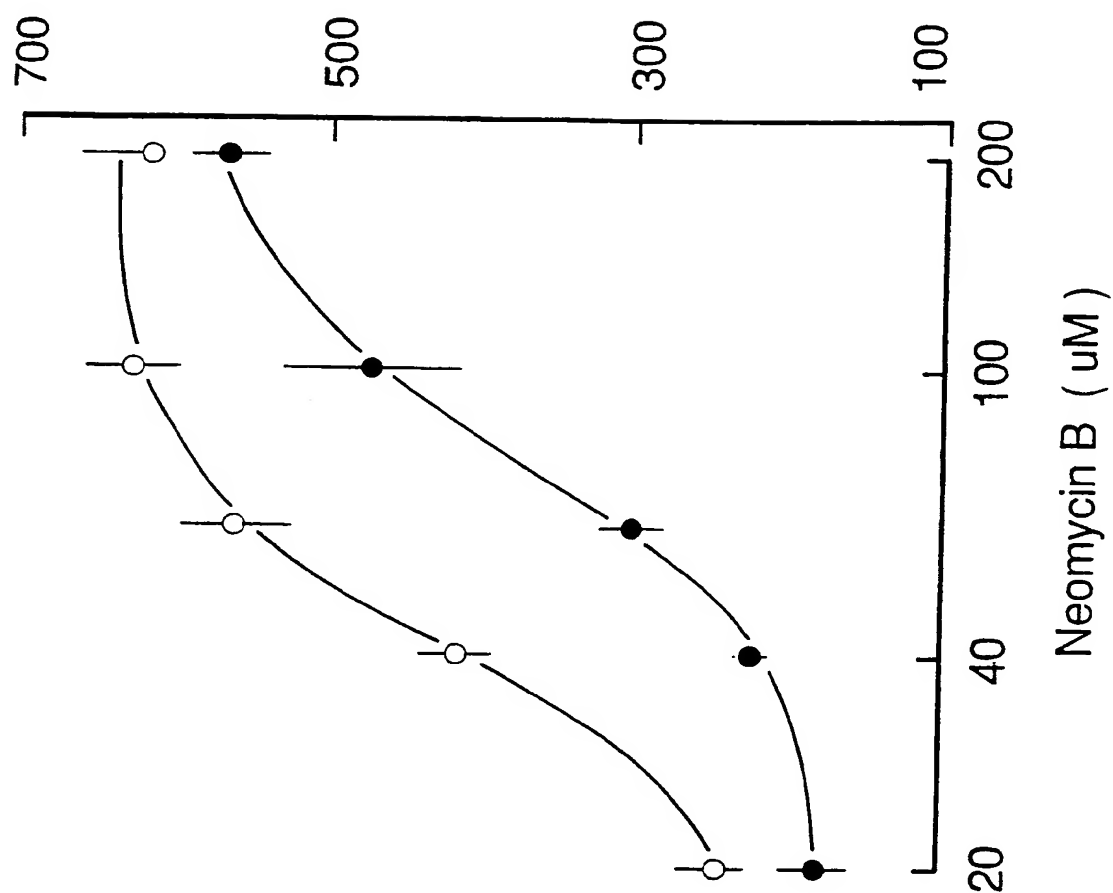


FIG. 15b.



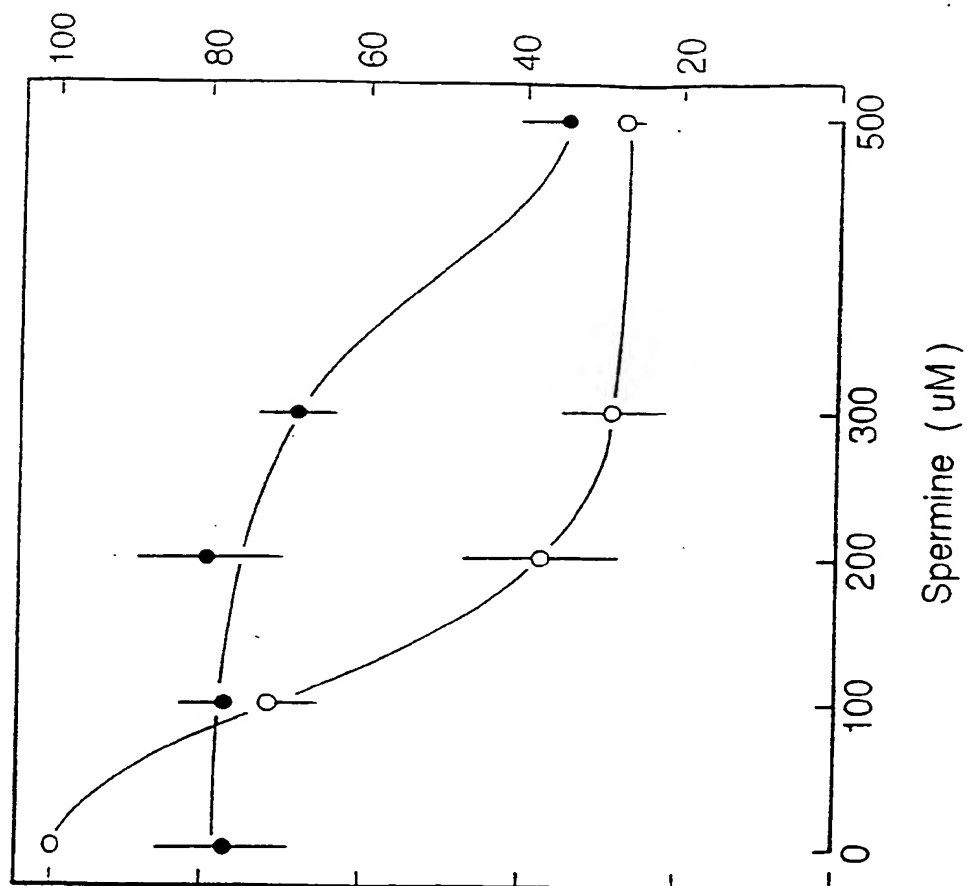


FIG. 16b.

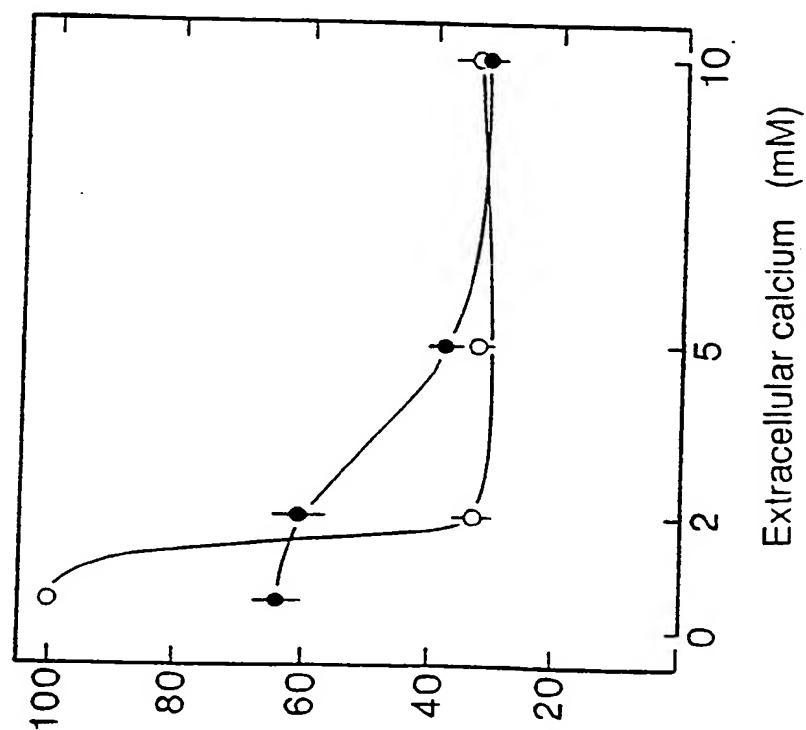


FIG. 16a.

PTH Secretion  
(percent of control)

**SUBSTITUTE SHEET**

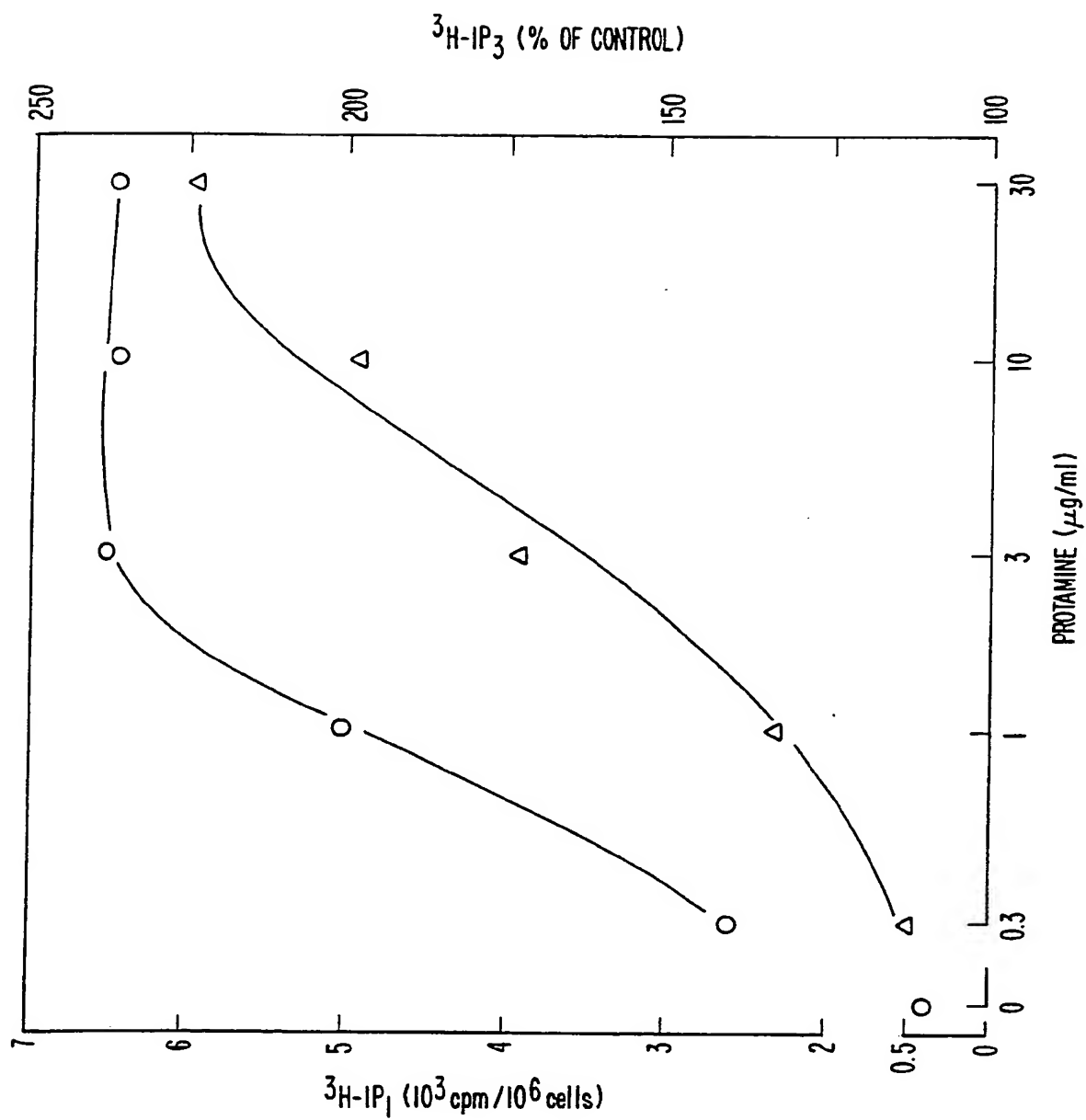


FIG. 17.

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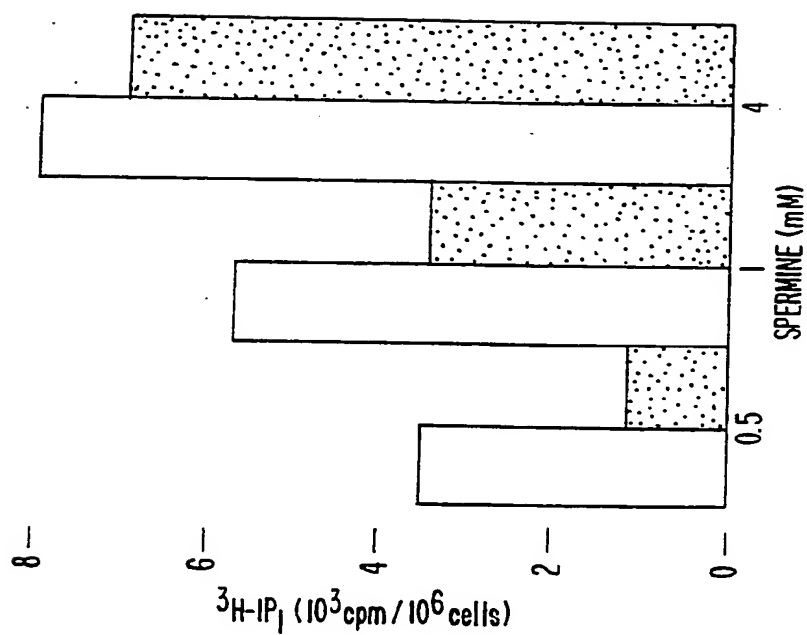


FIG. 18b.

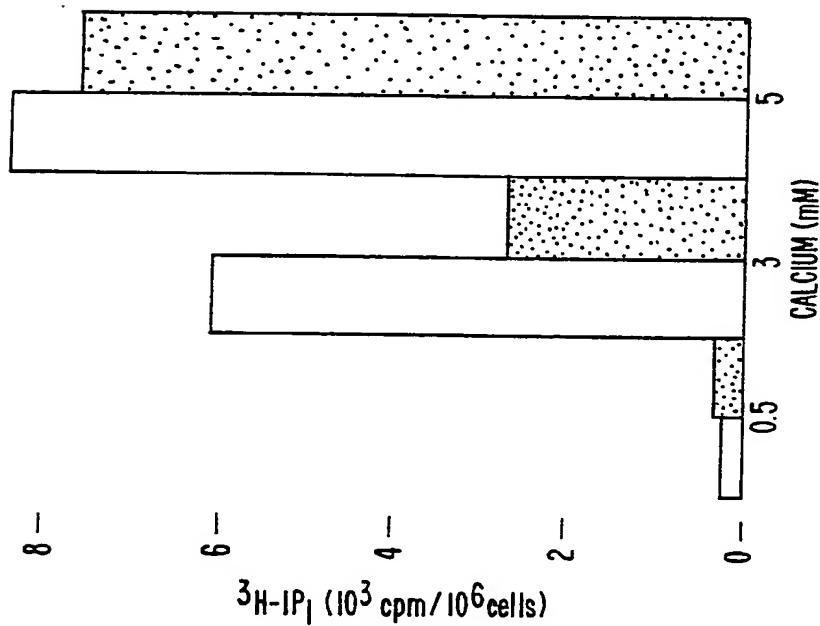


FIG. 18a.

SUBSTITUTE SHEET

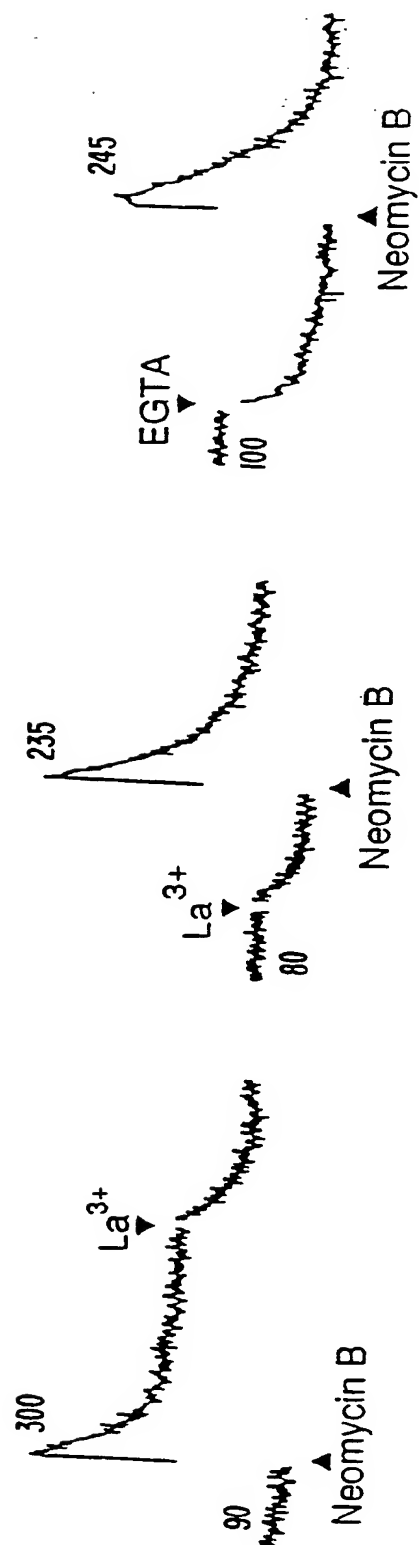


FIG. 19.

FIG. 20a.

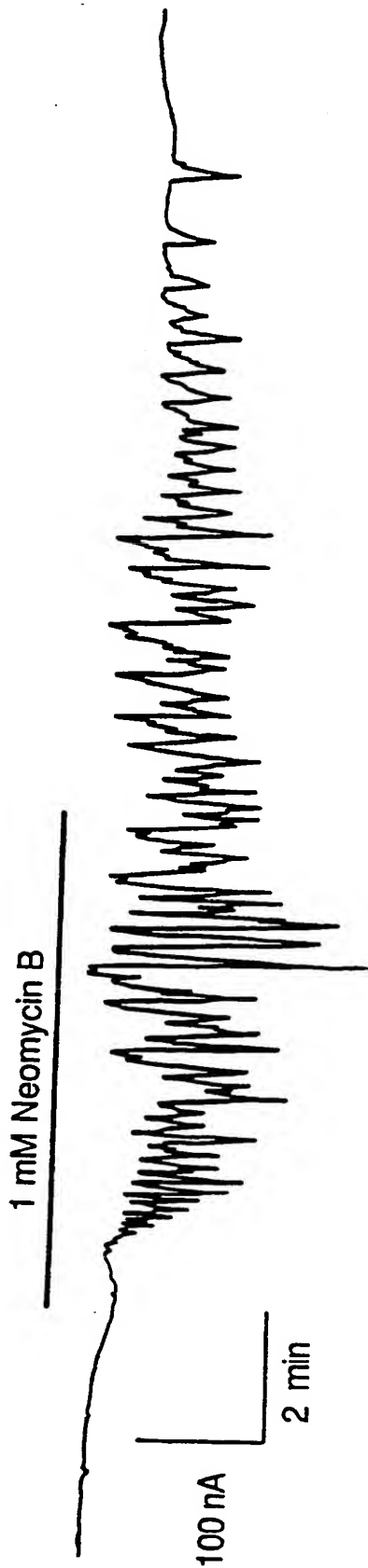
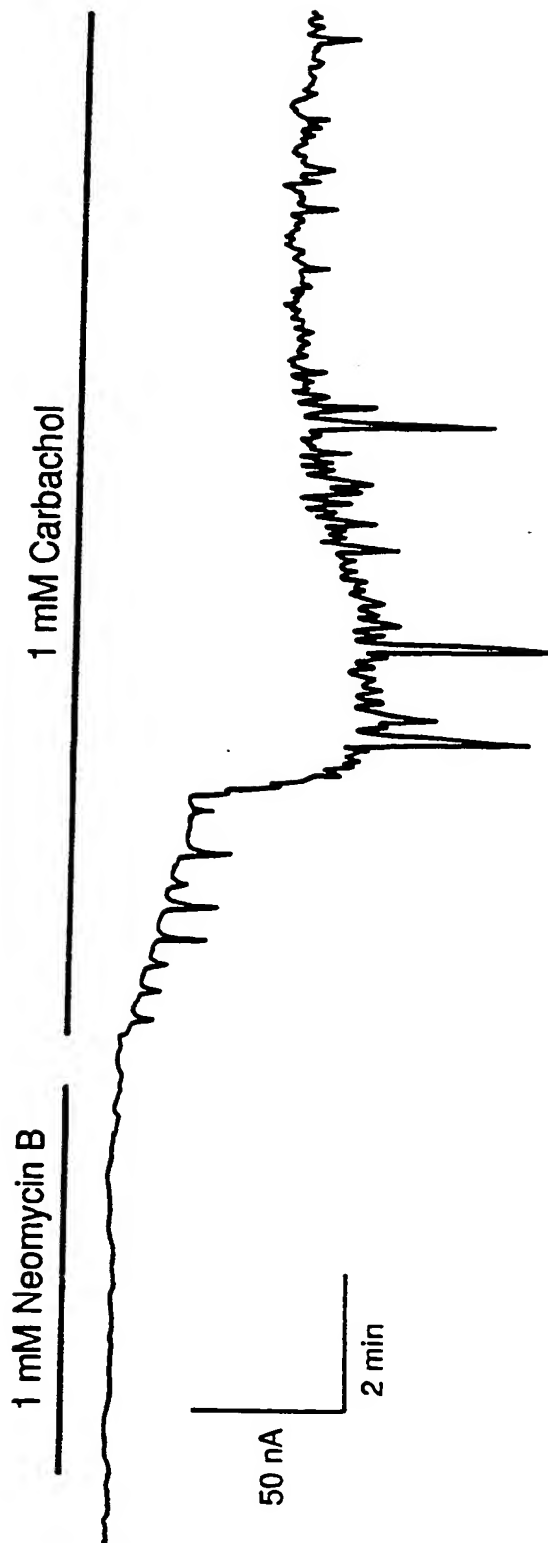
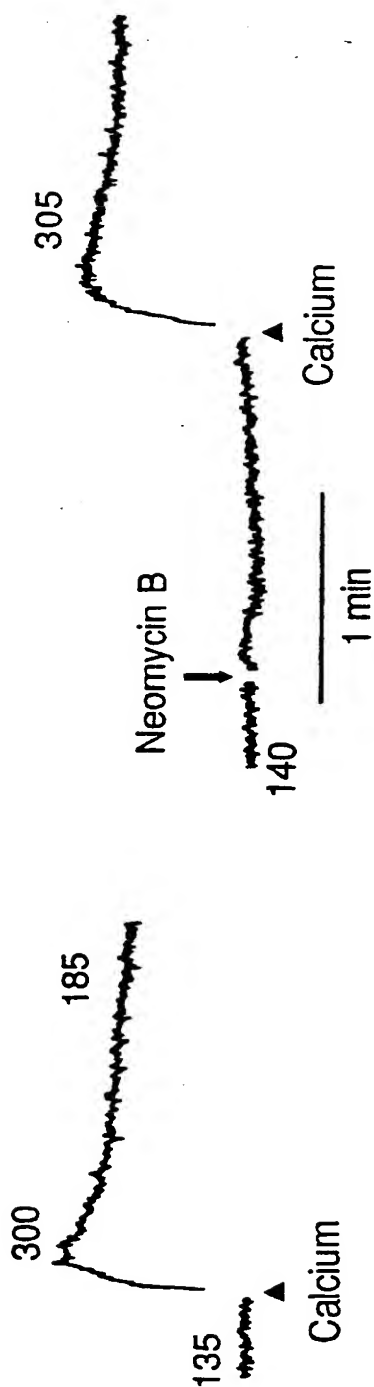


FIG. 20b.



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FIG. 21.



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FIG. 22.

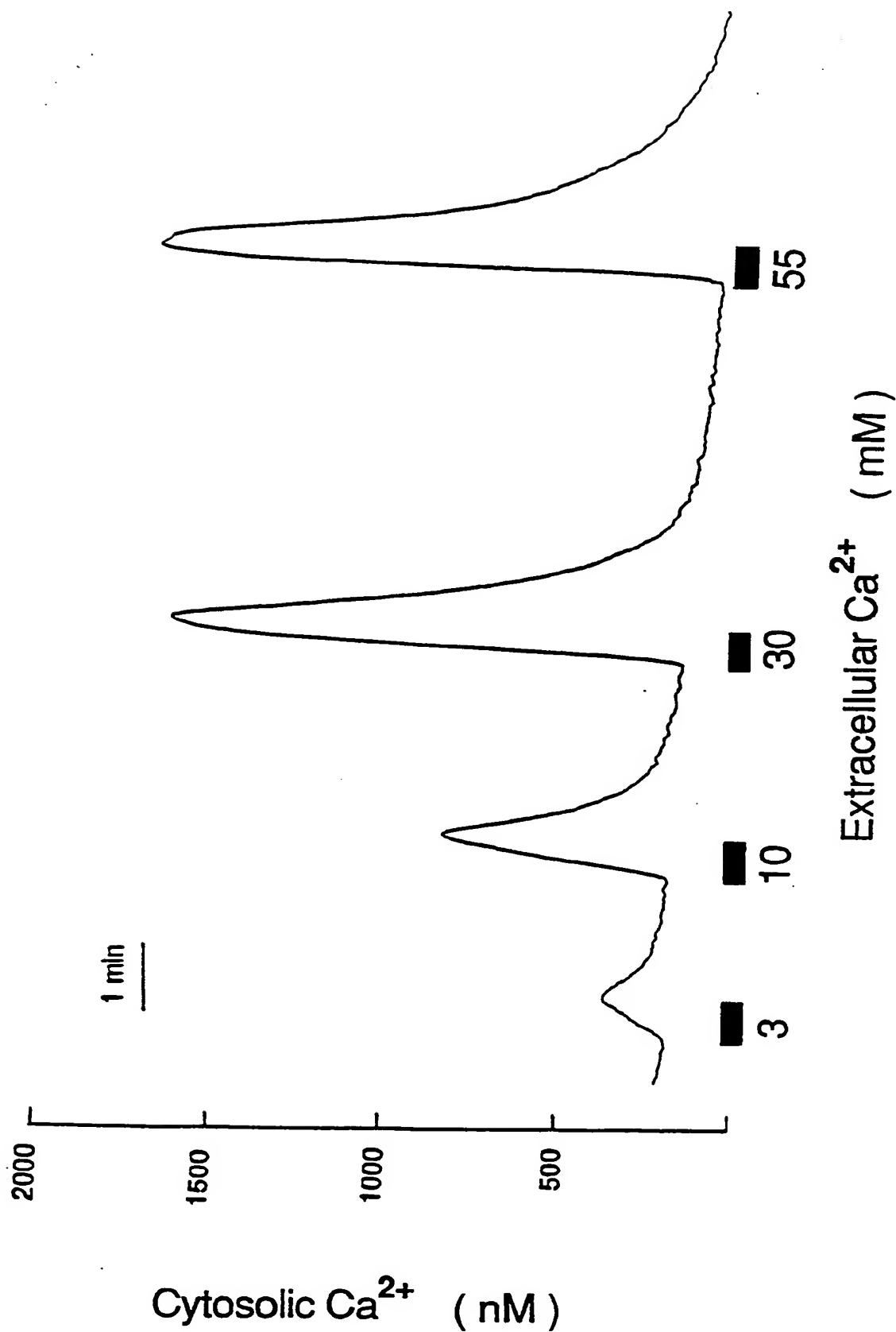
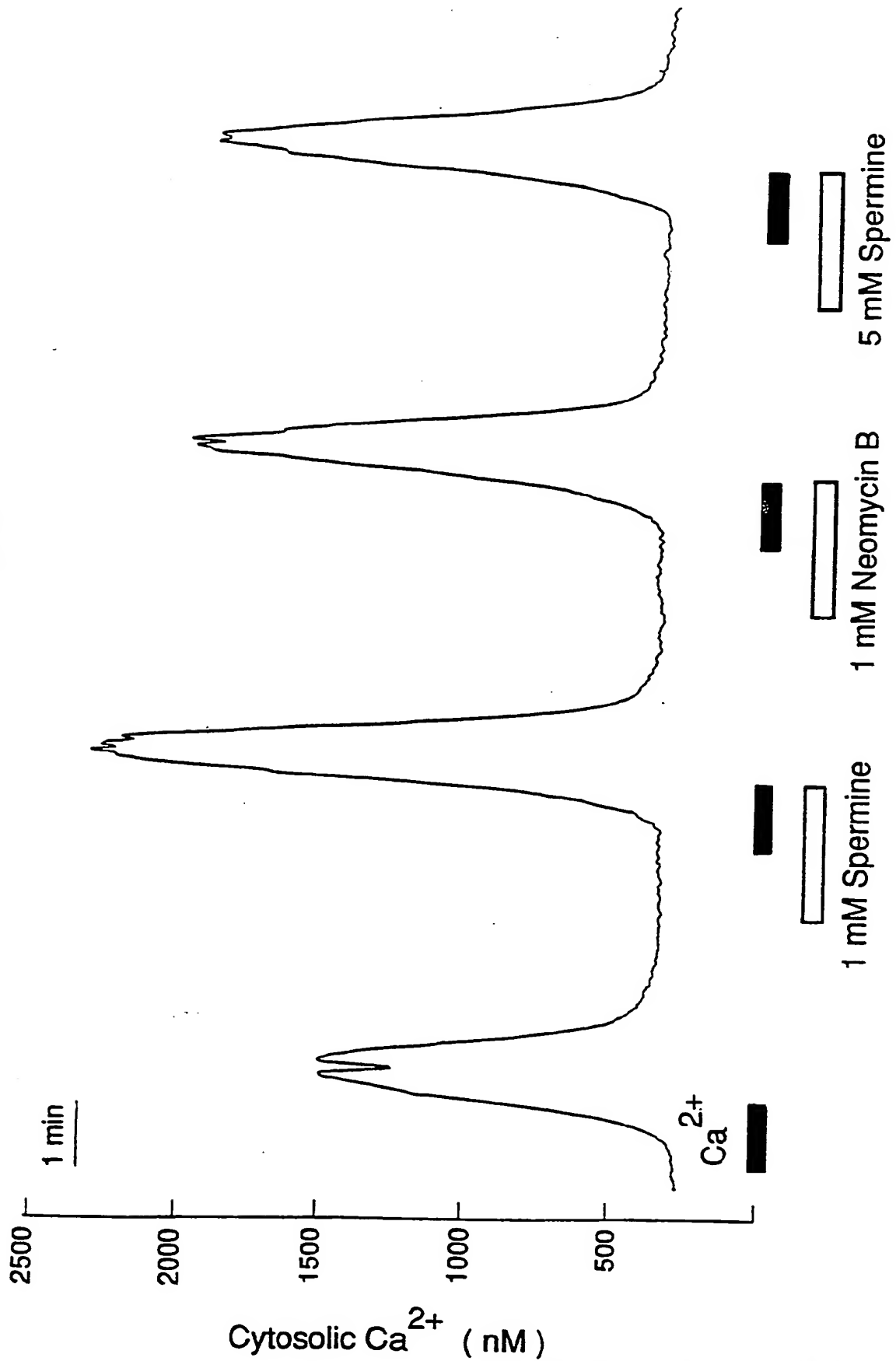


FIG. 23.



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FIG. 24.

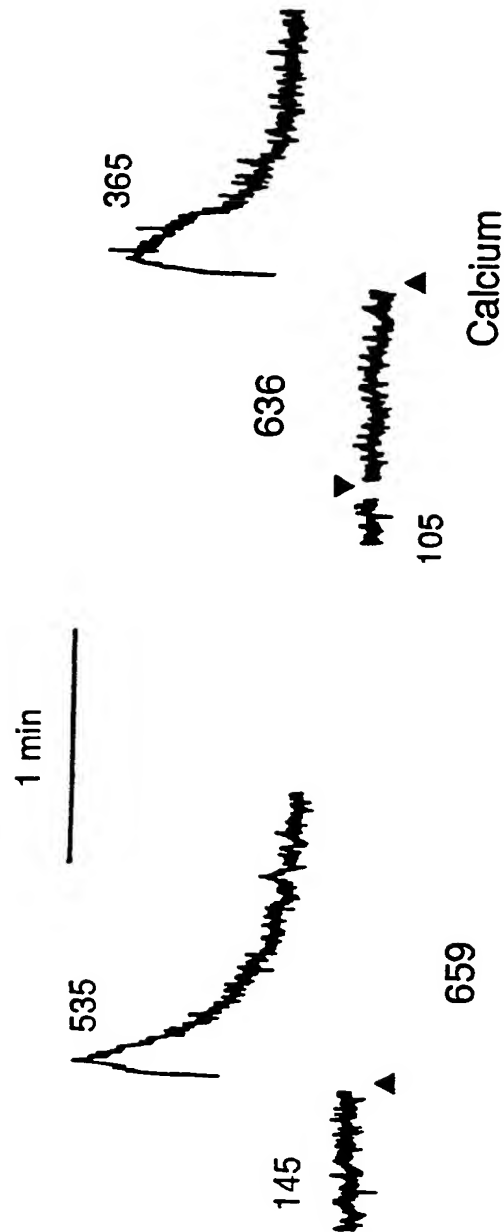


FIG. 25a.

10 mM  $Mg^{2+}$



FIG. 25b.

600  $\mu M$   $Gd^{3+}$

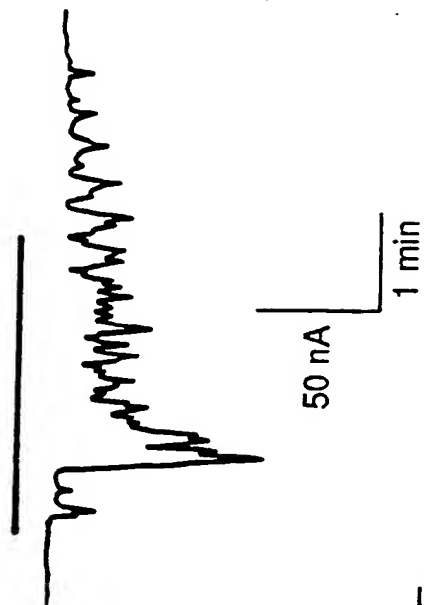


FIG. 25c.

10 mM  $Mg^{2+}$

100 nM Substance K



FIG. 26.

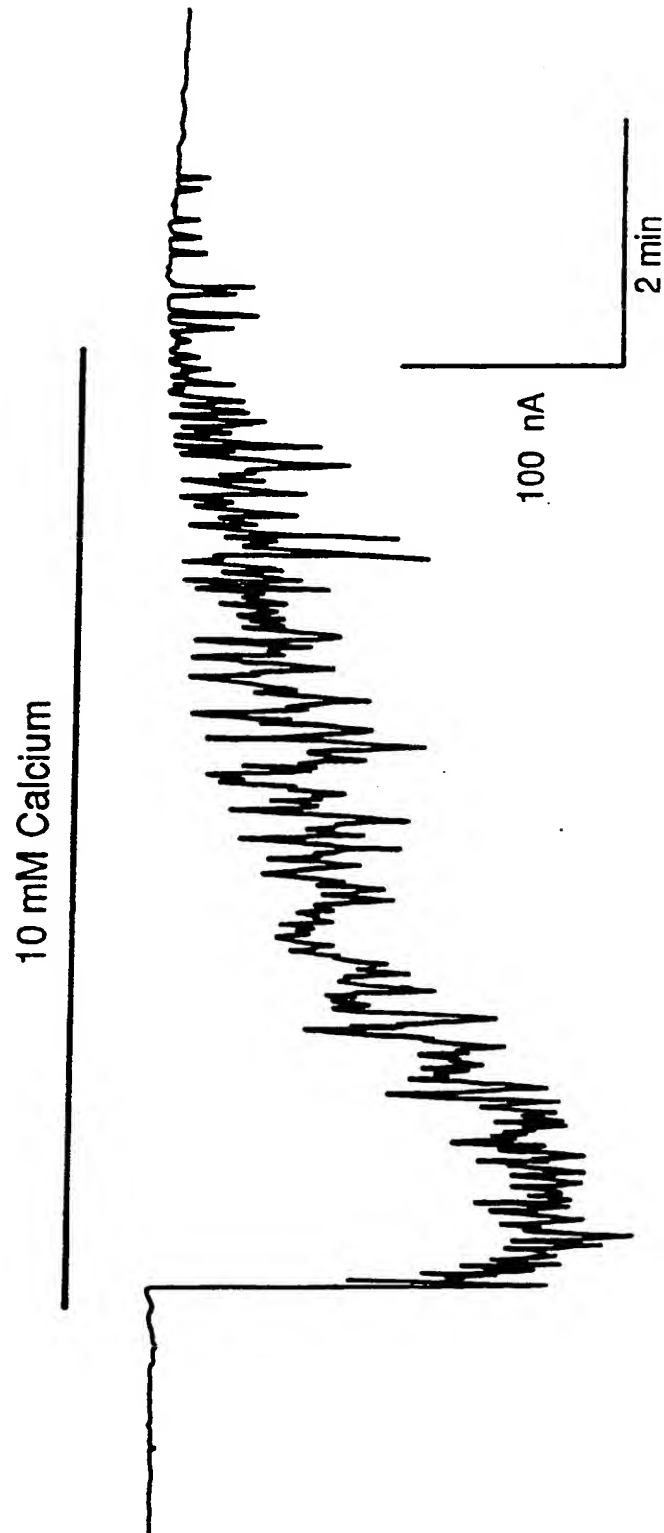
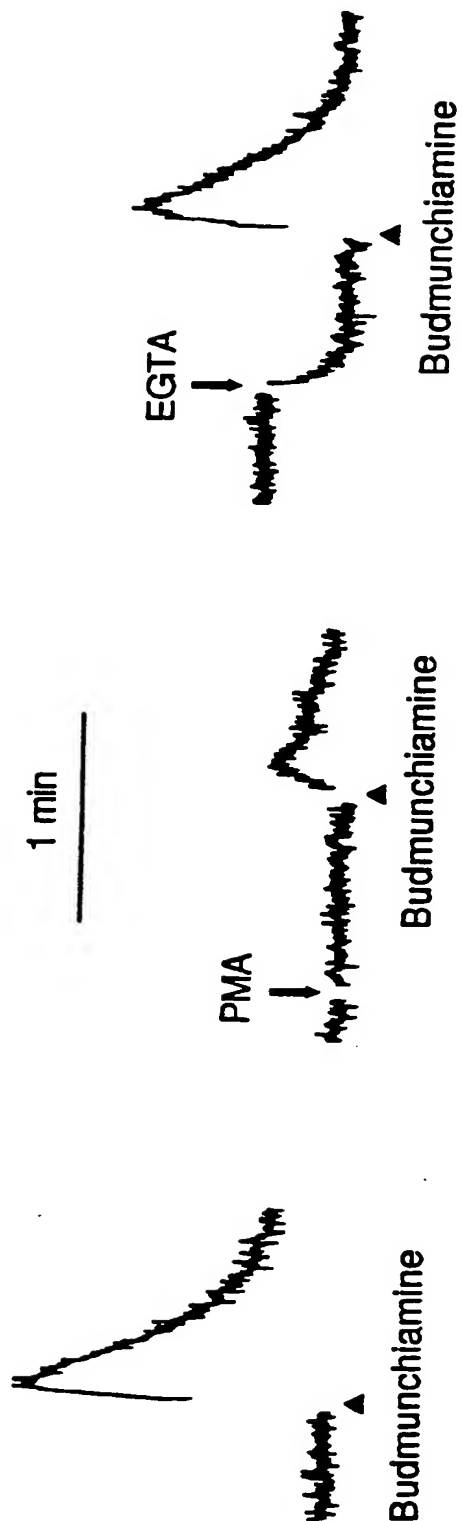


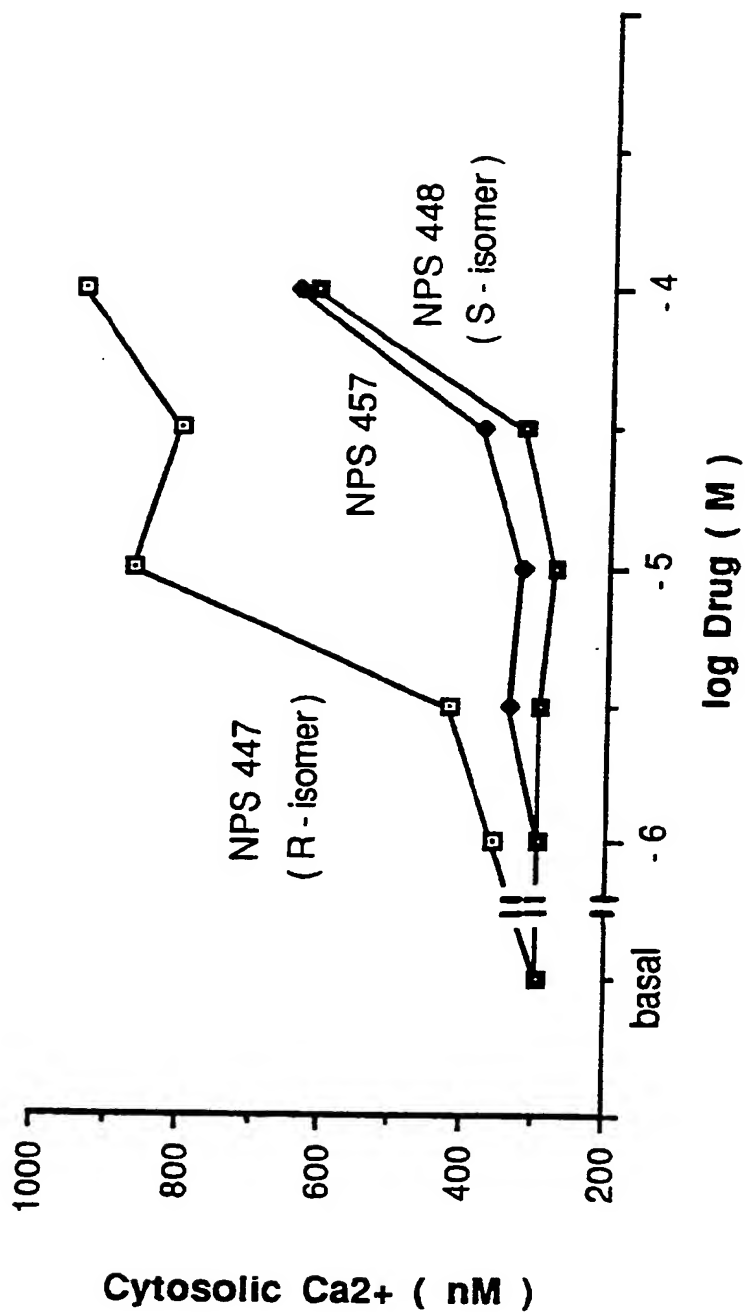
FIG. 27.



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FIG. 28.



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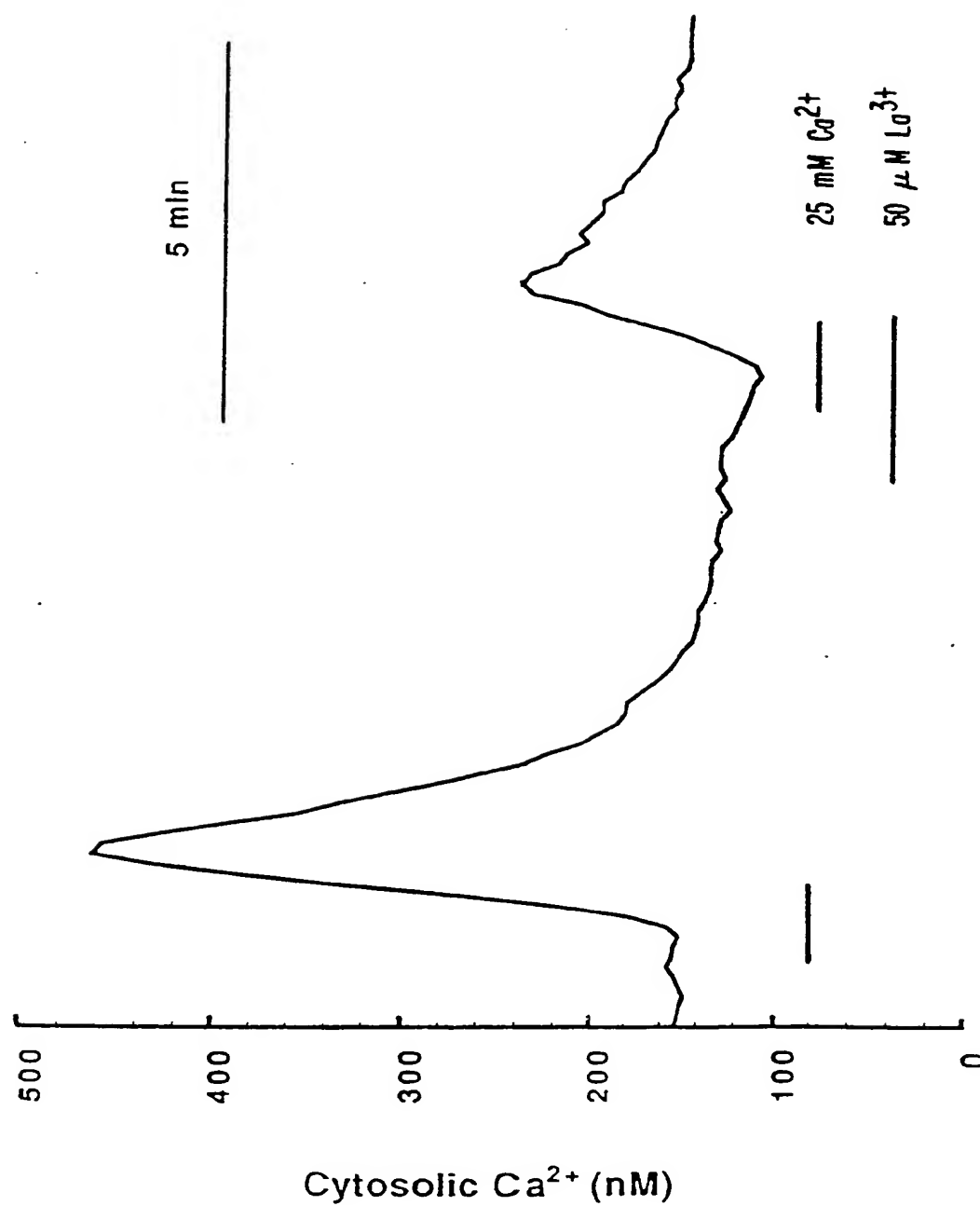


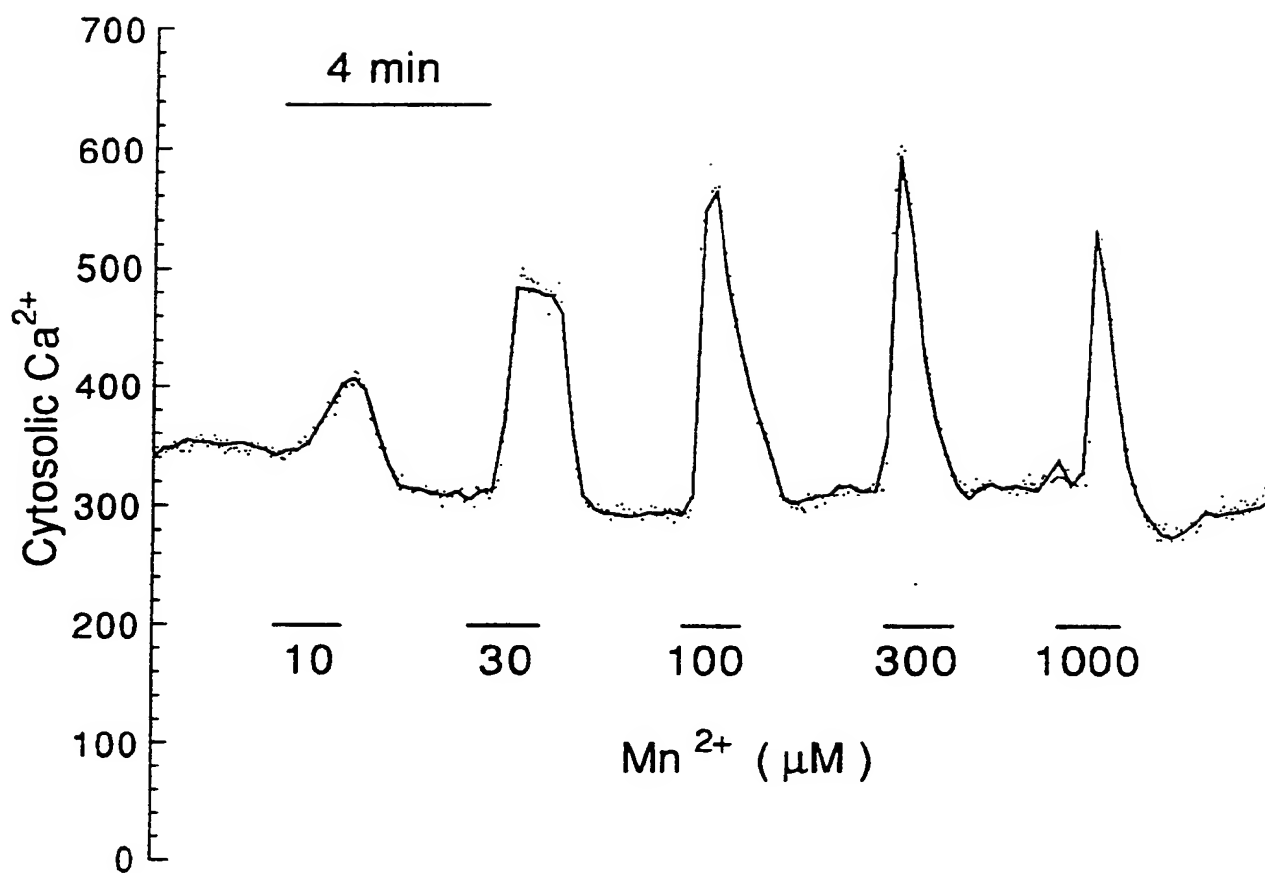
FIG. 29.

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FIG. 30a.



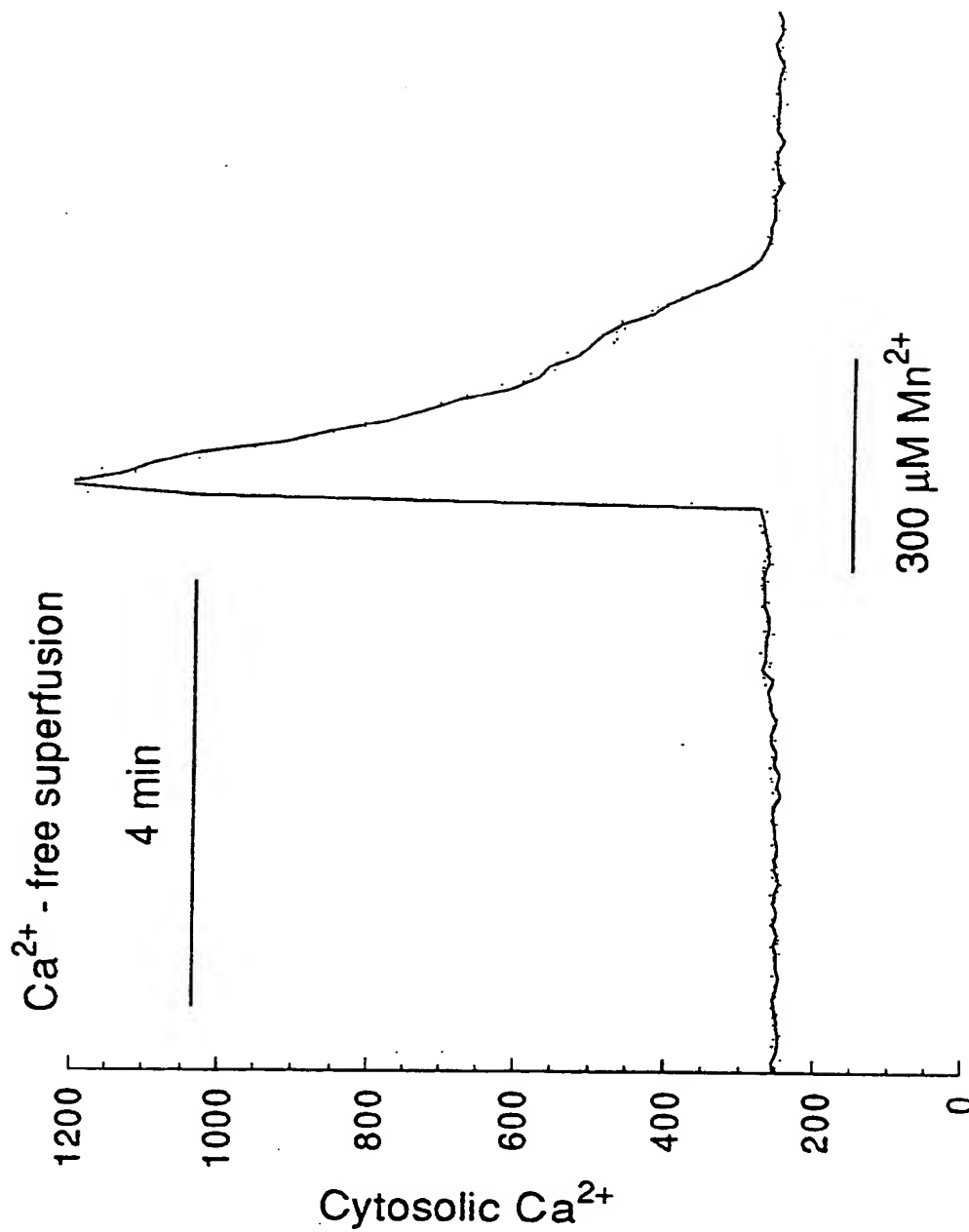
(a.)

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FIG. 30b.

(b.)



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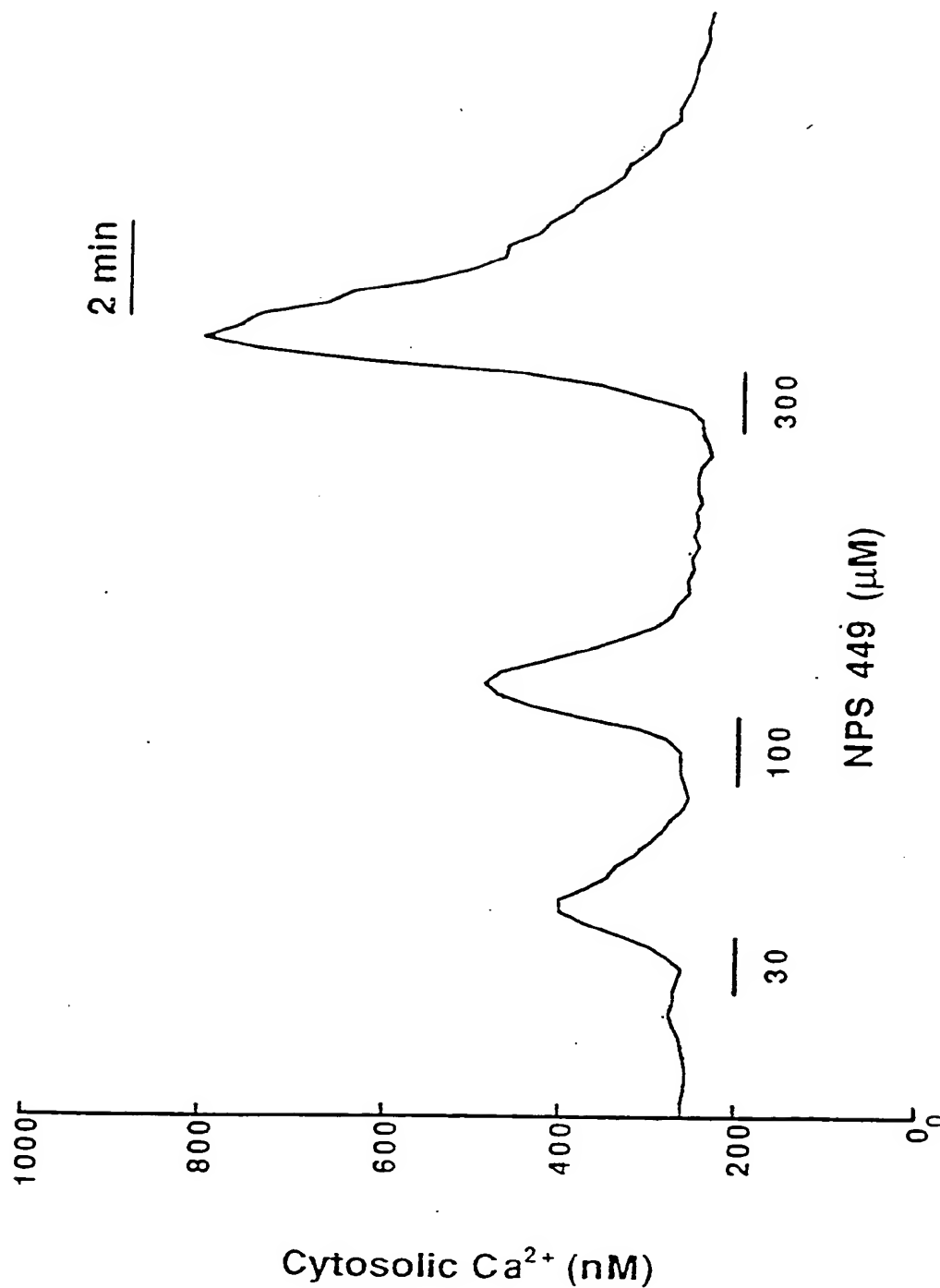


FIG. 31a.

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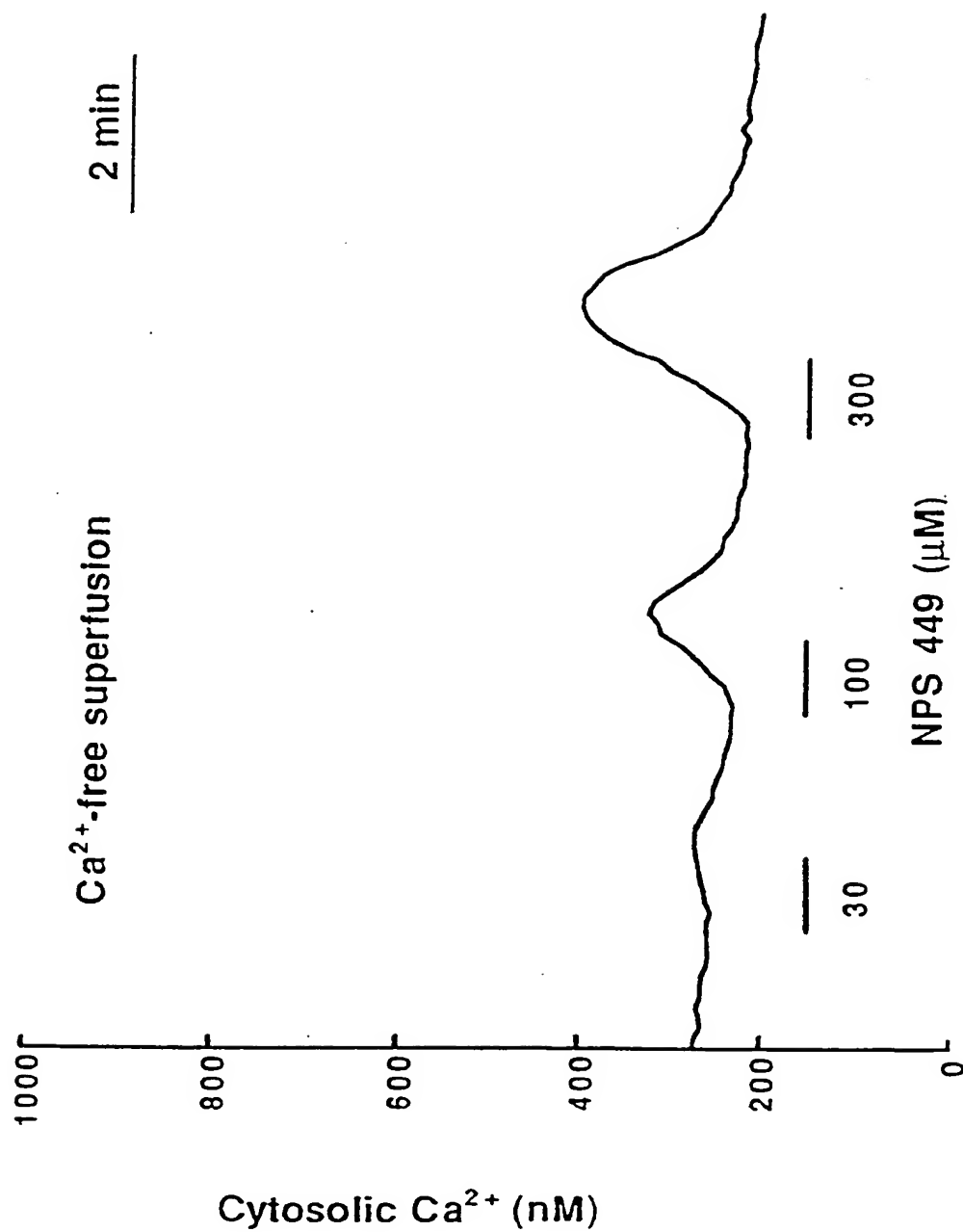


FIG. 31b.

SUBSTITUTE SHEET

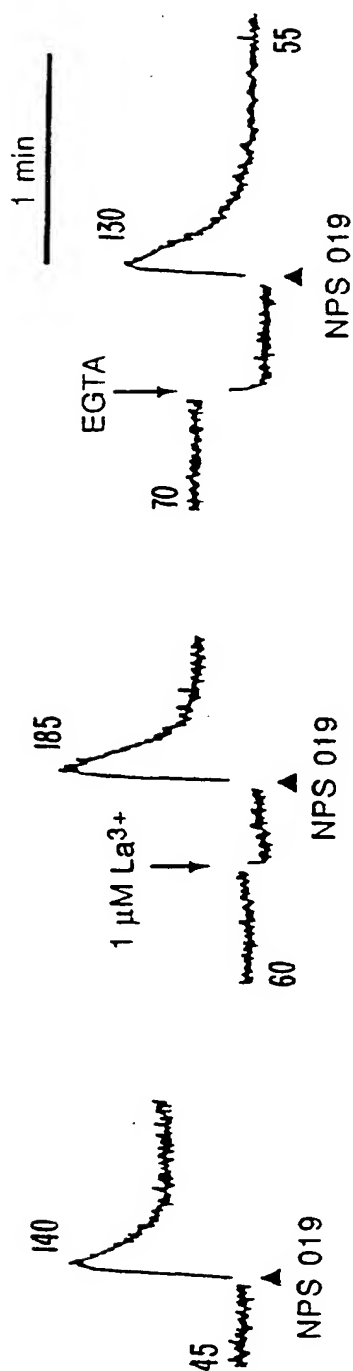
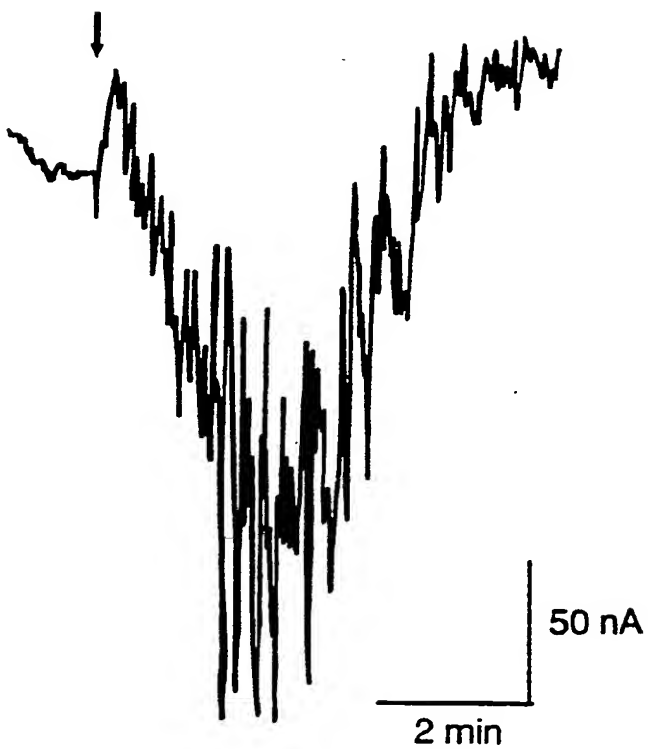


FIG. 32.

SUBSTITUTE SHEET

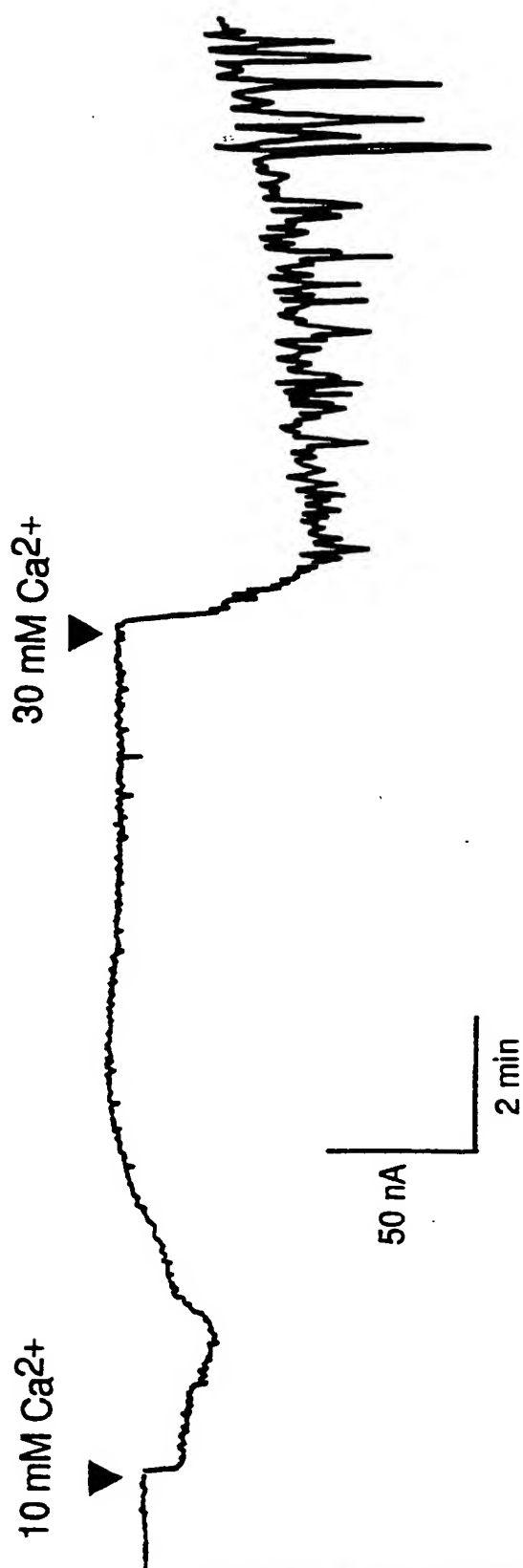
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FIG. 33.

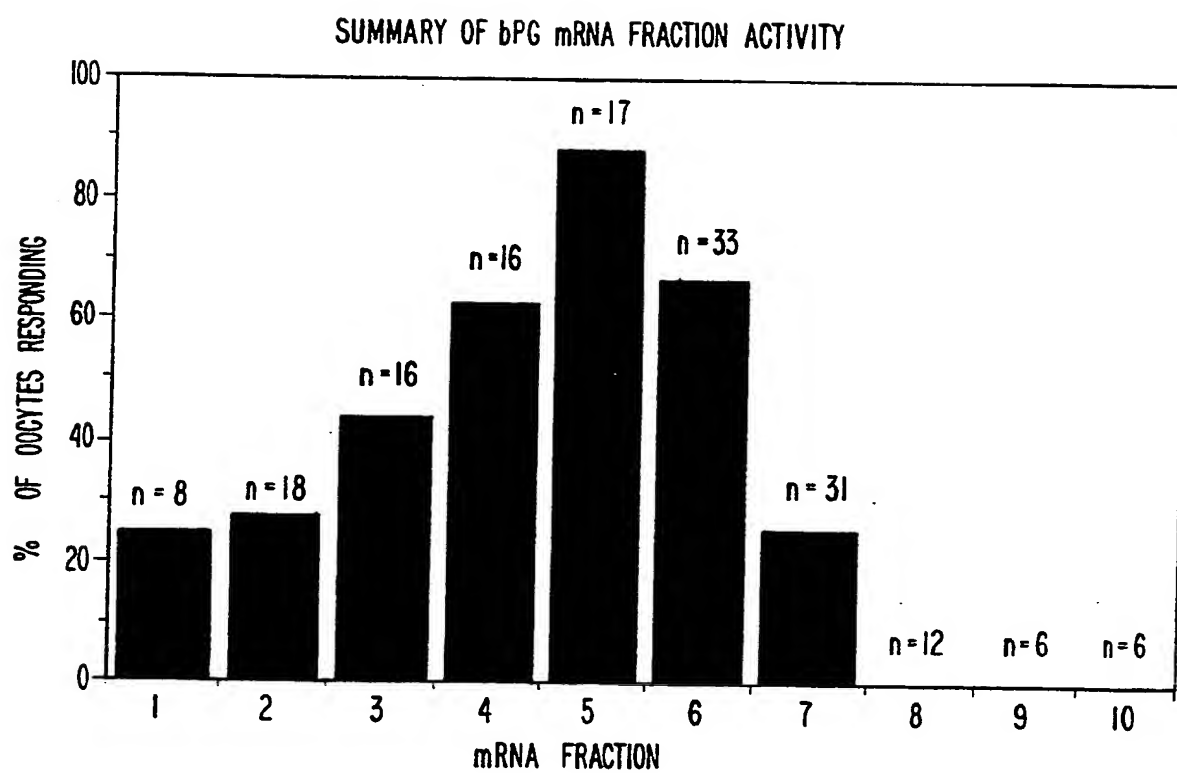
100  $\mu$ M NPS 456

SUBSTITUTE SHEET

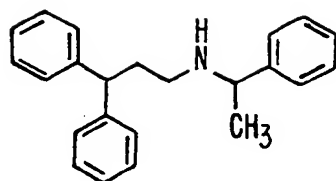
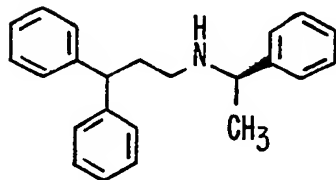
FIG. 34.



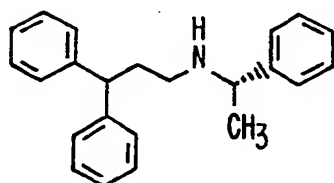
SUBSTITUTE SHEET

*FIG. 35.***SUBSTITUTE SHEET**

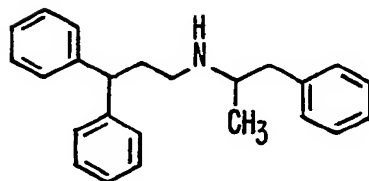


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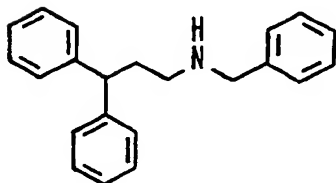
447



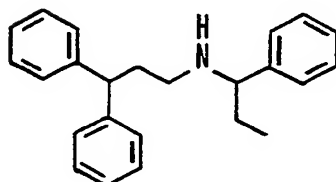
448



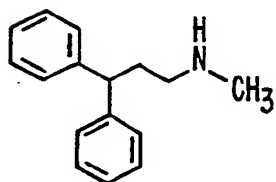
449



455



460

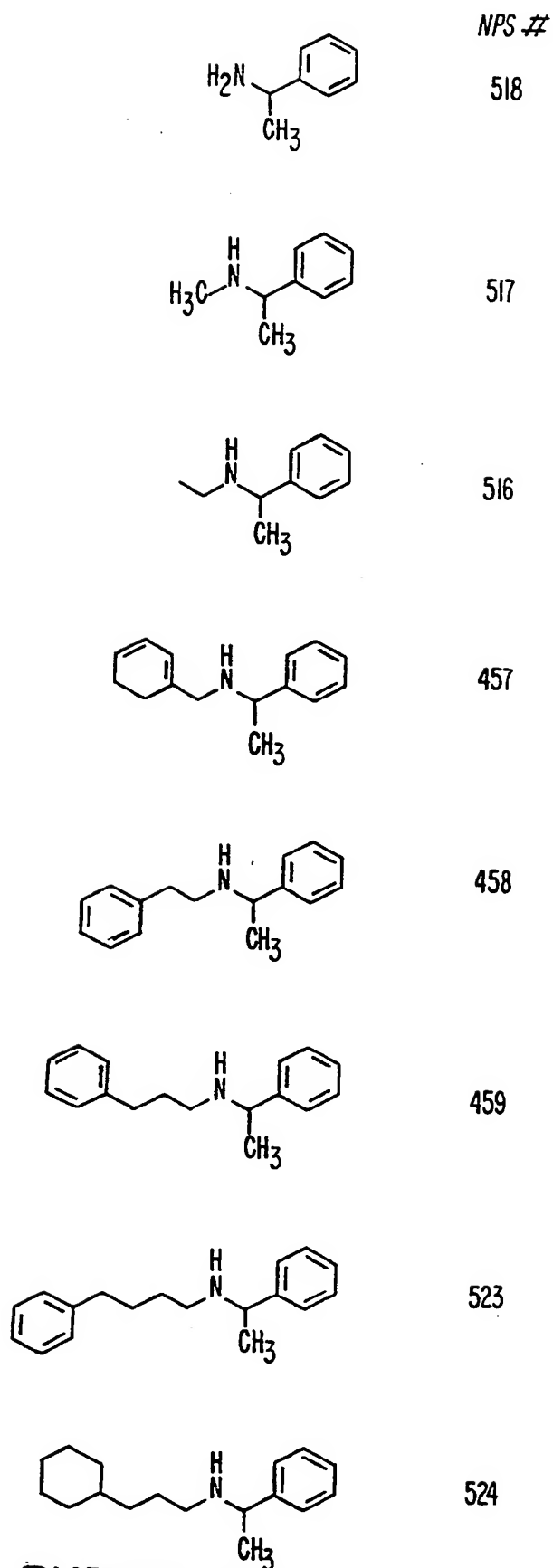


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FIG. 36a.

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FIG. 36b.

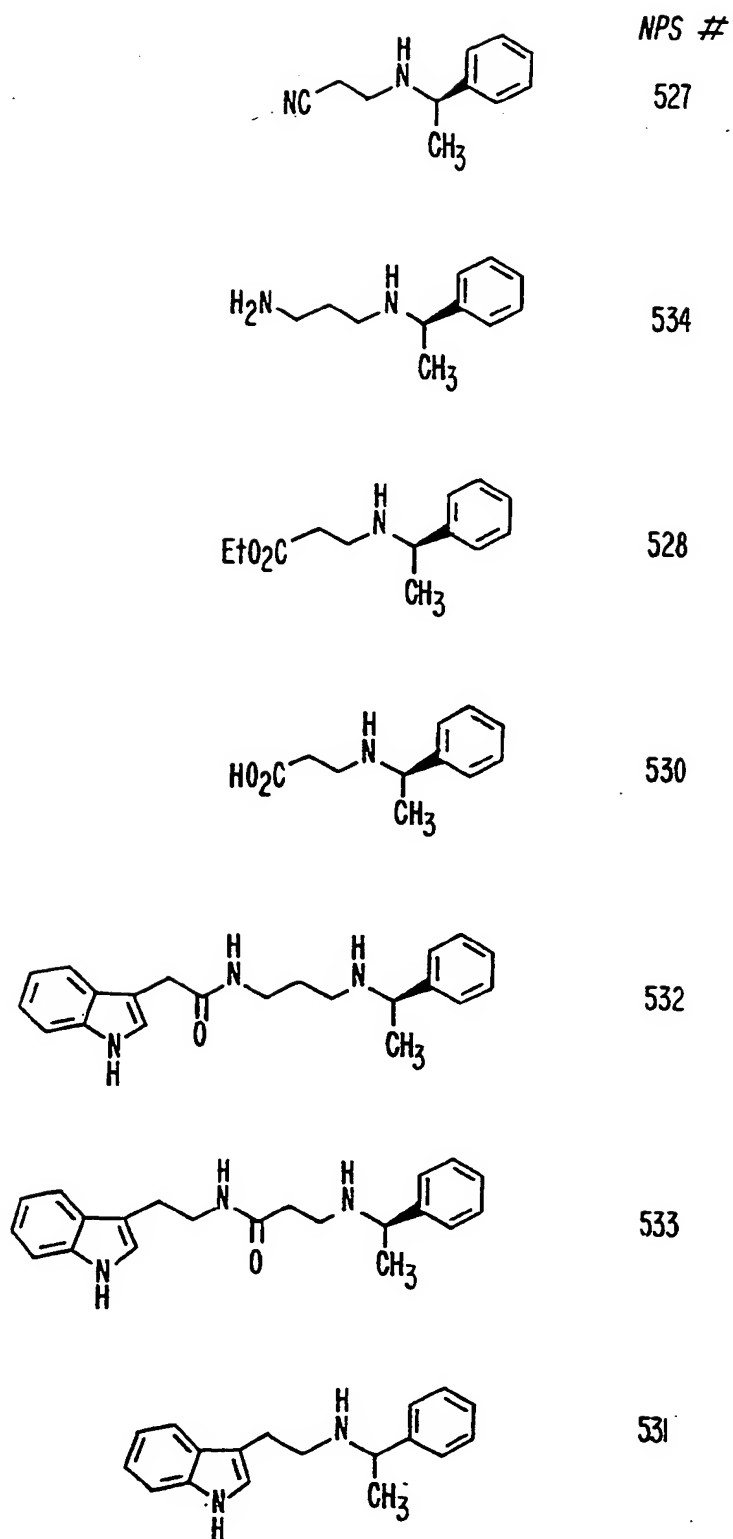


FIG. 36c.

SUBSTITUTE SHEET

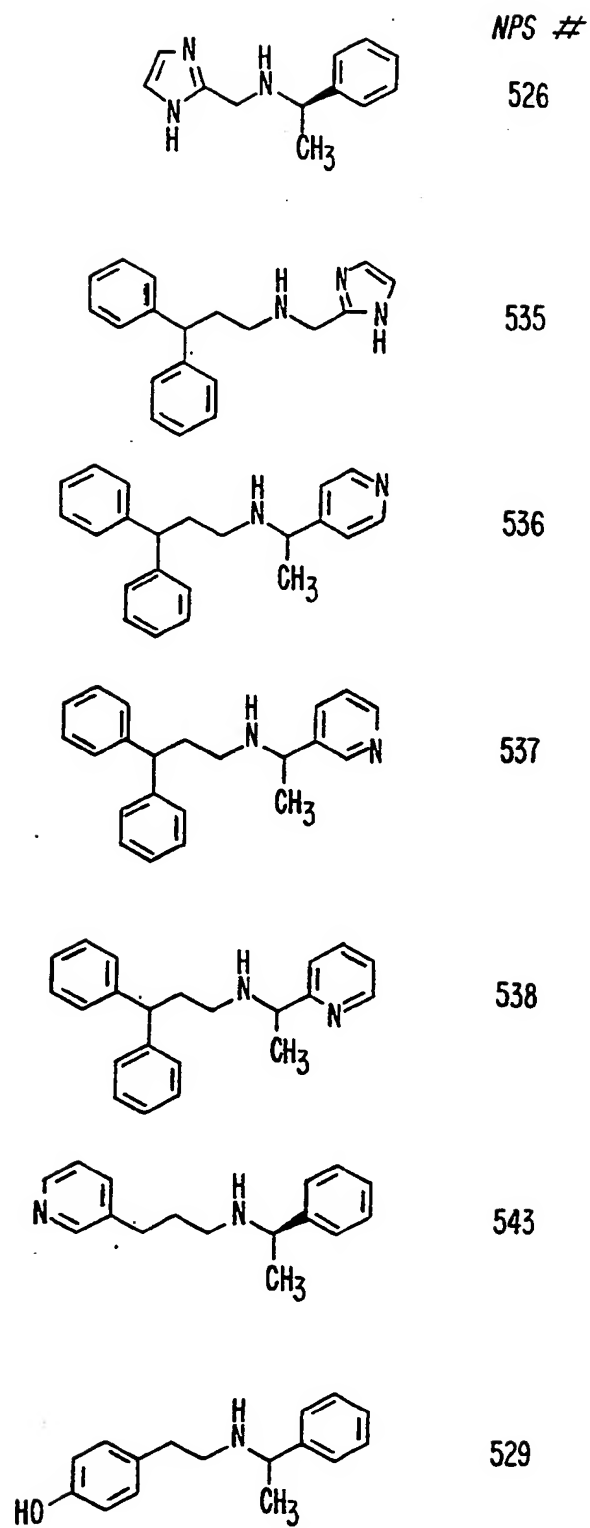


FIG. 36d.

SUBSTITUTE SHEET

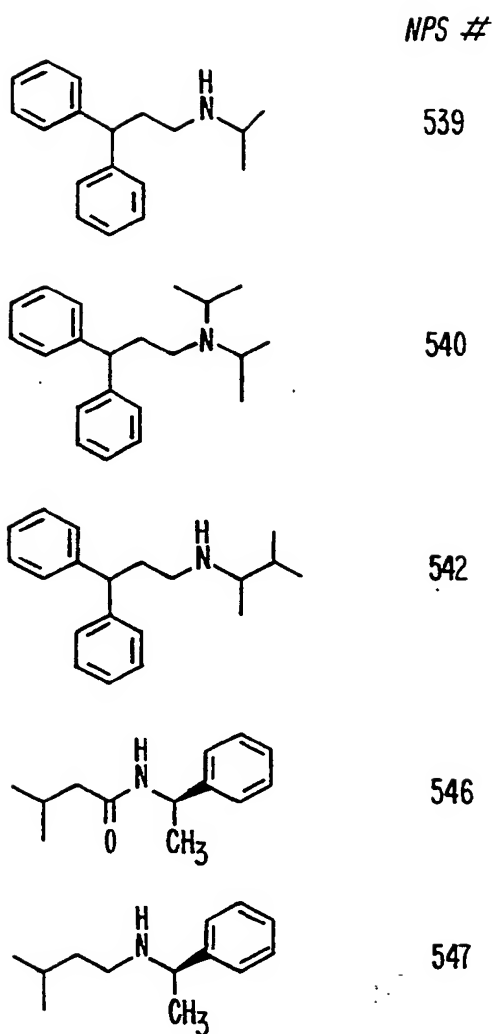


FIG. 36e.

SUBSTITUTE SHEET

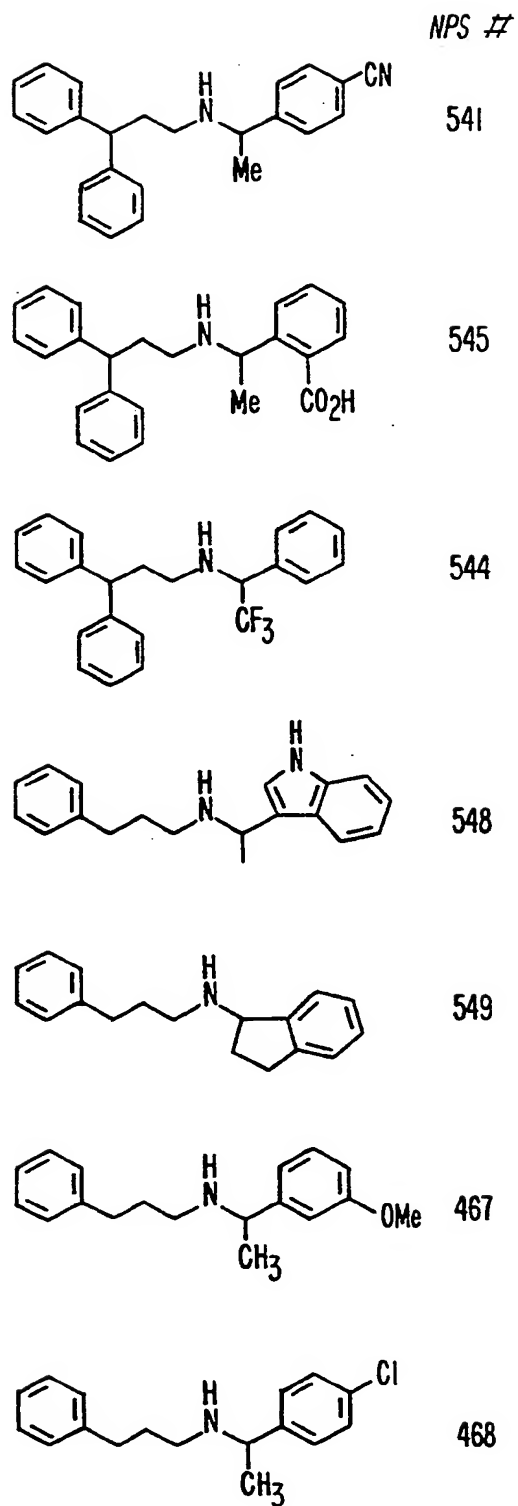


FIG. 36f.

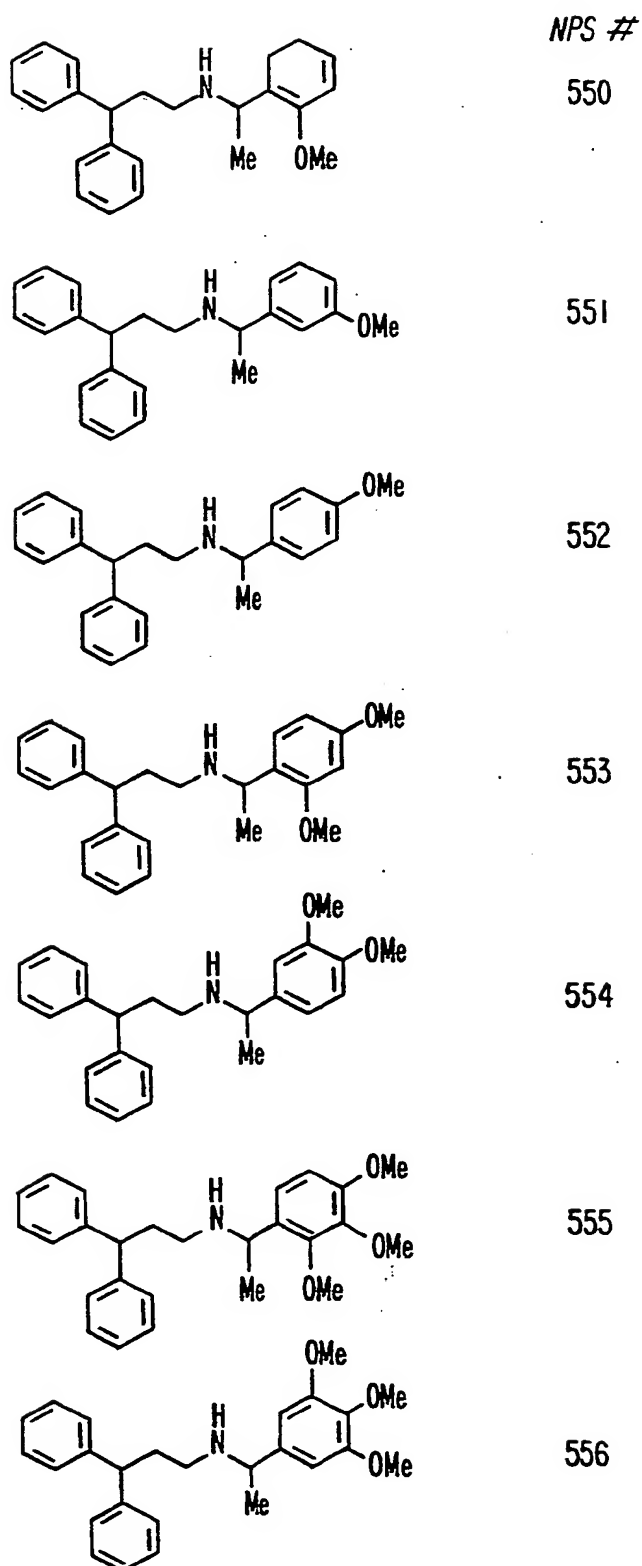
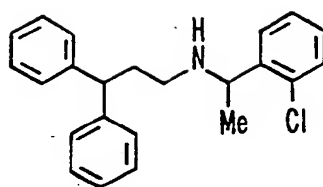
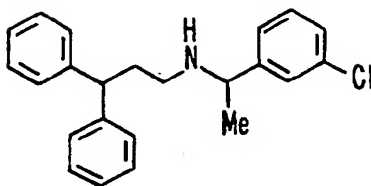
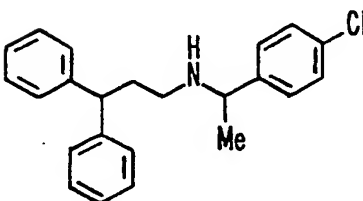


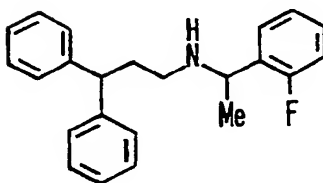
FIG. 36g.

NPS #  
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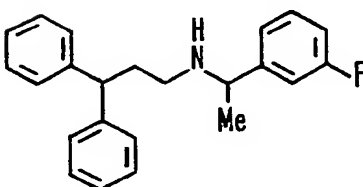
558



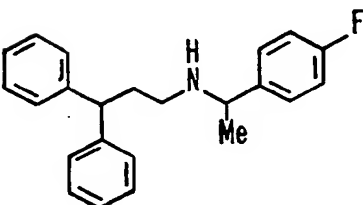
559



560



561

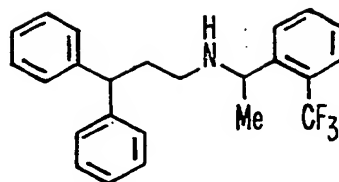


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FIG. 36h.

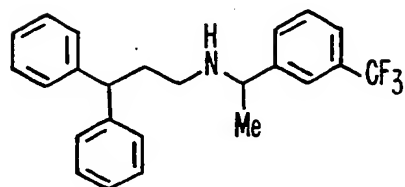
SUBSTITUTE SHEET



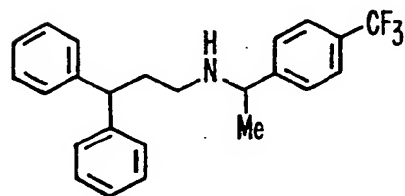


NPS #

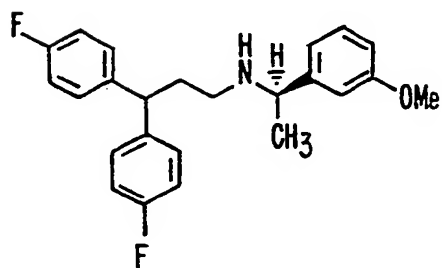
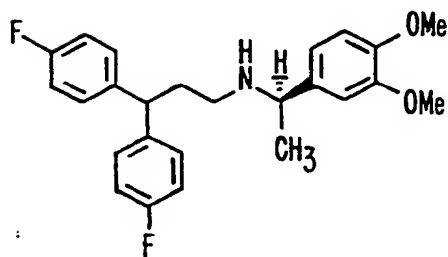
563



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565

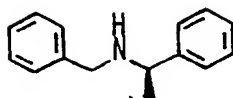


606

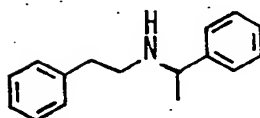
FIG. 36i.

SUBSTITUTE SHEET

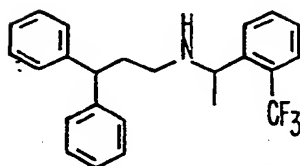
NPS #



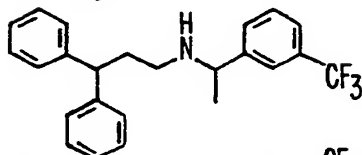
457



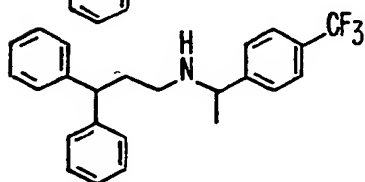
458



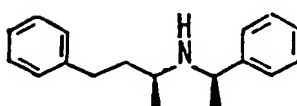
563



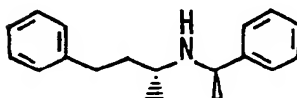
564



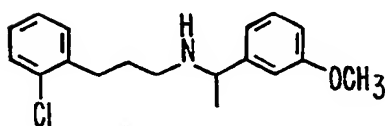
565



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568

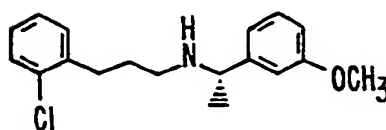
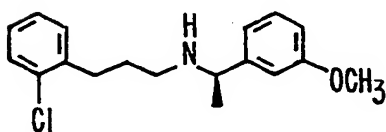


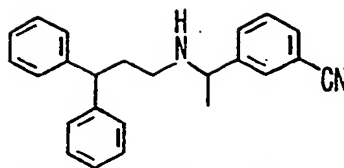
FIG. 36j.

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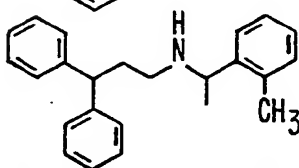
54/71

NPS #

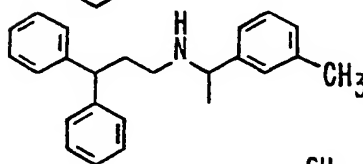
570



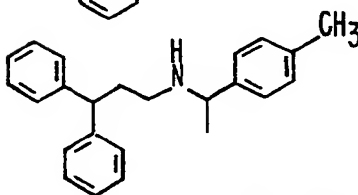
571



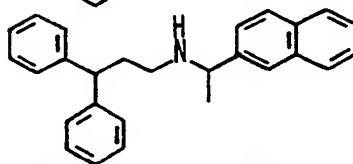
572



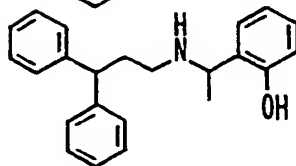
573



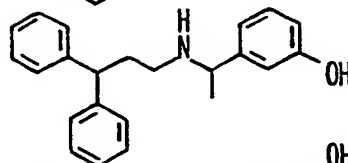
574



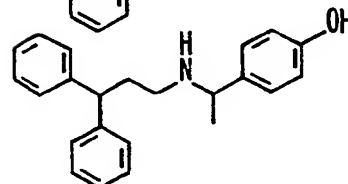
575



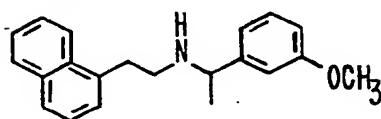
576



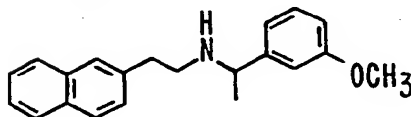
577



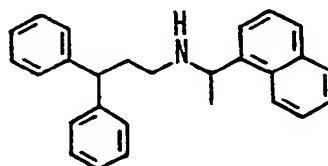
578



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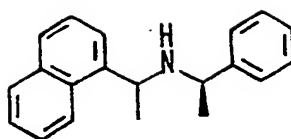
580



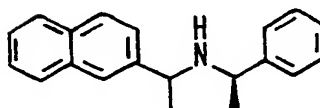
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FIG. 36k.

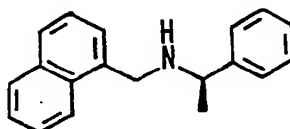
NPS #



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593

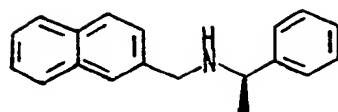


594

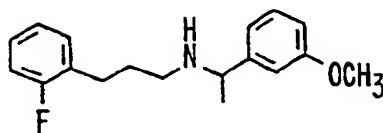
FIG. 36L.

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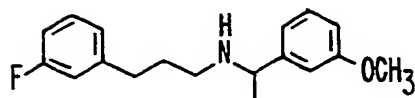
NPS #



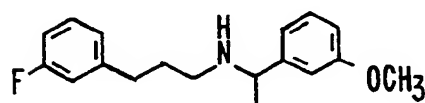
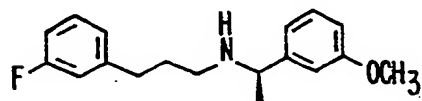
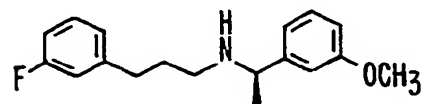
595



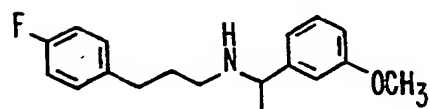
601



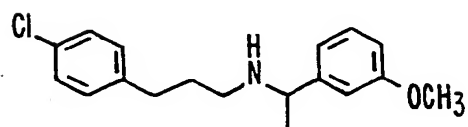
602



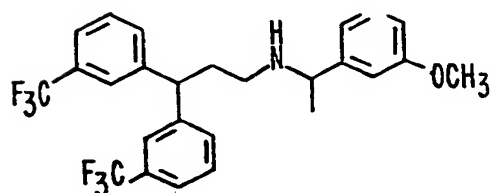
603



604



605



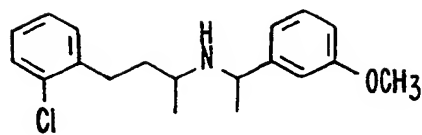
607

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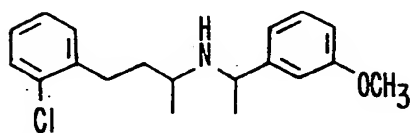
FIG. 36m.

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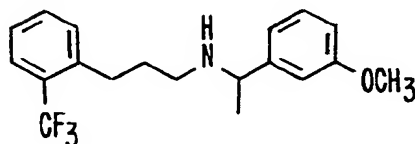
609



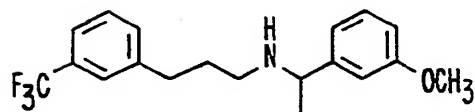
610



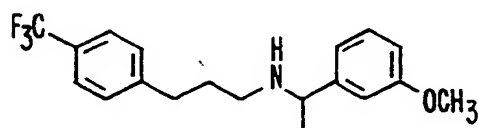
611



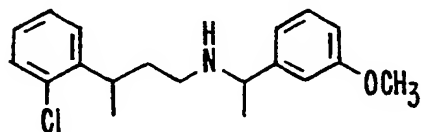
612



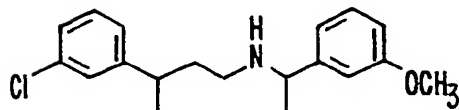
613



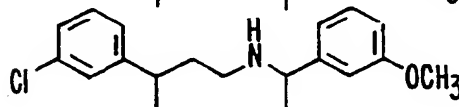
614



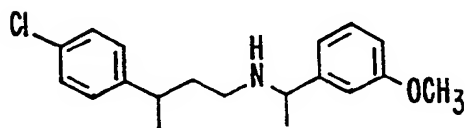
615



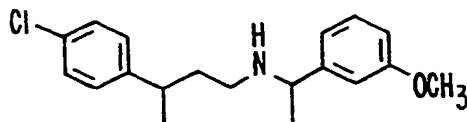
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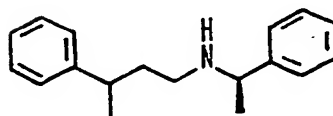
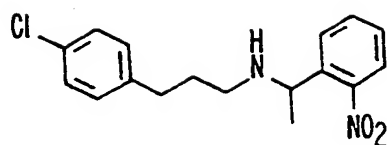
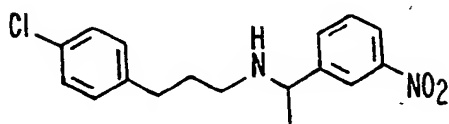
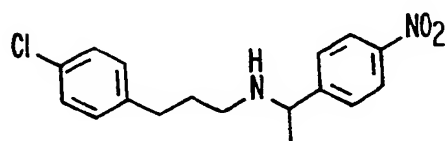


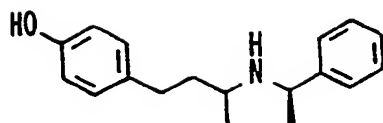
FIG. 36n

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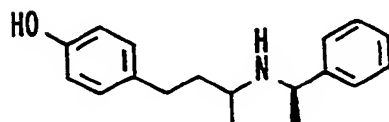
624



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631

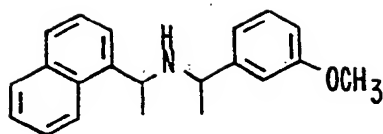


632

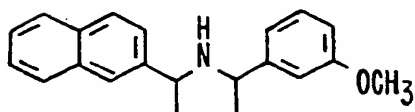
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FIG. 360.

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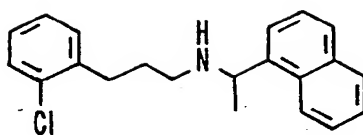
641



642

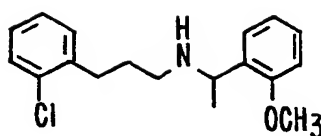
*FIG. 36p.***SUBSTITUTE SHEET**



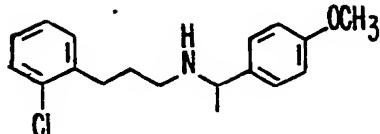


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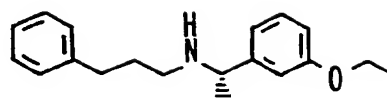
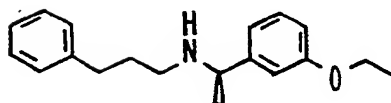
646



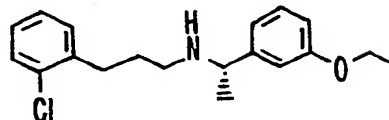
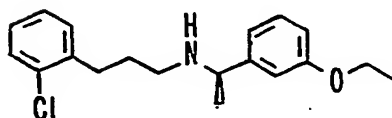
650



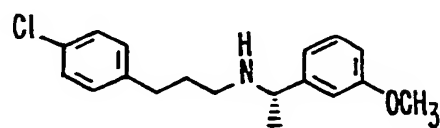
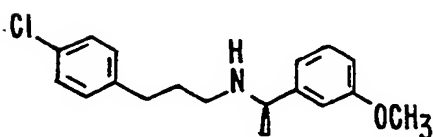
652



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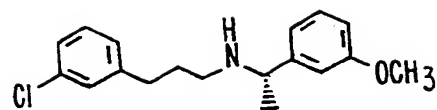
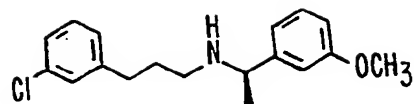
654



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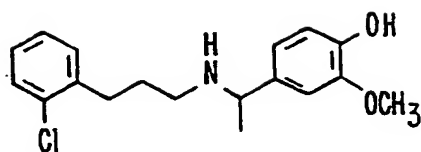
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FIG. 36q.



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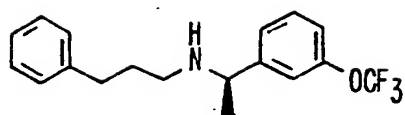


684

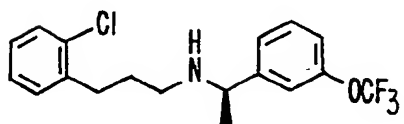
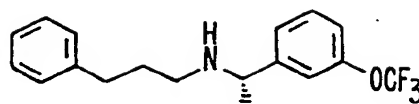
FIG. 36r.

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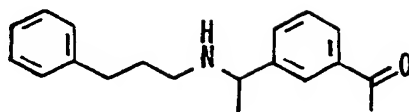
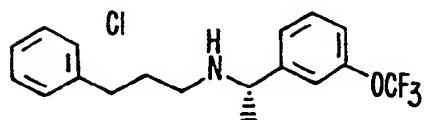
NPS #



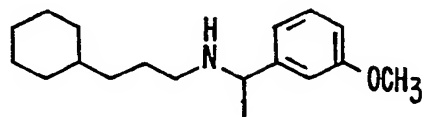
689



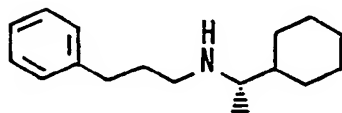
691



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FIG. 36s.

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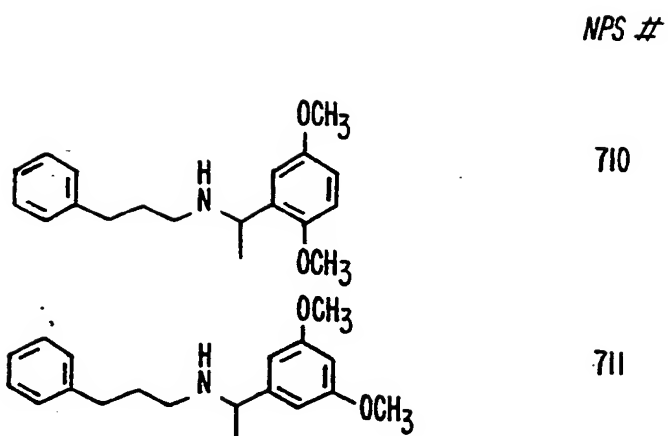


FIG. 36t.

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FIG. 37b.

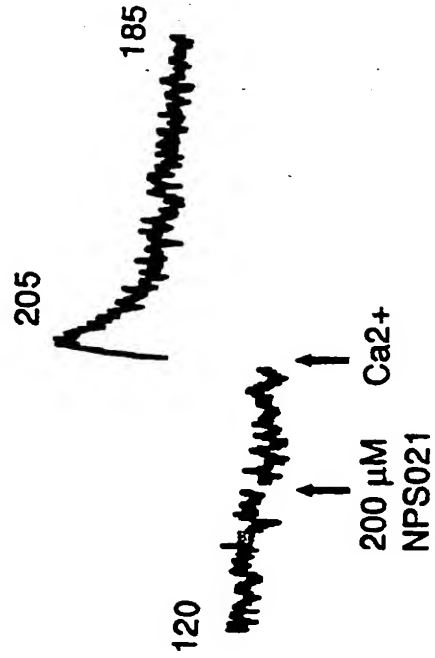
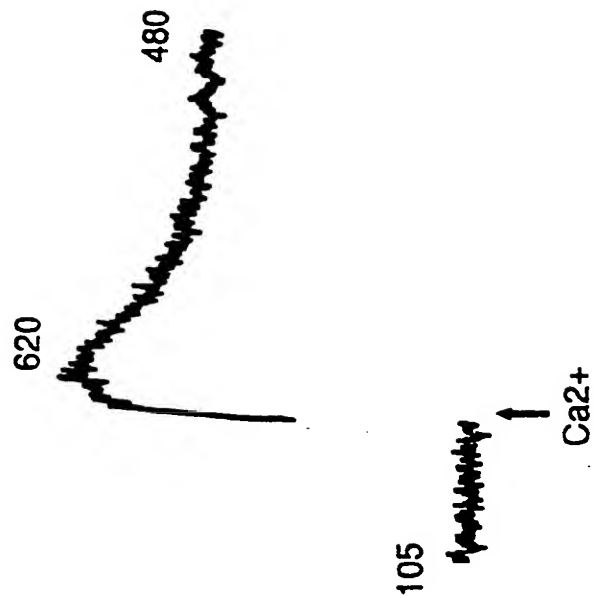


FIG. 37a.

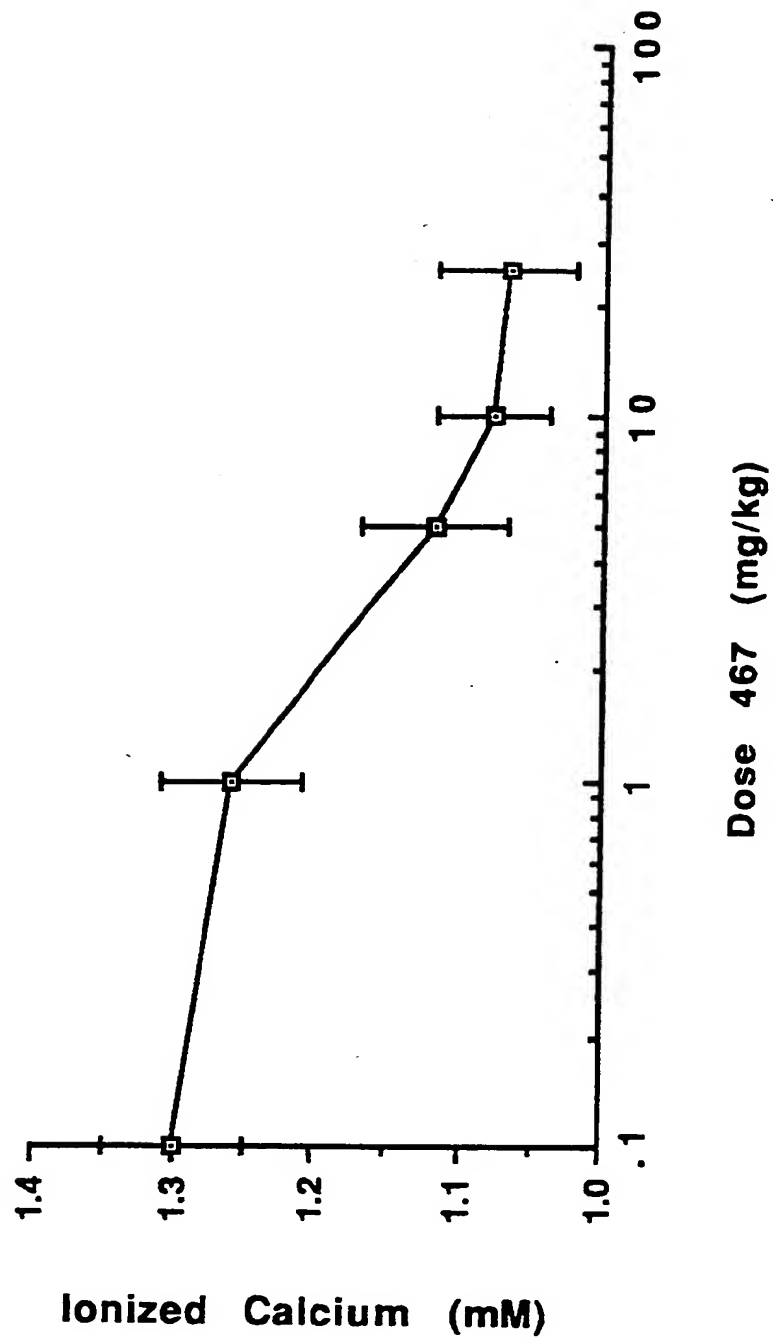


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FIG. 38

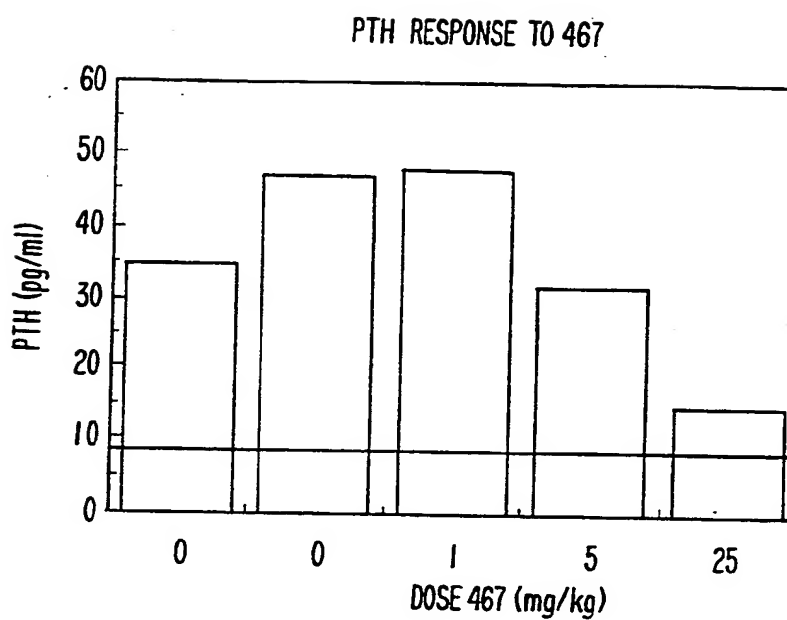
Ionized Calcium Response to NPS 467



Ionized Calcium (mM)

Dose 467 (mg/kg)

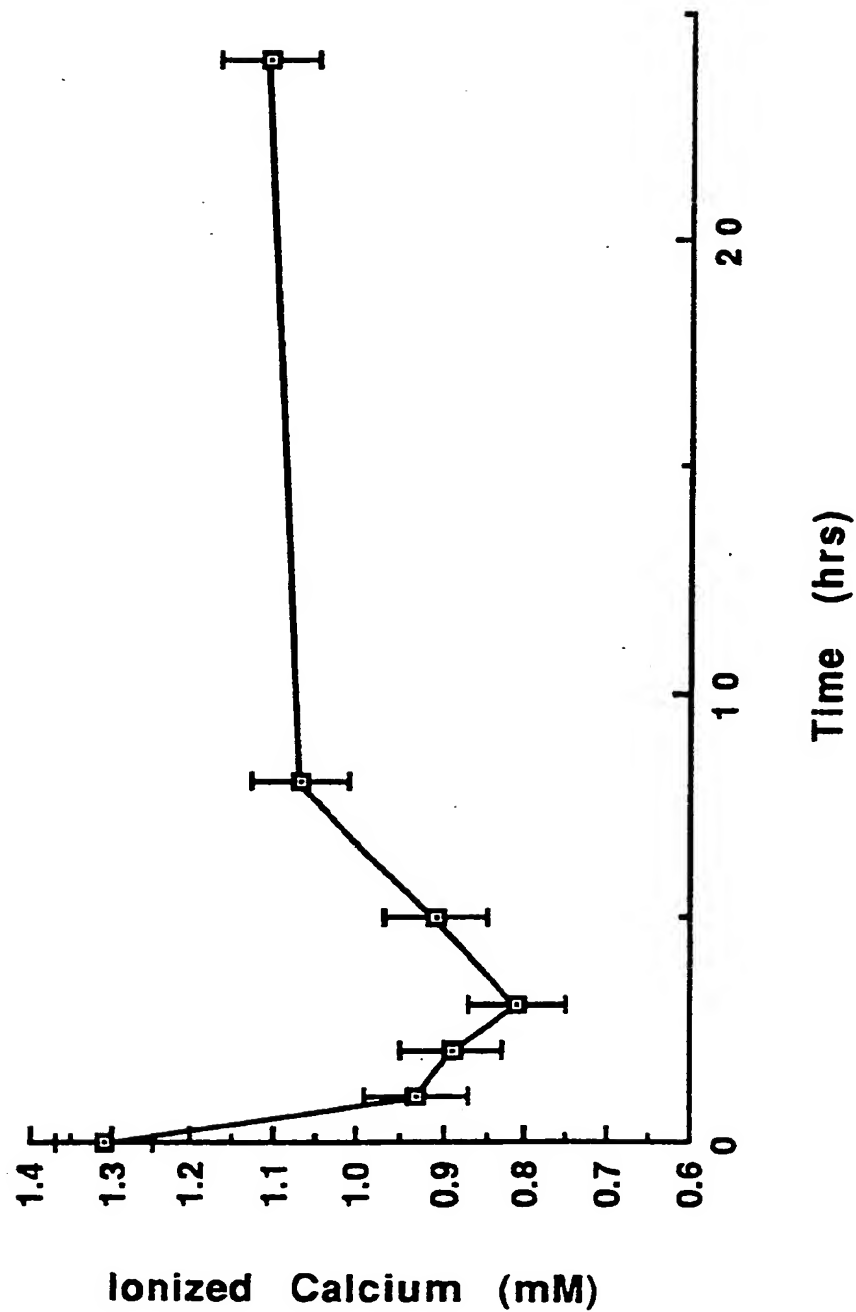
SUBSTITUTE SHEET

*FIG. 39.***SUBSTITUTE SHEET**

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FIG. 40.

Ionized Calcium Response to 25 mg/kg 467

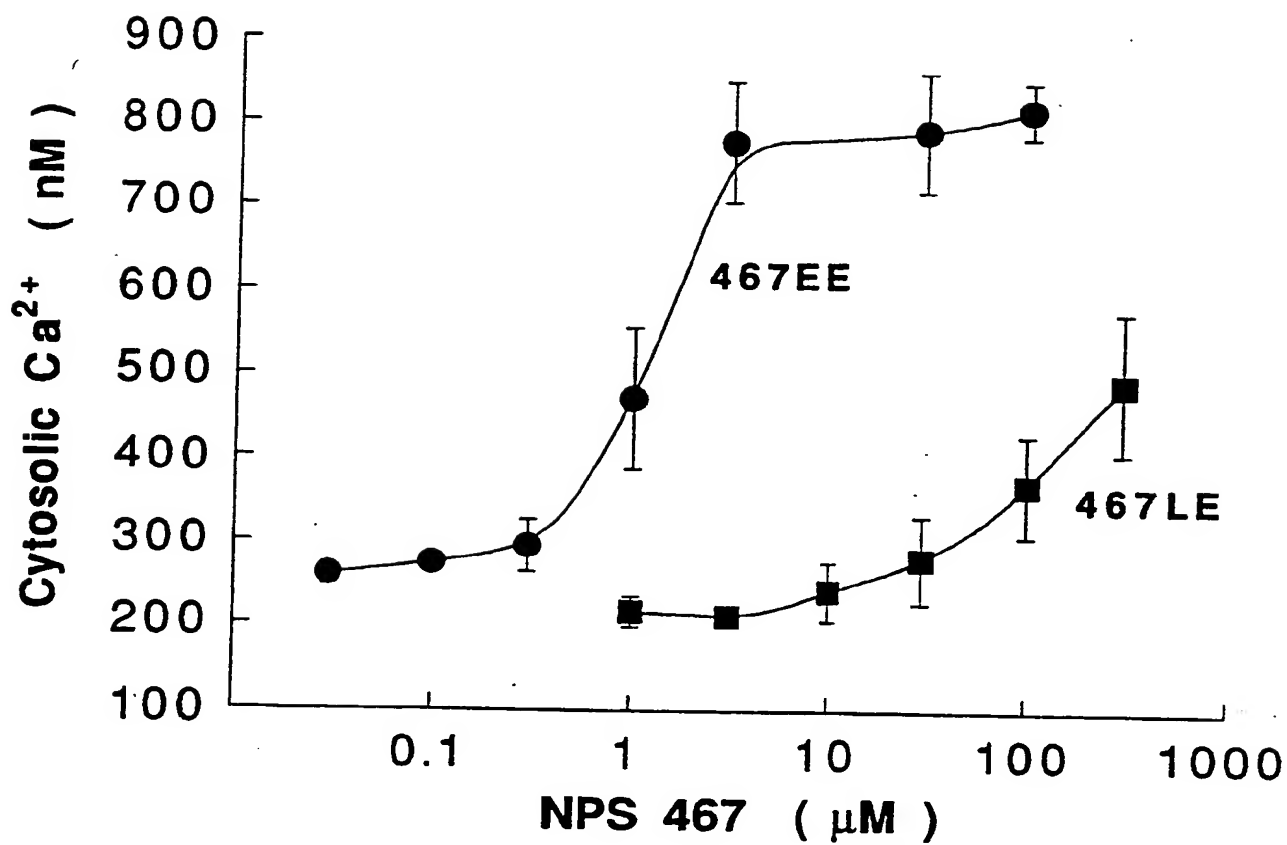


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FIG. 41.



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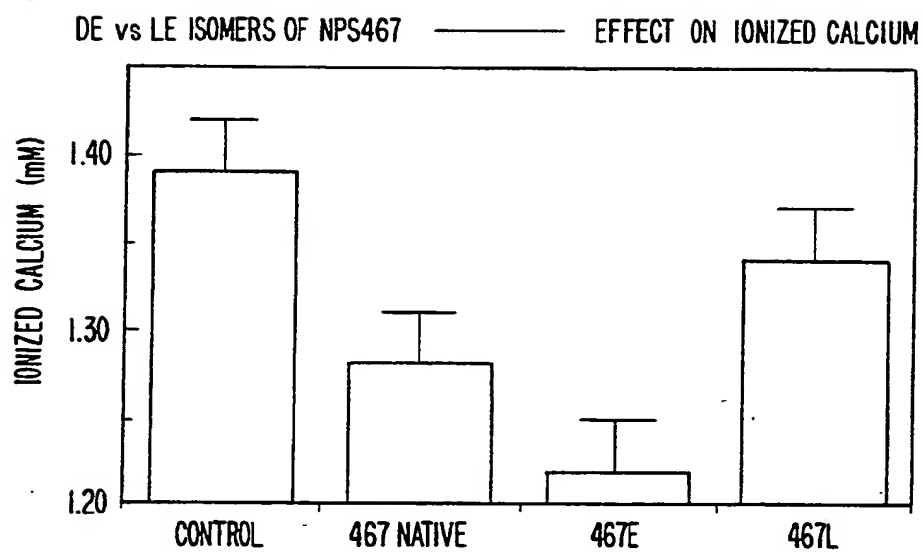


FIG. 42.

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FIG. 43a.

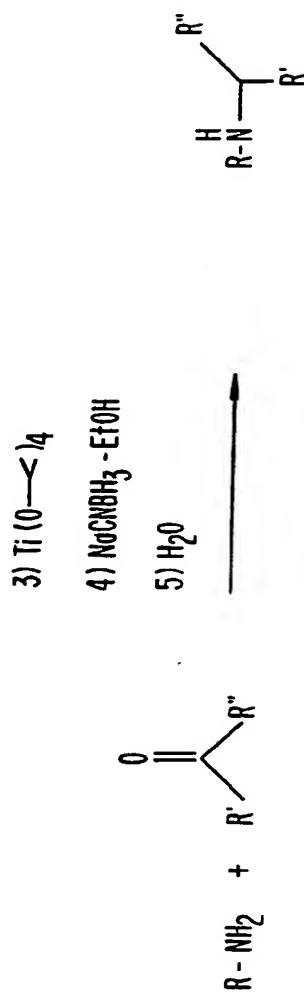
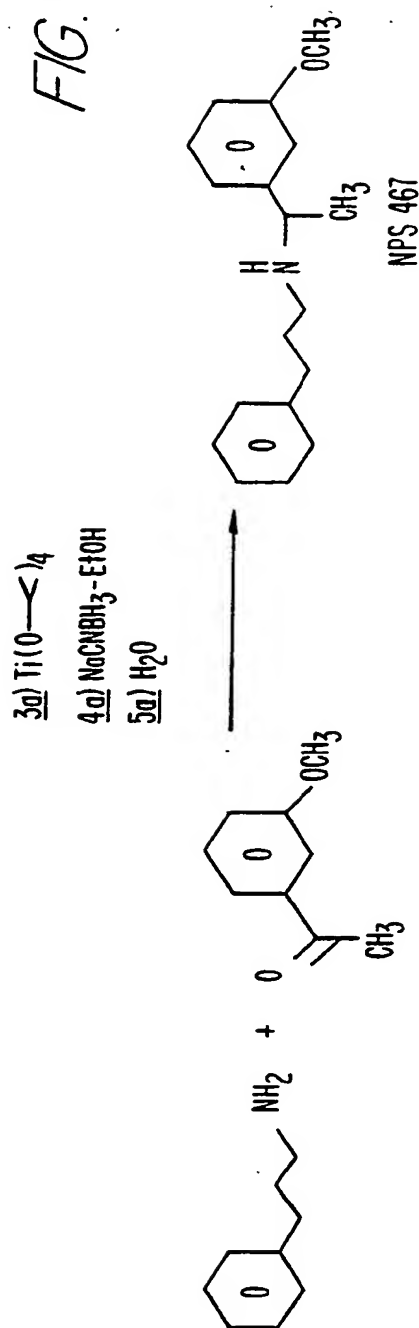
621

FIG. 43b.

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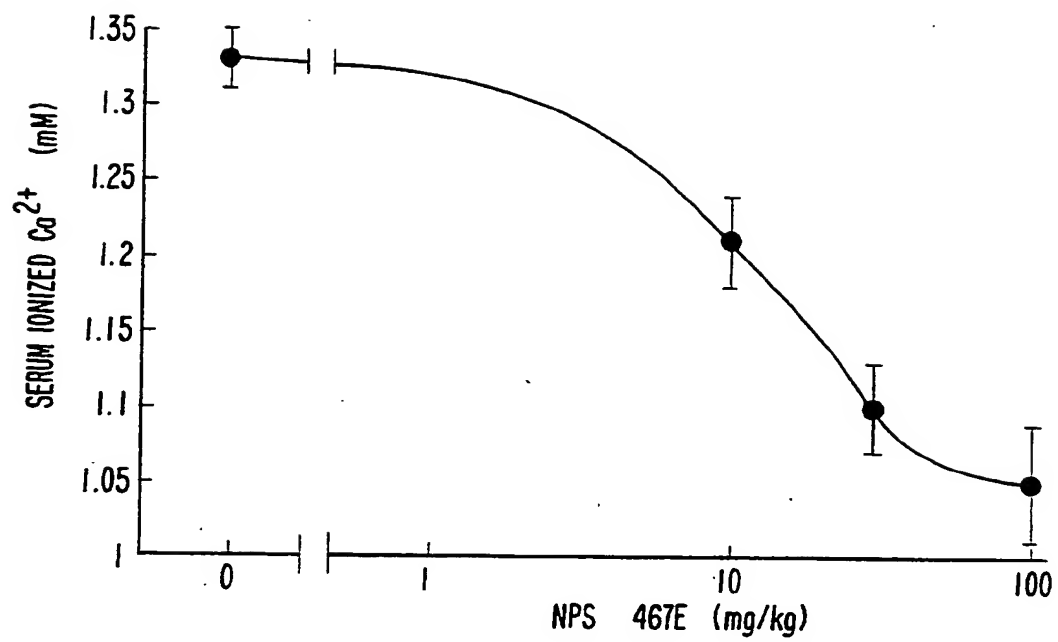


FIG. 44.

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## INTERNATIONAL SEARCH REPORT

International application No.  
PCT/US92/07175

## A. CLASSIFICATION OF SUBJECT MATTER

IPC(5) : Please See Extra Sheet.

US CL : 436/501, 503, 69.1; 536/27; 564/511, 305; 530/324, 350; 564/374

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 436/501, 503, 69.1; 536/27; 564/511, 305; 530/324, 350; 564/374

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

CAS ONLINE, APS,  
search terms: structure search, calcium agonists and antagonists

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
<u>X</u> Y	US, A, 4,988,730 (Korbonits et al) 29 January 1991, see entire document, and Table 2.	1-23, 25-27, 29-31, 36-61, 63-65, <u>67-69, 74-82, 89, 98, 99</u> 32-35, 70-73
Y	CIRCULATION RESEARCH, Vol. 64, No. 3, Issued March 1989, Koenig et al, "Polyamines Mediate Androgenic Stimulation of Calcium Fluxes and Membrane transport in Rat Heart Myocytes", pages 415-426, see the abstract and page 415, column 2.	23, 24
<u>X</u> Y	WO, A, 89/09834 (Ellis et al) 19 October 1989, see pages 6-8, 9, 10-11, 27-28.	<u>83, 84, 90, 92, 93, 97</u> 85-88, 94-96
<u>X</u> Y	QUANTITATIVE STRUCTURE-ACTIVITY RELATIONSHIPS, Vol. 8, Issued 1989, Holtje et al, "Conformational Analysis on Calcium Channel Active Diphenylalkylamines, Diphenylbutylpiperidines, Phenylalkylamines, and Perhexiline" pages 259-265, see Figure 1.	<u>99</u> 1-23, 25-35

☒ Further documents are listed in the continuation of Box C. ☐ See patent family annex.

* Special categories of cited documents:	*T	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
*A		document defining the general state of the art which is not considered to be part of particular relevance
*E	*X	document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
*L		document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
*O	*Y	document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
*P	*Z	document published prior to the international filing date but later than the priority date claimed
		document member of the same patent family

Date of the actual completion of the international search 02 December 1992	Date of mailing of the international search report 09 DEC 1992
Name and mailing address of the ISA/ Commissioner of Patents and Trademarks Box PCT Washington, D.C. 20231 Facsimile No. NOT APPLICABLE	Authorized officer LORA M. GREEN Telephone No. (703) 308-0196

Form PCT/ISA/210 (second sheet)(July 1992)\*

## INTERNATIONAL SEARCH REPORT

International application No.  
PCT/US92/07175

## C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	ANTICANCER RESEARCH, Vol. 11, Issued May-June 1991, Batra et al, "Effects of Diverse Categories of Drugs on Human Colon Tumour Cell Proliferation", pages 1221-1224, see abstract.	36-48, 50, 54, 58, 60, 61, 63-65, 68, 69, 77, 79-82
X	CARDIOVASCULAR DRUGS AND THERAPY, Vol. 2, No. 1, Issued May 1988, Opie, "Calcium Channel Antagonists Part V: Second-Generation Agents", pages 191-203, see entire article.	1-23, 25-27, 30, 31, 36-61, 63-65, 68-69, 74-77, 89
A	MEDICINAL RESEARCH REVIEWS, Vol. 9, No. 1, issued 1989, Triggle et al, "Ca <sup>2+</sup> Channel Ligands; Structure-Function Relationships of the 1,4-Dihydropyridines", pages 123-180, see entire article.	1-99

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# INTERNATIONAL SEARCH REPORT

International application No.  
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## A. CLASSIFICATION OF SUBJECT MATTER:

IPC (5):

G01N 33/566, 33/567; C07C 211/02, 211/16, 211/27.; C07H 21/00; C07K 5/00, 7/00; C12N 15/12; A61K 37/02